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(21) International Application Number: PCT/US95/07628 (22) International Filing Date: 15 June 1995 (15.06.95) (30) Priority Data: <table border="0" style="width: 100%;"> <tr> <td style="width: 40%;">08/260,190</td> <td style="width: 40%;">15 June 1994 (15.06.94)</td> <td style="width: 20%;">US</td> </tr> <tr> <td>08/485,049</td> <td>7 June 1995 (07.06.95)</td> <td>US</td> </tr> <tr> <td>08/486,756</td> <td>7 June 1995 (07.06.95)</td> <td>US</td> </tr> <tr> <td>08/447,504</td> <td>7 June 1995 (07.06.95)</td> <td>US</td> </tr> <tr> <td>08/481,658</td> <td>7 June 1995 (07.06.95)</td> <td>US</td> </tr> <tr> <td>08/485,862</td> <td>7 June 1995 (07.06.95)</td> <td>US</td> </tr> <tr> <td>08/485,863</td> <td>7 June 1995 (07.06.95)</td> <td>US</td> </tr> <tr> <td>08/487,077</td> <td>7 June 1995 (07.06.95)</td> <td>US</td> </tr> </table> (71) Applicant (for AM BF BJ CM MG MR TD TG only): CIBA CORNING DIAGNOSTICS CORP. [US/US]; 63 North Street, Medfield, MA 02052 (US). (71) Applicant (for all designated States except US): INSTITUTE OF VIROLOGY [SK/SK]; Slovak Academy of Sciences, Dubravska Cesta 9, 84 246 Bratislava (SK).		08/260,190	15 June 1994 (15.06.94)	US	08/485,049	7 June 1995 (07.06.95)	US	08/486,756	7 June 1995 (07.06.95)	US	08/447,504	7 June 1995 (07.06.95)	US	08/481,658	7 June 1995 (07.06.95)	US	08/485,862	7 June 1995 (07.06.95)	US	08/485,863	7 June 1995 (07.06.95)	US	08/487,077	7 June 1995 (07.06.95)	US	(72) Inventors; and (75) Inventors/Applicants (for US only): ZAVADA, Jan [CZ/CZ]; Na pekne vyhlidce 1, 160 00 Prague 6 (CZ). PASTOREKOVA, Silvia [SK/SK]; I. Bukovcna 18, 841 07 Bratislava (SK). PASTOREK, Jaromir [SK/SK]; I. Bukovcna 18, 841 07 Bratislava (SK). (74) Agent: LAUDER, Leona, L.; 6 Mariposa Court, Tiburon, CA 94920-2017 (US). (81) Designated States: AM, AT, AU, BB, BG, BR, BY, CA, CH, CN, CZ, DE, DK, EE, ES, FI, GB, GE, HU, IS, JP, KE, KG, KP, KR, KZ, LK, LR, LT, LU, LV, MD, MG, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, TJ, TM, TT, UA, UG, US, UZ, VN, European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG), ARIPO patent (KE, MW, SD, SZ, UG). Published <i>Without international search report and to be republished upon receipt of that report.</i>
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(54) Title: MN GENE AND PROTEIN (57) Abstract <p>A complete genomic sequence including a full-length cDNA sequence for the MN gene, a putative oncogene, is disclosed, as well as proteins/polypeptides encoded thereby. Recombinant nucleic acid molecules for expressing MN proteins/polypeptides and recombinant proteins are also provided. Expression of the MN gene is disclosed as being associated with tumorigenicity, and the invention concerns methods and compositions for detecting and/or quantitating MN antigen and/or MN-specific antibodies in vertebrate samples that are diagnostic/prognostic for neoplastic and preneoplastic disease. Test kits embodying the immunoassays of this invention are provided. MN-specific antibodies are disclosed that can be used diagnostically/prognostically, therapeutically, for imaging, and/or for affinity purification of MN proteins/polypeptides. Also provided are nucleic acid probes for the MN gene as well as test kits comprising said probes. The invention also concerns vaccines comprising MN proteins/polypeptides which are effective to immunize a vertebrate against neoplastic diseases associated with the expression of MN proteins. The invention still further concerns antisense nucleic acid sequences that can be used to inhibit MN gene expression, and polymerase chain reaction (PCR) assays to detect genetic rearrangements in MN genes.</p>																										

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MN GENE AND PROTEIN
FIELD OF THE INVENTION

The present invention is in the general area of medical genetics and in the fields of biochemical engineering and immunochemistry. More specifically, it relates to the identification of a new gene--the MN gene--a cellular gene
5 coding for the MN protein. The inventors hereof found MN proteins to be associated with tumorigenicity. Evidence indicates that the MN protein appears to represent a potentially novel type of oncoprotein. Identification of MN antigen as well as antibodies specific therefor in patient
10 samples provides the basis for diagnostic/prognostic assays for cancer.

BACKGROUND OF THE INVENTION

A novel quasi-viral agent having rather unusual properties was detected by its capacity to complement mutants
15 of vesicular stomatitis virus (VSV) with heat-labile surface G protein in HeLa cells (cell line derived from human cervical adenocarcinoma), which had been cocultivated with human breast carcinoma cells. [Zavada et al., Nature New Biol., 240: 124 (1972); Zavada et al., J. Gen. Virol., 24: 327 (1974);
20 Zavada, J., Arch. Virol., 50: 1 (1976); Zavada, J., J. Gen. Virol., 63: 15-24 (1982); Zavada and Zavadova, Arch. Virol., 118: 189 (1991).] The quasi viral agent was called MaTu as it was presumably derived from a human mammary tumor.

There was significant medical interest in studying
25 and characterizing MaTu as it appeared to be an entirely new type of molecular parasite of living cells, and possibly originated from a human tumor. Zavada et al., International Publication Number WO 93/18152 (published 1 September 1993), describes the elucidation of the biological and molecular
30 nature of MaTu which resulted in the discovery of the MN gene and protein. MaTu was found by the inventors to be a two-

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component system, having an exogenous transmissible component, MX, and an endogenous cellular component, MN. The MN component was found to be a cellular gene, showing only very little homology with known DNA sequences. The MN gene was
5 found to be present in the chromosomal DNA of all vertebrates tested, and its expression was found to be strongly correlated with tumorigenicity.

The exogenous MaTu-MX transmissible agent was identified as lymphocytic choriomeningitis virus (LCMV) which
10 persistently infects HeLa cells. The inventors discovered that the MN expression in HeLa cells is positively regulated by cell density, and also its expression level is increased by persistent infection with LCMV.

Research results provided herein show that cells
15 transfected with MN cDNA undergo changes indicative of malignant transformation. Further research findings indicate that the disruption of cell cycle control is one of the mechanisms by which MN may contribute to the complex process of tumor development.

20 Described herein is the cloning and sequencing of the MN gene and the recombinant production of MN proteins. The full-length MN cDNA sequence [SEQ. ID. NO.: 1], the amino acid sequence deduced therefrom [SEQ. ID. NO.: 2], a full-length genomic sequence for MN [SEQ. ID. NO.: 5] including a
25 proposed promoter sequence [SEQ. ID. NO.: 27] are provided. Eleven exons [SEQ. ID. NOS. 28-38] and ten introns [SEQ. ID. NOS.: 39-48] are comprised by the MN gene. Also a 1.4 kilobase region [SEQ. ID. NO. 49] within the middle of the MN genomic sequence is described herein, which has the character
30 of a typical CpG-rich island, and which contains multiple putative binding sites for transcription factors AP2 and Sp1.

Also described are antibodies prepared against proteins/polypeptides. MN proteins/ polypeptides can be used in serological assays according to this invention to detect
35 MN-specific antibodies. Further, MN proteins/polypeptides and/or antibodies reactive with MN antigen can be used in immunoassays according to this invention to detect and/or

quantitate MN antigen. Such assays may be diagnostic and/or prognostic for neoplastic/pre-neoplastic disease.

SUMMARY OF THE INVENTION

This invention is directed to the MN gene, fragments thereof and the related cDNA which are useful, for example, as follows: 1) to produce MN proteins/ polypeptides by biochemical engineering; 2) to prepare nucleic acid probes to test for the presence of the MN gene in cells of a subject; 3) to prepare appropriate polymerase chain reaction (PCR) primers for use, for example, in PCR-based assays or to produce nucleic acid probes; 4) to identify MN proteins and polypeptides as well as homologs or near homologs thereto; 5) to identify various mRNAs transcribed from MN genes in various tissues and cell lines, preferably human; and 6) to identify mutations in MN genes. The invention further concerns purified and isolated DNA molecules comprising the MN gene or fragments thereof, or the related cDNA or fragments thereof.

Thus, this invention in one aspect concerns isolated nucleic acid sequences that encode MN proteins or polypeptides wherein the nucleotide sequences for said nucleic acids are selected from the group consisting of:

- (a) SEQ. ID. NO.: 1;
- (b) nucleotide sequences that hybridize under stringent conditions to SEQ. ID. NO.: 1 or to its complement;
- (c) nucleotide sequences that differ from SEQ. ID. NO.: 1 or from the nucleotide sequences of (b) in codon sequence because of the degeneracy of the genetic code. Further, such nucleic acid sequences are selected from nucleotide sequences that but for the degeneracy of the genetic code would hybridize to SEQ. ID. NO.: 1 or to its complement under stringent hybridization conditions.

Further, such isolated nucleic acids that encode MN proteins or polypeptides can also include the MN nucleic acids of the genomic sequence shown in Figure 3a-d, that is, SEQ. ID. NO.: 5, as well as sequences that hybridize to it or its complement under stringent conditions, or would hybridize to SEQ. ID. NO.: 5 or to its complement under such conditions,

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but for the degeneracy of the genetic code. Degenerate variants of SEQ. ID. NOS.: 1 and 5 are within the scope of the invention.

Further, this invention concerns nucleic acid probes which are fragments of the isolated nucleic acids that encode MN proteins or polypeptides as described above. Preferably said nucleic acid probes are comprised of at least 29 nucleotides, more preferably of at least 50 nucleotides, still more preferably at least 100 nucleotides, and even more preferably at least 150 nucleotides.

Still further, this invention is directed to isolated nucleic acids containing at least twenty-seven nucleotides selected from the group consisting of:

(a) SEQ. ID. NOS.: 1, 5 and 27-49 and that are complementary to SEQ. ID. NOS.: 1, 5 and 27-49;

(b) nucleotide sequences that hybridize under standard stringent hybridization conditions to one or more of the following nucleotide sequences: SEQ. ID. NOS.: 1, 5, and 27-49 and the respective complements of SEQ. ID. NOS.: 1, 5 and 27-49; and

(c) nucleotide sequences that differ from the nucleotide sequences of (a) and (b) in codon sequence because of the degeneracy of the genetic code. The invention also concerns nucleic acids that but for the degeneracy of the genetic code would hybridize to the nucleic acids of (a) and (b) under standard stringent hybridization conditions. Further this invention concerns nucleic acids of (b) and (c) that hybridize partially or wholly to the non-coding regions of SEQ. ID. NO.: 5 or its complement as, for example, sequences that function as nucleic acid probes to identify MN nucleic acid sequences. Conventional technology can be used to determine whether the nucleic acids of (b) and (c) or of fragments of SEQ. ID. NO.: 5 are useful to identify MN nucleic acid sequences, for example, as outlined in Benton and Davis, Science, 196: 180 (1977) and Fuscoe et al. Genomics, 5: 100 (1989). In general, such nucleic acids are preferably at least 29 nucleotides, most preferably at least 50 nucleotides and still more preferably at least 100

nucleotides. An exemplary and preferred nucleic acid probe is SEQ. ID. NO.: 55 (a 470 bp probe useful in RNase portection assays).

Test kits of this invention can comprise the nucleic acid probes of the invention which are useful diagnostically/prognostically for neoplastic and/or pre-neoplastic disease. Preferred test kits comprise means for detecting or measuring the hybridization of said probes to the MN gene or to the mRNA product of the MN gene, such as a visualizing means.

Fragments of the isolated nucleic acids of the invention, can also be used as PCR primers to amplify segments of MN genes, and may be useful in identifying mutations in MN genes. Typically, said PCR primers are oligonucleotides, preferably at least 16 nucleotides, but they may be considerably longer. Exemplary primers may be from about 16 nucleotides to about 50 nucleotides, preferably from about 19 nucleotides to about 45 nucleotides.

Further, the invention concerns the use of such PCR primers in methods to detect mutations in an isolated MN gene and/or fragment(s) thereof. For example, such methods can comprise amplifying one or more fragment(s) of an MN gene by PCR, and determining whether any of said one or more fragments contain mutations, by, for example, comparing the size of the amplified fragments to those of similarly amplified corresponding fragments of MN genes known to be normal, by using a PCR-single-strand conformation polymorphism assay or a denaturing gradient gel electrophoretic assay.

This invention also concerns nucleic acids which encode MN proteins or polypeptides that are specifically bound by monoclonal antibodies designated M75 that are produced by the hybridoma VU-M75 deposited at the American Type Culture Collection (ATCC) at 12301 Parklawn Drive in Rockville, Maryland 20852 (USA) under ATCC No. HB 11128, and/or by monoclonal antibodies designated MN12 produced by the hybridoma MN 12.2.2 deposited at the ATCC under ATCC No. HB 11647.

This invention further concerns isolated nucleic acids containing at least sixteen nucleotides, preferably at least twenty-nine nucleotides, more preferably at least fifty nucleotides, wherein said nucleic acid is selected from the group consisting of:

(a) the MN nucleic acids contained in plasmids A4a, XE1 and XE3 which were deposited at the American Type Culture Collection (ATCC) in Rockville, Maryland in the United States of America under the respective ATCC Nos. 97199, 97200, and 97198;

(b) nucleic acids that hybridize under stringent conditions to the MN nucleic acids of (a); and

(c) nucleic acids that differ from the nucleic acids of (a) or (b) in codon sequence due to the degeneracy of the genetic code. Such isolated nucleic acids, for example, can be polymerase chain reaction (PCR) primers.

The invention further concerns isolated nucleic acids that code for an MN protein, MN fusion protein or MN polypeptide that is operatively linked to an expression control sequence within a vector; unicellular hosts, prokaryotic or eukaryotic, that are transformed or transfected therewith; and methods of recombinantly producing MN proteins, MN fusion proteins and MN polypeptides comprising transforming or transfecting unicellular hosts with said nucleic acid operatively linked to an expression control sequence, culturing said transformed or transfected unicellular hosts so that said MN proteins, fusion proteins or polypeptides are expressed, and extracting and isolating said MN protein fusion protein or polypeptide.

Recombinant nucleic acids that encode MN fusion proteins are claimed as consisting essentially of an MN protein or MN polypeptide and a non-MN protein or polypeptide wherein the nucleotide sequence for the portion of the nucleic acid encoding the MN protein or polypeptide is selected from the group consisting of:

(a) SEQ. ID. NO.: 1;

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(b) nucleotide sequences that hybridize under stringent conditions to SEQ. ID. NO.: 1 or to its complement; and

(c) degenerate variants of SEQ. ID. NO.: 1, and of
5 the nucleotide sequences of (b);

wherein the nucleic acid encoding said MN protein or polypeptide contains at least twenty-nine nucleotides.

Said non-MN protein or polypeptide may preferably be nonimmunogenic to humans and not typically reactive to
10 antibodies in human body fluids. Examples of such a DNA sequence is the alpha-peptide coding region of beta-galactosidase and a sequence coding for glutathione S-transferase or a fragment thereof. However, in some instances, a non-MN protein or polypeptide that is
15 serologically active, immunogenic and/or antigenic may be preferred as a fusion partner to a MN antigen. Further, claimed herein are such recombinant fusion proteins/polypeptides which are substantially pure and non-naturally occurring. Exemplary fusion proteins of this invention are
20 GEX-3X-MN, MN-Fc and MN-PA, described infra.

In HeLa and in tumorigenic HeLa x fibroblast hybrid (H/F-T) cells, MN protein is manifested as a "twin" protein p54/58N; it is glycosylated and forms disulfide-linked
25 oligomers. As determined by electrophoresis upon reducing gels, MN proteins have molecular weights in the range of from about 40 kd to about 70 kd, preferably from about 45 kd to about 65 kd, more preferably from about 48 kd to about 58 kd. Upon non-reducing gels, MN proteins in the form of oligomers
30 have molecular weights in the range of from about 145 kd to about 160 kd, preferably from about 150 to about 155 kd, still more preferably from about 152 to about 154 kd. A predicted amino acid sequence for a preferred MN protein of this invention is shown in Figure 1 [SEQ. ID. NO. 2].

Other particular MN proteins or polypeptides are
35 exemplified by the putative MN signal peptide shown as the first thirty-seven amino acids in Figure 1 [SEQ. ID. NO.: 6], preferred MN antigen epitopes [SEQ. ID. NOS.: 10-16], and domains of the MN protein represented in Figure 1 as amino

acids 38-135 [SEQ. IS. NO.: 50], 136-391 [SEQ. ID. NO.: 51],
414-433 [SEQ. ID. NO.: 52], and 434-459 [SEQ. ID. NO.: 53].

The discovery of the MN gene and protein and thus,
of substantially complementary MN genes and proteins encoded
5 thereby, led to the finding that the expression of MN proteins
was associated with tumorigenicity. That finding resulted in
the creation of methods that are diagnostic/ prognostic for
cancer and precancerous conditions. Methods and compositions
are provided for identifying the onset and presence of
10 neoplastic disease by detecting and/or quantitating MN antigen
in patient samples, including tissue sections and smears, cell
and tissue extracts from vertebrates, preferably mammals and
more preferably humans. Such MN antigen may also be found in
body fluids.

15 MN proteins and genes are of use in research
concerning the molecular mechanisms of oncogenesis, in cancer
diagnostics/prognostics, and may be of use in cancer
immunotherapy. The present invention is useful for detecting
a wide variety of neoplastic and/or pre-neoplastic diseases.
20 Exemplary neoplastic diseases include carcinomas, such as
mammary, bladder, ovarian, uterine, cervical, endometrial,
squamous cell and adenosquamous carcinomas; and head and neck
cancers; mesodermal tumors, such as neuroblastomas and
retinoblastomas; sarcomas, such as osteosarcomas and Ewing's
25 sarcoma; and melanomas. Of particular interest are head and
neck cancers, gynecologic cancers including ovarian, cervical,
vaginal, endometrial and vulval cancers; gastrointestinal
cancer, such as, stomach, colon and esophageal cancers;
urinary tract cancer, such as, bladder and kidney cancers;
30 skin cancer; liver cancer; prostate cancer; lung cancer; and
breast cancer. Of still further particular interest are
gynecologic cancers; breast cancer; urinary tract cancers,
especially bladder cancer; lung cancer; and liver cancer.
Even further of particular interest are gynecologic cancers
35 and breast cancer. Gynecologic cancers of particular interest
are carcinomas of the uterine cervix, endometrium and ovaries;
more particularly such gynecologic cancers include cervical
squamous cell carcinomas, adenosquamous carcinomas,

adenocarcinomas as well as gynecologic precancerous conditions, such as metaplastic cervical tissues and condylomas.

The invention further relates to the biochemical engineering of the MN gene, fragments thereof or related cDNA. For example, said gene or a fragment thereof or related cDNA can be inserted into a suitable expression vector, wherein it is operatively linked to an expression control sequence; host cells, preferably unicellular, can be transformed or transfected with such an expression vector; and an MN protein/polypeptide, preferably an MN protein, is expressed therein. Such a recombinant protein or polypeptide can be glycosylated or nonglycosylated, preferably glycosylated, and can be purified to substantial purity. The invention further concerns MN proteins/polypeptides which are synthetically or otherwise biologically prepared.

Said MN proteins/polypeptides can be used in assays to detect MN antigen in patient samples and in serological assays to test for MN-specific antibodies. MN proteins/polypeptides of this invention are serologically active, immunogenic and/or antigenic. They can further be used as immunogens to produce MN-specific antibodies, polyclonal and/or monoclonal, as well as an immune T-cell response.

The invention further is directed to MN-specific antibodies, which can be used diagnostically/prognostically and may be used therapeutically. Preferred according to this invention are MN-specific antibodies reactive with the epitopes represented respectively by the amino acid sequences of the MN protein shown in Figure 1 as follows: from AA 62 to AA 67 [SEQ. ID. NO.: 10]; from AA 55 to AA 60 [SEQ. ID. NO.: 11]; from AA 127 to AA 147 [SEQ. ID. NO.: 12]; from AA 36 to AA 51 [SEQ. ID. NO.: 13]; from AA 68 to AA 91 [SEQ. ID. NO.: 14]; from AA 279 to AA 291 [SEQ. ID. NO.: 15]; and from AA 435 to AA 450 [SEQ. ID. NO.: 16]. More preferred are antibodies reactive with epitopes represented by SEQ. ID. NOS.: 10, 11 and 12. Still more preferred are antibodies reactive with the epitopes represented by SEQ. ID NOS: 10 and

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11, as for example, respectively Mabs M75 and MN12. Most preferred are monoclonal antibodies reactive with the epitope represented by SEQ. ID. NO.: 10.

Also preferred according to this invention are antibodies prepared against recombinantly produced MN proteins as, for example, GEX-3X-MN, MN 20-19, MN-Fc and MN-PA. Also preferred are MN-specific antibodies prepared against glycosylated MN proteins, such as, MN 20-19 expressed in baculovirus infected Sf9 cells.

10 A hybridoma that produces a representative MN-specific antibody, the monoclonal antibody M75 (Mab M75), was deposited at the under ATCC Number HB 11128 as indicated above. The M75 antibody was used to discover and identify the MN protein and can be used to identify readily MN antigen in
15 Western blots, in radioimmunoassays and immunohistochemically, for example, in tissue samples that are fresh, frozen, or formalin-, alcohol-, acetone- or otherwise fixed and/or paraffin-embedded and deparaffinized. Another representative MN-specific antibody, Mab MN12, is secreted by the hybridoma
20 MN 12.2.2, which was deposited at the ATCC under the designation HB 11647.

MN-specific antibodies can be used, for example, in laboratory diagnostics, using immunofluorescence microscopy or immunohistochemical staining; as a component in immunoassays
25 for detecting and/or quantitating MN antigen in, for example, clinical samples; as probes for immunoblotting to detect MN antigen; in immunoelectron microscopy with colloid gold beads for localization of MN proteins and/or polypeptides in cells; and in genetic engineering for cloning the MN gene or
30 fragments thereof, or related cDNA. Such MN-specific antibodies can be used as components of diagnostic/prognostic kits, for example, for in vitro use on histological sections; such antibodies can also and used for in vivo diagnostics/prognostics, for example, such antibodies can be labeled
35 appropriately, as with a suitable radioactive isotope, and used in vivo to locate metastases by scintigraphy. Further such antibodies may be used in vivo therapeutically to treat cancer patients with or without toxic and/or cytostatic agents

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attached thereto. Further, such antibodies can be used in vivo to detect the presence of neoplastic and/or pre-neoplastic disease. Still further, such antibodies can be used to affinity purify MN proteins and polypeptides.

5 This invention also concerns methods of treating neoplastic disease and/or pre-neoplastic disease comprising inhibiting the expression of MN genes by administering antisense nucleic acid sequences that are substantially complementary to mRNA transcribed from MN genes. Said
10 antisense nucleic acid sequences are those that hybridize to such mRNA under stringent hybridization conditions. Preferred are antisense nucleic acid sequences that are substantially complementary to sequences at the 5' end of the MN cDNA sequence shown in Figure 1. Preferably said antisense nucleic
15 acid sequences are oligonucleotides.

This invention also concerns vaccines comprising an immunogenic amount of one or more substantially pure MN proteins and/or polypeptides dispersed in a physiologically acceptable, nontoxic vehicle, which amount is effective to
20 immunize a vertebrate, preferably a mammal, more preferably a human, against a neoplastic disease associated with the expression of MN proteins. Said proteins can be recombinantly, synthetically or otherwise biologically produced. A particular use of said vaccine would be to
25 prevent recidivism and/or metastasis. For example, it could be administered to a patient who has had an MN-carrying tumor surgically removed, to prevent recurrence of the tumor.

The immunoassays of this invention can be embodied in test kits which comprise MN proteins/polypeptides and/or
30 MN-specific antibodies. Such test kits can be in solid phase formats, but are not limited thereto, and can also be in liquid phase format, and can be based on immunohistochemical assays, ELISAS, particle assays, radiometric or fluorometric assays either unamplified or amplified, using, for example,
35 avidin/biotin technology.

Abbreviations

The following abbreviations are used herein:

-12-

	AA	-	amino acid
	ATCC	-	American Type Culture Collection
	bp	-	base pairs
	BLV	-	bovine leukemia virus
5	BSA	-	bovine serum albumin
	BRL	-	Bethesda Research Laboratories
	CA	-	carbonic anhydrase
	CAT	-	chloramphenicol acetyltransferase
	Ci	-	curie
10	cm	-	centimeter
	CMV	-	cytomegalovirus
	cpm	-	counts per minute
	C-terminus	-	carboxyl-terminus
	°C	-	degrees centigrade
15	DEAE	-	diethylaminoethyl
	DMEM	-	Dulbecco modified Eagle medium
	EDTA	-	ethylenediaminetetraacetate
	EIA	-	enzyme immunoassay
	ELISA	-	enzyme-linked immunosorbent assay
20	F	-	fibroblasts
	FCS	-	fetal calf serum
	FITC	-	fluorescein isothiocyanate
	GEX-3X-MN	-	fusion protein MN glutathione S-transferase
	H	-	HeLa cells
25	HEF	-	human embryo fibroblasts
	HeLa K	-	standard type of HeLa cells
	HeLa S	-	Stanbridge's mutant HeLa D98/AH.2
	H/F-T	-	hybrid HeLa fibroblast cells that are tumorigenic; derived from HeLa D98/AH.2
30	H/F-N	-	hybrid HeLa fibroblast cells that are nontumorigenic; derived from HeLa D98/AH.2
	HRP	-	horseradish peroxidase
	Inr	-	initiator
	IPTG	-	isopropyl-Beta-D-thiogalacto-pyranoside
35	kb	-	kilobase
	kbp	-	kilobase pairs
	kd	-	kilodaltons
	LCMV	-	lymphocytic choriomeningitis virus

	LTR	-	long terminal repeat
	M	-	molar
	mA	-	milliampere
	MAB	-	monoclonal antibody
5	ME	-	mercaptoethanol
	MEM	-	minimal essential medium
	min.	-	minute(s)
	mg	-	milligram
	ml	-	milliliter
10	mM	-	millimolar
	MMC	-	mitomycin C
	MLV	-	murine leukemia virus
	N	-	normal concentration
	NEG	-	negative
15	ng	-	nanogram
	nt	-	nucleotide
	N-terminus	-	amino-terminus
	ODN	-	oligodeoxynucleotide
	ORF	-	open reading frame
20	PA	-	Protein A
	PBS	-	phosphate buffered saline
	PCR	-	polymerase chain reaction
	PEST	-	combination of one-letter abbreviations for proline, glutamic acid, serine, threonine
25	pI	-	isoelectric point
	PMA	-	phorbol 12-myristate 13-acetate
	POS	-	positive
	Py.	-	pyrimidine
	RIA	-	radioimmunoassay
30	RIP	-	radioimmunoprecipitation
	RIPA	-	radioimmunoprecipitation assay
	RNP	-	RNase protection assay
	SDRE	-	serum dose response element
	SDS	-	sodium dodecyl sulfate
35	SDS-PAGE	-	sodium dodecyl sulfate-polyacrylamide gel electrophoresis
	SINE	-	short interspersed repeated sequence
	SSDS	-	synthetic splice donor site

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	SP-RIA	-	solid-phase radioimmunoassay
	SSDS	-	synthetic splice donor site
	SSPE	-	NaCl (0.18 M), sodium phosphate (0.01 M), EDTA (0.001 M)
5	TBE	-	Tris-borate/EDTA electrophoresis buffer
	TCA	-	trichloroacetic acid
	TC media	-	tissue culture media
	TMB	-	tetramethylbenzidine
	Tris	-	tris (hydroxymethyl) aminomethane
10	μ Ci	-	microcurie
	μ g	-	microgram
	μ l	-	microliter
	μ M	-	micromolar
	VSV	-	vesicular stomatitis virus
15	X-MLV	-	xenotropic murine leukemia virus

Cell Lines

	HeLa K	--	standard type of HeLa cells; aneuploid, epithelial-like cell line isolated from a human cervical adenocarcinoma [Gey et al., <u>Cancer Res.</u> , 12: 264 (1952); Jones et al., <u>Obstet. Gynecol.</u> , 38: 945-949 (1971)] obtained from Professor B. Korych, [Institute of Medical Microbiology and Immunology, Charles University; Prague, Czech Republic]
20			
25	HeLa D98/AH.2 (also HeLa S)	--	Mutant HeLa clone that is hypoxanthine guanine phosphoribosyl transferase-deficient (HGPRT ⁻) kindly provided by Eric J. Stanbridge [Department of Microbiology, College of Medicine, University of California, Irvine, CA (USA)] and reported in Stanbridge et al., <u>Science</u> , 215: 252-259 (15 Jan. 1982); parent of hybrid cells H/F-N and H/F-T, also obtained from E.J. Stanbridge.
30			
35	NIH-3T3	--	murine fibroblast cell line reported in Aaronson, <u>Science</u> , 237: 178 (1987).

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XC -- cells derived from a rat rhabdomyosarcoma induced with Rous sarcoma virus-induced rat sarcoma [Svoboda, J., Natl. Cancer Center Institute Monograph No. 17, IN:

5 "International Conference on Avian Tumor Viruses" (J.W. Beard ed.), pp. 277-298 (1964)], kindly provided by Jan Svoboda [Institute of Molecular Genetics, Czechoslovak Academy of Sciences; Prague, Czech Republic]; and

10

CGL1 -- H/F-N hybrid cells (HeLa D98/AH.2 derivative)

CGL2 -- H/F-N hybrid cells (HeLa D98/AH.2 derivative)

CGL3 -- H/F-T hybrid cells (HeLa D98/AH.2 derivative)

CGL4 -- H/F-T hybrid cells (HeLa D98/Ah.2 derivative)

15 Nucleotide and Amino Acid Sequence Symbols

The following symbols are used to represent nucleotides herein:

	<u>Base Symbol</u>	<u>Meaning</u>
20	A	adenine
	C	cytosine
	G	guanine
	T	thymine
	U	uracil
25	I	inosine
	M	A or C
	R	A or G
	W	A or T/U
	S	C or G
30	Y	C or T/U
	K	G or T/U
	V	A or C or G
	H	A or C or T/U

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D	A or G or T/U
B	C or G or T/U
N/X	A or C or G or T/U

There are twenty main amino acids, each of which is specified by a different arrangement of three adjacent nucleotides (triplet code or codon), and which are linked together in a specific order to form a characteristic protein. A three-letter or one-letter convention is used herein to identify said amino acids, as, for example, in Figure 1 as follows:

	<u>Amino acid name</u>	3 Ltr. <u>Abbrev.</u>	1 Ltr. <u>Abbrev.</u>
	Alanine	Ala	A
	Arginine	Arg	R
15	Asparagine	Asn	N
	Aspartic Acid	Asp	D
	Cysteine	Cys	C
	Glutamic Acid	Glu	E
	Glutamine	Gln	Q
20	Glycine	Gly	G
	Histidine	His	H
	Isoleucine	Ile	I
	Leucine	Leu	L
	Lysine	Lys	K
25	Methionine	Met	M
	Phenylalanine	Phe	F
	Proline	Pro	P
	Serine	Ser	S
	Threonine	Thr	T
30	Tryptophan	Trp	W
	Tyrosine	Tyr	Y
	Valine	Val	V
	Unknown or other		X

BRIEF DESCRIPTION OF THE FIGURES

Figure 1 provides the nucleotide sequence for a full-length MN cDNA [SEQ. ID. NO.: 1] clone isolated as

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described herein. Figure 1 also sets forth the predicted amino acid sequence [SEQ. ID. NO.: 2] encoded by the cDNA.

Figure 2 compares the results of immunizing baby rats to XC tumor cells with rat serum prepared against the fusion protein MN glutathione S-transferase (GEX-3X-MN) (the IM group) with the results of immunizing baby rats with control rat sera (the C group). Each point on the graph represents the tumor weight of a tumor from one rat. Example 2 details those experiments.

Figure 3a-d provides a 10,898 bp complete genomic sequence of MN [SEQ. ID. NO.: 5]. The base count is as follows: 2654 A; 2739 C; 2645 G; and 2859 T. The 11 exons are shown in capital letters.

Figure 4 is a restriction map of the full-length MN cDNA. The open reading frame is shown as an open box. The thick lines below the restriction map illustrate the sizes and positions of two overlapping cDNA clones. The horizontal arrows indicate the positions of primers R1 [SEQ. ID. NO.: 7] and R2 [SEQ. ID. NO.: 8] used for the 5' end RACE. Relevant restriction sites are BamHI (B), EcoRV (V), EcoRI (E), PstI (Ps), PvuII (Pv).

Figure 5 is a map of the human MN gene. The numbered black boxes represent exons. The box designated LTR denotes a region of homology to HERV-K LTR. The empty boxes are Alu-related sequences.

Figure 6 is a nucleotide sequence for the proposed promoter of the human MN gene [SEQ. ID. No.: 27]. The nucleotides are numbered from the transcription initiation site according to RNase protection assay. Potential regulatory elements are overlined. Transcription start sites are indicated by asterisks (RNase protection) and dots (RACE). The sequence of the 1st exon begins under the asterisks.

Figure 7 provides a schematic of the alignment of MN genomic clones according to their position related to the transcription initiation site. All the genomic fragments except Bd3 were isolated from a lambda FIX III genomic library derived from HeLa cells. Clone Bd3 was derived from a human fetal brain library.

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Figure 8 shows the construction and cloning of a series of 5' deletion mutants of MN's putative promoter region linked to the bacterial CAT gene.

DETAILED DESCRIPTION

5 The MN gene is shown herein to be organized into 11 exons and 10 introns. Described herein is the cloning and sequencing of the MN cDNA and genomic sequences, and the genetic engineering of MN proteins -- such as the GEX-3X-MN, MN-PA, MN-Fc and MN 20-19 proteins. The recombinant MN
10 proteins can be conveniently purified by affinity chromatography.

 MN is manifested in HeLa cells by a twin protein, p54/58N. Immunoblots using a monoclonal antibody reactive with p54/58N (MAb M75) revealed two bands at 54 kd and 58 kd.
15 Those two bands may correspond to one type of protein that differs by glycosylation pattern or by how it is processed. Herein, the phrase "twin protein" indicates p54/58N.

 The expression of MN proteins appears to be diagnostic/prognostic for neoplastic disease. The MN twin
20 protein, p54/58N, was found to be expressed in HeLa cells and in Stanbridge's tumorigenic (H/F-T) hybrid cells [Stanbridge et al., Somatic Cell Genet. 7: 699-712 (1981); and Stanbridge et al., Science, 215: 252-259 (1982)] but not in fibroblasts or in non-tumorigenic (H/F-N) hybrid cells [Stanbridge et al.,
25 id.]. In early studies reported in Zavada et al. WO 93/18152, supra, MN proteins were found in immunoblots prepared from human ovarian, endometrial and uterine cervical carcinomas, and in some benign neoplasias (as mammary papilloma) but not from normal ovarian, endometrial, uterine or placental
30 tissues. Example 1 herein details further research on MN gene expression wherein MN antigen, as detected by immunohistochemical staining, was found to be prevalent in tumor cells of a number of cancers, including cervical, bladder, head and neck, and renal cell carcinomas among
35 others. Further, the immunohistochemical staining experiments of Example 1 show that among normal tissues tested, only normal stomach tissues showed routinely and extensively the

presence of MN antigen. MN antigen is further shown herein to be present sometimes in morphologically normal-appearing areas of tissue specimens exhibiting dysplasia and/or malignancy.

MN Gene--Cloning and Sequencing

5 Figure 1 provides the nucleotide sequence for a full-length MN cDNA clone isolated as described below [SEQ. ID. NO.: 1]. Figure 3a-d provides a complete MN genomic sequence [SEQ. ID. NO.: 5]. Figure 6 shows the nucleotide sequence for a proposed MN promoter [SEQ. ID. NO.: 27].

10 It is understood that because of the degeneracy of the genetic code, that is, that more than one codon will code for one amino acid [for example, the codons TTA, TTG, CTT, CTC, CTA and CTG each code for the amino acid leucine (leu)], that variations of the nucleotide sequences in, for example,
15 SEQ. ID. NOS.: 1 and 5 wherein one codon is substituted for another, would produce a substantially equivalent protein or polypeptide according to this invention. All such variations in the nucleotide sequences of the MN cDNA and complementary nucleic acid sequences are included within the scope of this
20 invention.

 It is further understood that the nucleotide sequences herein described and shown in Figures 1, 3a-d and 6, represent only the precise structures of the cDNA, genomic and promoter nucleotide sequences isolated and described herein.

25 It is expected that slightly modified nucleotide sequences will be found or can be modified by techniques known in the art to code for substantially similar or homologous MN proteins and polypeptides, for example, those having similar epitopes, and such nucleotide sequences and proteins/
30 polypeptides are considered to be equivalents for the purpose of this invention. DNA or RNA having equivalent codons is considered within the scope of the invention, as are synthetic nucleic acid sequences that encode proteins/polypeptides homologous or substantially homologous to MN

35 proteins/polypeptides, as well as those nucleic acid sequences that would hybridize to said exemplary sequences [SEQ. ID. NOS. 1, 5 and 27] under stringent conditions, or that, but for

the degeneracy of the genetic code would hybridize to said cDNA nucleotide sequences under stringent hybridization conditions. Modifications and variations of nucleic acid sequences as indicated herein are considered to result in sequences that are substantially the same as the exemplary MN sequences and fragments thereof.

Partial cDNA clone

In Zavada et al., *id.*, the isolation of a partial MN cDNA clone of 1397 bp in length was described. A lambda gt11 cDNA library of LMCV-infected HeLa cells was prepared and subjected to immunoscreening with Mab M75 in combination with goat anti-mouse antibodies conjugated with alkaline phosphatase. One positive clone was picked and subcloned into the NotI site of pBluescript KS [Stratagen; La Jolla, CA (USA)] thereby creating pBluscript-MN.

Two oppositely oriented nested deletions were made using Erase-a-Base™ kit [Promega; Madison, WI (USA)] and sequenced by dideoxy method with a T7 sequencing kit [Pharmacia; Piscataway, NJ (USA)]. The sequencing showed a partial cDNA clone, the insert being 1397 bp long. The sequence comprises a large 1290 bp open reading frame and 107 bp 3' untranslated region containing a polyadenylation signal (AATAAA). However, the sequence surrounding the first ATG codon in the open reading frame (ORF) did not fit the definition of a translational start site. In addition, as followed from a comparison of the size of the MN clone with that of the corresponding mRNA in a Northern blot, the cDNA was shown to be missing about 100 bp from the 5' end of its sequence.

Full-Length cDNA Clone

Attempts to isolate a full-length clone from the original cDNA library failed. Therefore, the inventors performed a rapid amplification of cDNA ends (RACE) using MN-specific primers, R1 and R2 [SEQ. ID. NOS.: 7 and 8], derived from the 5' region of the original cDNA clone. The RACE product was inserted into pBluescript, and the entire

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population of recombinant plasmids was sequenced with an MN-specific primer ODN1 [SEQ. ID. NO.: 3]. In that way, a reliable sequence at the very 5' end of the MN cDNA as shown in Figure 1 [SEQ. ID. NO.: 1] was obtained.

5 Specifically, RACE was performed using 5' RACE System [GIBCO BRL; Gaithersburg, MD (USA)] as follows. 1 µg of mRNA (the same as above) was used as a template for the first strand cDNA synthesis which was primed by the MN-specific antisense oligonucleotide, R1 (5'-
10 TGGGGTTCTTGAGGATCTCCAGGAG-3') [SEQ. ID. NO.: 7]. The first strand product was precipitated twice in the presence of ammonium acetate and a homopolymeric C tail was attached to its 3' end by TdT. Tailed cDNA was then amplified by PCR using a nested primer, R2 (5'-CTCTAACTTCAGGGAGCCCTCTTCTT-3')
15 [SEQ. ID. NO.: 8] and an anchor primer that anneals to the homopolymeric tail (5'-CUACUACUACUAGGCCACGCGTCGACTAGTACGGGI
IGGGIIGGGIIG-3') [SEQ. ID. NO.: 9]. The amplified product was digested with BamHI and SalI restriction enzymes and cloned into pBluescript II KS plasmid. After transformation,
20 plasmid DNA was purified from the whole population of transformed cells and used as a template for sequencing with the MN-specific primer ODN1 [SEQ. ID. NO.: 3; a 29-mer 5'
CGCCCAGTGGGTCATCTTCCCCAGAAGAG 3'].

Based upon results of the RACE analysis, the full-
25 length MN cDNA sequence was seen to contain a single ORF starting at position 12, with an ATG codon that is in a good context (GCGCATGG) with the rule proposed for translation initiation [Kozak, J. Cell. Biol., 108: 229-241 (1989)].
[See below under Mapping of MN Gene Transcription Initiation
30 Site for fine mapping of the 5' end of the MN gene.] The AT rich 3' untranslated region contains a polyadenylation signal (AATAAA) preceding the end of the cDNA by 10 bp.
Surprisingly, the sequence from the original clone as well as from four additional clones obtained from the same cDNA
35 library did not reveal any poly(A) tail. Moreover, just downstream of the poly(A) signal, an ATTTA motif that is thought to contribute to mRNA instability [Shaw and Kamen, Cell, 46: 659-667 (1986)] was found. That fact raised the

possibility that the poly (A) tail is missing due to the specific degradation of the MN mRNA.

Genomic clones

To study MN regulation, MN genomic clones were
5 isolated. One MN genomic clone (Bd3) was isolated from a human cosmid library prepared from fetal brain using both MN cDNA as a probe and the MN-specific primers derived from the 5' end of the cDNA ODN1 [SEQ. ID. NO.: 3, supra] and ODN2 [SEQ. ID NO.: 4; 19-mer (5' GGAATCCTCCTGCATCCGG 3')].
10 Sequence analysis revealed that that genomic clone covered a region upstream from a MN transcription start site and ending with the BamHI restriction site localized inside the MN cDNA. Other MN genomic clones can be similarly isolated.

In order to identify the complete genomic region of
15 MN, the human genomic library in Lambda FlX II vector (Stratagene) was prepared from HeLa chromosomal DNA and screened by plaque hybridization using MN cDNA as described below. Several independent MN recombinant phages were identified, isolated and characterized by restriction mapping
20 and hybridization analyses. Four overlapping recombinants covering the whole genomic region of MN were selected, digested and subcloned into pBluescript. The subclones were then subjected to bidirectional nested deletions and sequencing. DNA sequences were compiled and analyzed by
25 computer using the DNASIS software package.

The details of isolating genomic clones covering the complete genomic region for MN are provided below. Figure 7 provides a schematic of the alignment of MN genomic clones according to the transcription initiation site. Plasmids
30 containing the A4a clone and the XE1 and XE3 subclones were deposited at the American Type Culture Collection (ATCC) at 12301 Parklawn Drive, Rockville, MD 20852 (USA) on June 6, 1995, respectively under ATCC Deposit Nos. 97199, 97200, and 97198.

Isolation of Genomic DNA Clones

The Sau3AI human HeLa genomic library was prepared in Lambda FIX II vector [Stratagene; La Jolla, CA (USA)] according to manufacturer's protocol. Human fetal brain cosmid library in SuperCos cosmid was from Stratagene. Recombinant phages or bacteria were plated at 1×10^5 plaque forming units on 22x22 cm Nunc plates or 5×10^4 cells on 150 mm Petri dishes, and plaques or colonies were transferred to Hybond N membranes (Amersham). Hybridization was carried out with the full-length MN cDNA labeled with [P^{32}]PdCTP by the Multiprime DNA labeling method (Amersham) at 65°C in 6 x SSC, 0.5% SDS, 10 x Denhardt's and 0.2 mg/1 ml salmon sperm DNA. Filters were washed twice in 2 x SSC, 0.1% SDS at 65°C for 20 min. The dried filters were exposed to X-ray films, and positive clones were picked up. Phages and bacteria were isolated by 3-4 sequential rounds of screening.

Subcloning and DNA Sequencing

Genomic DNA fragments were subcloned into a pBluescript KS and templates for sequencing were generated by serial nested deletions using the Erase-a-Base system. Sequencing was performed by the dideoxynucleotide chain termination method using T7 sequencing kit (Pharmacia). Nucleotide sequence alignments and analyses were carried out using the DNASIS software package (Hitachi Software Engineering).

Exon-Intron Structure of Complete MN Genomic Region

The complete sequence of the overlapping clones contains 10,898 bp (SEQ. ID. NO.: 5). Figure 5 depicts the organization of the human MN gene, showing the location of all 11 exons as well as the 2 upstream and 6 intronic Alu repeat elements. All the exons are small, ranging from 27 to 191 bp, with the exception of the first exon which is 445 bp. The intron sizes range from 89 to 1400 bp.

Table 1 below lists the splice donor and acceptor sequences that conform to consensus splice sequences including

the AG-GT motif [Mount, "A catalogue of splice junction sequences," Nucleic Acids Res. 10: 459-472 (1982)].

TABLE 1
Exon-Intron Structure of the Human MN Gene

	Exon	Size	Genomic Position**	SEQ ID NO	5'splice donor	SEQ ID No
5	1	445	*3507-3951	28	AGAAG gtaagt	67
	2	30	5126-5155	29	TGGAG gtgaga	68
	3	171	5349-5519	30	CAGTC gtgagg	69
10	4	143	5651-5793	31	CCGAG gtgagc	70
	5	93	5883-5975	32	TGGAG gtacca	71
	6	67	7376-7442	33	GGAAG gtcagt	72
	7	158	8777-8934	34	AGCAG gtgggc	73
	8	145	9447-9591	35	GCCAG gtacag	74
15	9	27	9706-9732	36	TGCTG gtgagt	75
	10	82	10350-10431	37	CACAG gtatta	76
	11	191	10562-10752	38	ATAAT end	

	Intron	Size	Genomic Position**	SEQ ID NO	3'splice acceptor	SEQ ID NO
	1	1174	3952-5125	39	atacag GGGAT	77
20	2	193	5156-5348	40	ccccag GCGAC	78
	3	131	5520-5650	41	acgcag TGCAA	79
	4	89	5794-5882	42	tttcag ATCCA	80
	5	1400	5976-7375	43	ccccag GAGGG	81
	6	1334	7443-8776	44	tcacag GCTCA	82
25	7	512	8935-9446	45	ccctag CTCCA	83
	8	114	9592-9705	46	ctccag TCCAG	84
	9	617	9733-10349	47	tcgcag GTGACA	85
	10	130	10432-10561	48	acacag AAGGG	86

** positions are related to nt numbering in whole genomic sequence including the 5' flanking region [Figure 3a-d]

* number corresponds to transcription initiation site determined below by RNase protection assay

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A search for sequences related to MN gene in the EMBL Data Library did not reveal any specific homology except for 6 complete and 2 partial Alu-type repeats with homology to Alu sequences ranging from 69.8% to 91% [Jurka and
5 Milosavljevic, "Reconstruction and analysis of human Alu genes," J. Mol. Evol. 32: 105-121 (1991)]. Below under the Characterization of the 5' Flanking Region, also a 222 bp sequence proximal to the 5' end of the genomic region is shown to be closely homologous to a region of the HERV-K LTR.

10 Mapping of MN Gene Transcription Initiation Site

In the earlier attempt to localize the site of transcription initiation of the MN gene by RACE (above), the obtained a major PCR fragment whose sequence placed the start site 12 bp upstream from the first codon of the ORF. That
15 result was obtained probably due to a preferential amplification of the shortest form of mRNA. Therefore, the inventors used an RNase protection assay (RNP) for fine mapping of the 5' end of the MN gene. The probe was a uniformly labeled 470 nucleotide copy RNA (nt -205 to +265)
20 [SEQ. ID. NO.: 55], which was hybridized to total RNA from MN-expressing HeLa and CGL3 cells and analyzed on a sequencing gel. That analysis has shown that the MN gene transcription initiates at multiple sites, the 5' end of the longest MN transcript being 30 nt longer than that previously
25 characterized by RACE.

RNase Protection Assay

³²P-labeled RNA probes were prepared with an RNA Transcription kit (Stratagene). In vitro transcription reactions were carried out using 1 µg of the linearized
30 plasmid as a template, 50 µCi of [P³²P]rUTP (800 Ci/mmol), 10 U of either T3 or T7 RNA polymerase and other components of the Transcription Kit following instructions of the supplier. For mapping of the 5' end of MN mRNA, the 470 bp NcoI-BamHI fragment (NcoI filled in by Klenow enzyme) of Bd3 clone (nt -
35 205 to +265 related to transcription start) was subcloned to EcoRV-BamHI sites of pBluescript SK+, linearized with HindIII

and labeled with T3 RNA polymerase. For the 3' end mRNA analysis, probe, that was prepared using T7 RNA polymerase on KS-dXE3-16 template (one of the nested deletion clones of MN genomic region XE3 subclone) digested with Sau3AI (which cuts
5 exon 11 at position 10,629), was used. Approximately 3×10^5 cpm of RNA probe were used per one RNase protection assay reaction.

RNase protection assays (RNP) were performed using Lysate RNase Protection Kit (USB/Amersham) according to
10 protocols of the supplier. Briefly, cells were lysed using Lysis Solution at concentration of approximately 10^7 cells/ml, and 45 μ l of the cell homogenate were used in RNA/RNA hybridization reactions with 32 P-labeled RNA probes prepared as described above. Following overnight hybridizations at 42°C,
15 homogenates were treated for 30 min at 37°C with RNase cocktail mix. Protected RNA duplexes were run on polyacrylamide/urea denaturing sequencing gels. Fixed and dried gels were exposed to X-ray film for 24 - 72 hours.

Mapping of MN Gene Transcription Termination Site

20 An RNase protection assay, as described above, was also used to verify also the 3' end of the MN cDNA. That was important with respect to our previous finding that the cDNA contains a poly(A) signal but lacks a poly(A) tail, which could be lost during the proposed degradation of MN mRNA due
25 to the presence of an instability motif in its 3' untranslated region. RNP analysis of MN mRNA with the fragment of the genomic clone XE3 covering the region of interest corroborated our data from MN cDNA sequencing, since the 3' end of the protected fragment corresponded to the last base of MN cDNA
30 (position 10,752 of the genomic sequence). That site also meets the requirement for the presence of a second signal in the genomic sequence that is needed for transcription termination and polyadenylation [McLauchlan et al., Nucleic Acids Res., 13: 1347 (1985)]. Motif TGTGTTAGT (nt 10,759-
35 10,767) corresponds well to both the consensus sequence and the position of that signal within 22 bp downstream from the polyA signal (nt 10,737-10,742).

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Characterization of the 5' Flanking Region

The Bd3 genomic clone isolated from human fetal brain cosmid library was found to cover a region of 3.5 kb upstream from the transcription start site of the MN gene. It contains no significant coding region. Two Alu repeats are situated at positions -2587 to -2296 [SEQ. ID. NO.: 56] and -1138 to -877 [SEQ. ID. NO.: 57] (with respect to the transcription start determined by RNP). The sequence proximal to the 5' end is strongly homologous (91.4% identity) to the U3 region of long terminal repeats of human endogenous retroviruses HERV-K [Ono, M., "Molecular cloning and long terminal repeat sequences of human endogenous retrovirus genes related to types A and B retrovirus genes," J. Virol. 58: 937-944 (1986)]. The LTR-like fragment is 222 bp long with an A-rich tail at its 3' end. Most probably, it represents part of SINE (short interspersed repeated sequence) type nonviral retroposon derived from HERV-K [Ono et al., "A novel human nonviral retroposon derived from an endogenous retrovirus," Nucleic Acids Res. 15: 8725-8373 (1987)]. There are no sequences corresponding to regulatory elements in this fragment, since the 3' part of U3, and the entire R and U5 regions of LTR are absent from the Bd3 genomic clone, and the glucocorticoid responsive element as well as the enhancer core sequences are beyond its 5' border.

However, two keratinocyte-dependent enhancers were identified in the sequence downstream from the LTR-like fragment at positions -3010 and -2814. Those elements are involved in transcriptional regulation of the E6-E7 oncogenes of human papillomaviruses and are thought to account for their tissue specificity [Cripe et al., "Transcriptional regulation of the human papilloma-virus-16 E6-E7 promoter by a keratinocyte-dependent enhancer, and by viral E2 trans-activator and repressor gene products: implications for cervical carcinogenesis," EMBO J. 6: 3745-3753 (1987)].

Nucleotide sequence analysis of the DNA 5' to the transcription start (from nt -507) revealed no recognizable TATA box within the expected distance from the beginning of the first exon (Figure 6). However, the presence of potential

binding sites for transcription factors suggests that this region might contain a promoter for the MN gene. There are several consensus sequences for transcription factors AP1 and AP2 as well as for other regulatory elements, including a p53 binding site [Locker and Buzard, "A dictionary of transcription control sequences," J. DNA Sequencing and Mapping, 1: 3-11 (1990); Imagawa et al., "Transcription factor AP-2 mediates induction by two different signal-transduction pathways: protein kinase C and cAMP," Cell, 51: 251-260 (1987); El Deiry et al., "Human genomic DNA sequences define a consensus binding site for p53," Nat. Genet., 1: 44-49 (1992)]. Although the putative promoter region contains 59.3% C+G, it does not have additional attributes of CpG-rich islands that are typical for TATA-less promoters of housekeeping genes [Bird, "CpG-rich islands and the function of DNA methylation," Nature, 321: 209-213 (1986)]. Another class of genes lacking TATA box utilizes the initiator (Inr) element as a promoter. Many of these genes are not constitutively active, but they are rather regulated during differentiation or development. The Inr has a consensus sequence of PyPyPyCAPyPyPyPyPy [SEQ. ID. NO.: 23] and encompasses the transcription start site [Smale and Baltimore, "The 'initiator' as a transcription control element," Cell, 57: 103-113 (1989)]. There are two such consensus sequences in the MN putative promoter; however, they do not overlap the transcription start (Figure 6).

In the initial experiments, the inventors were unable to show promoter activity in human carcinoma cells HeLa and CGL3 that express MN, using the 3.5 kb Bd3 fragment and series of its deletion mutants (from nt -933 to -30) [SEQ. ID. NO.: 58] fused to chloramphenicol acetyl transferase (CAT) gene in a transient system. This might indicate that either the promoter activity of the region 5' to the MN transcription start is below the sensitivity of the CAT assay, or additional regulatory elements not present in our constructs are required for driving the expression of MN gene.

With respect to this fact, an interesting region was found in the middle of the MN gene. The region is about 1.4

kb in length [nt 4,600-6,000 of the genomic sequence; SEQ. ID. NO.: 49] and spans from the 3' part of the 1st intron to the end of the 5th exon. The region has the character of a typical CpG-rich island, with 62.8% C+G content and 82 CpG: 5 131 GpC dinucleotides. Moreover, there are multiple putative binding sites for transcription factors AP2 and Sp1 [Locker and Buzard, supra; Briggs et al., "Purification and biochemical characterization of the promoter-specific transcription factor Sp-1," Science, 234: 47-52 (1986)] 10 concentrated in the center of this area. Particularly the 3rd intron of 131 bp in length contains three Sp1 and three AP2 consensus sequences. That data indicates the possible involvement of that region in the regulation of MN gene expression. However, functionality of that region, as well as 15 other regulatory elements found in the proposed 5' MN promoter, remains to be determined.

MN Promoter Analysis

To define sequences necessary for MN gene expression, a series of 5' deletion mutants of the putative 20 promoter region were fused to the bacterial chloramphenicol acetyltransferase (CAT) gene. [See Figure 8.] The pMN-CAT deletion constructs were transfected using a DEAE dextran method for transient expression into HeLa and CGL3 cells. Those cells were used since they naturally express MN protein, 25 and thus, should contain all the required transcription factors.

After 48 hours, crude cell lysates were prepared and the activity of the expressed CAT was evaluated according to acetylation of [¹⁴C]chloramphenicol by thin layer 30 chromatography. However, no MN promoter CAT activity was detected in either the HeLa or the CGL3 cells in a transient system. On the other hand, reporter CAT plasmids with viral promoters (e.g. pBLV-LTR + tax transactivator, pRSV CAT and pSV2 CAT), that served as positive controls, gave strong 35 signals on the chromatogram. [pSV2 CAT carries the SV40 origin and expresses CAT from the SV40 early promoter (P_E).

pRSV CAT expresses CAT from the Rous sarcoma virus (RSV) LTR promoter (P_{LTR}).]

No detectable CAT activity was observed in additional experiments using increasing amounts of transfected plasmids (from 2 to 20 g DNA per dish) and prolonged periods of cell incubation after transcription. Increased cell density also did not improve the results (in contrast to the expectations based on density-dependent expression of native MN protein in HeLa cells). Since the inventors had found consensus sequences for transcription factors AP2 and AP1 in the putative MN promoter, they studied the effect of their inducers dexamethasone (1 M) and phorbol ester phorbol 12-myristate 13-acetate (PMA 50 ng/ml) on CAT activity. However, the MN promoter was unresponsive to those compounds.

The following provides explanations for the results: --the putative MN promoter immediately preceding the transcription initiation site is very weak, and its activity is below the sensitivity of a standard CAT assay; --additional sequences (e.g. enhancers) are necessary for MN transcription.

To further shed light on the regulation of MN expression at the level of transcription, constructs, analogously prepared to the MN-CAT constructs, are prepared, wherein the MN promoter region is upstream from the neomycin phosphotransferase gene engineered for mammalian expression. Such constructs are then transfected to cells which are subjected to selection with G418. Activity of the promoter is then evaluated on the basis of the number of G418 resistant colonies that result. That method has the capacity to detect activity of a promoter that is 50 to 100 times weaker in comparison to promoters detectable by a CAT assay.

Deduced Amino Acid Sequence

The ORF of the MN cDNA shown in Figure 1 has the coding capacity for a 459 amino acid protein with a calculated molecular weight of 49.7 kd. MN protein has an estimated pI of about 4. As assessed by amino acid sequence analysis, the deduced primary structure of the MN protein can be divided into four distinct regions. The initial hydrophobic region of

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37 amino acids (AA) corresponds to a signal peptide. The mature protein has an N-terminal part of 377 AA, a hydrophobic transmembrane segment of 20 AA and a C-terminal region of 25 AA. Alternatively, the MN protein can be viewed as having

5 five domains as follows: (1) a signal peptide [amino acids (AA) 1-37; SEQ. ID. NO.: 6]; (2) a region of homology to collagen alpha1 chain (AA 38-135; SEQ. ID. NO.: 50); (3) a carbonic anhydrase domain (AA 136-391; SEQ. ID. NO.: 51); (4) a transmembrane region (AA 414-433; SEQ. ID. NO.: 52); and

10 (5) an intracellular C terminus (AA 434-459; SEQ. ID. NO.: 53). (The AA numbers are keyed to Figure 1.)

More detailed insight into MN protein primary structure disclosed the presence of several consensus sequences. One potential N-glycosylation site was found at

15 position 346 of Figure 1. That feature, together with a predicted membrane-spanning region are consistent with the results, in which MN was shown to be an N-glycosylated protein localized in the plasma membrane. MN protein sequence deduced from cDNA was also found to contain seven S/TPXX sequence

20 elements [SEQ. ID. NOS.: 25 AND 26] (one of them is in the signal peptide) defined by Suzuki, J. Mol. Biol., 207: 61-84 (1989) as motifs frequently found in gene regulatory proteins. However, only two of them are composed of the suggested consensus amino acids.

25 Experiments have shown that the MN protein is able to bind zinc cations, as shown by affinity chromatography using Zn-charged chelating sepharose. MN protein immunoprecipitated from HeLa cells by Mab M75 was found to have weak catalytic activity of CA. The CA-like domain of MN

30 has a structural predisposition to serve as a binding site for small soluble domains. Thus, MN protein could mediate some kind of signal transduction.

MN protein from LCMV-infected HeLa cells was shown by using DNA cellulose affinity chromatography to bind to

35 immobilized double-stranded salmon sperm DNA. The binding activity required both the presence of zinc cations and the absence of a reducing agent in the binding buffer.

Sequence Similarities

Computer analysis of the MN cDNA sequence was carried out using DNASIS and PROSIS (Pharmacia Software packages). GenBank, EMBL, Protein Identification Resource and SWISS-PROT databases were searched for all possible sequence similarities. In addition, a search for proteins sharing sequence similarities with MN was performed in the MIPS databank with the FastA program [Pearson and Lipman, PNAS (USA), 85: 2444 (1988)].

The MN gene was found to clearly be a novel sequence derived from the human genome. Searches for amino acid sequence similarities in protein databases revealed as the closest homology a level of sequence identity (38.9% in 256 AA or 44% in an 170 AA overlap) between the central part of the MN protein [AAs 136-391 (SEQ. ID. NO: 51)] or 221-390 [SEQ. ID. NO.: 54] of Figure 1 and carbonic anhydrases (CA). However, the overall sequence homology between the cDNA MN sequence and cDNA sequences encoding different CA isoenzymes is in a homology range of 48-50% which is considered by ones in the art to be low. Therefore, the MN cDNA sequence is not closely related to any CA cDNA sequences.

Only very closely related nt sequences having a homology of at least 80-90% would hybridize to each other under stringent conditions. A sequence comparison of the MN cDNA sequence shown in Figure 1 and a corresponding cDNA of the human carbonic anhydrase II (CA II) showed that there are no stretches of identity between the two sequences that would be long enough to allow for a segment of the CA II cDNA sequence having 50 or more nucleotides to hybridize under stringent hybridization conditions to the MN cDNA or vice versa.

Although MN deduced amino acid sequences show some homology to known carbonic anhydrases, they differ from them in several respects. Seven carbonic anhydrases are known [Dodgson et al. (eds.), The Carbonic Anhydrases, (Plenum Press; New York/London (1991))]. All of the known carbonic anhydrases are proteins of about 30 kd, smaller than the p54/58N-related products of the MN gene. Further, the

carbonic anhydrases do not form oligomers as do the MN-related proteins.

The N-terminal part of the MN protein (AA 38-135; SEQ. ID. NO.: 50) shows a 27-30% identity with human collagen
5 alpha1 chain, which is an important component of the extracellular matrix.

MN Proteins and/or Polypeptides

The phrase "MN proteins and/or polypeptides" (MN proteins/polypeptides) is herein defined to mean proteins
10 and/or polypeptides encoded by an MN gene or fragments thereof. An exemplary and preferred MN protein according to this invention has the deduced amino acid sequence shown in Figure 1. Preferred MN proteins/polypeptides are those
15 proteins and/or polypeptides that have substantial homology with the MN protein shown in Figure 1. For example, such substantially homologous MN proteins/ polypeptides are those that are reactive with the MN-specific antibodies of this invention, preferably the Mabs M75, MN12, MN9 and MN7 or their equivalents.

20 A "polypeptide" is a chain of amino acids covalently bound by peptide linkages and is herein considered to be composed of 50 or less amino acids. A "protein" is herein defined to be a polypeptide composed of more than 50 amino acids.

25 MN proteins exhibit several interesting features: cell membrane localization, cell density dependent expression in HeLa cells, correlation with the tumorigenic phenotype of HeLa x fibroblast somatic cell hybrids, and expression in several human carcinomas among other tissues. As demonstrated
30 herein, for example, in Example 1, MN protein can be found directly in tumor tissue sections but not in general in counterpart normal tissues (exceptions noted infra in Example 1 as in normal stomach tissues). MN is also expressed
sometimes in morphologically normal appearing areas of tissue
35 specimens exhibiting dysplasia and/or malignancy. Taken together, these features suggest a possible involvement of MN

in the regulation of cell proliferation, differentiation and/or transformation.

It can be appreciated that a protein or polypeptide produced by a neoplastic cell in vivo could be altered in sequence from that produced by a tumor cell in cell culture or by a transformed cell. Thus, MN proteins and/or polypeptides which have varying amino acid sequences including without limitation, amino acid substitutions, extensions, deletions, truncations and combinations thereof, fall within the scope of this invention. It can also be appreciated that a protein extant within body fluids is subject to degradative processes, such as, proteolytic processes; thus, MN proteins that are significantly truncated and MN polypeptides may be found in body fluids, such as, sera. The phrase "MN antigen" is used herein to encompass MN proteins and/or polypeptides.

It will further be appreciated that the amino acid sequence of MN proteins and polypeptides can be modified by genetic techniques. One or more amino acids can be deleted or substituted. Such amino acid changes may not cause any measurable change in the biological activity of the protein or polypeptide and result in proteins or polypeptides which are within the scope of this invention, as well as, MN muteins.

The MN proteins and polypeptides of this invention can be prepared in a variety of ways according to this invention, for example, recombinantly, synthetically or otherwise biologically, that is, by cleaving longer proteins and polypeptides enzymatically and/or chemically. A preferred method to prepare MN proteins is by a recombinant means. Particularly preferred methods of recombinantly producing MN proteins are described below for the GEX-3X-MN, MN 20-19, MN-Fc and MN-PA proteins.

Recombinant Production of MN Proteins and Polypeptides

A representative method to prepare the MN proteins shown in Figure 1 or fragments thereof would be to insert the full-length or an appropriate fragment of MN cDNA into an appropriate expression vector as exemplified below. In Zavada et al., WO 93/18152, supra, production of a fusion protein

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GEX-3X-MN using the partial cDNA clone (described above) in the vector pGEX-3X (Pharmacia) is described. Nonglycosylated GEX-3X-MN (the Mn fusion protein MN glutathione S-transferase) from XL1-Blue cells. Herein described is the recombinant production of both a glycosylated MN protein expressed from insect cells and a nonglycosylated MN protein expressed from E. coli using the expression plasmid pEt-22b [Novagen Inc.; Madison, WI (USA)].

Baculovirus Expression Systems. Recombinant baculovirus express vectors have been developed for infection into several types of insect cells. For example, recombinant baculoviruses have been developed for among others: Aedes aegypti, Autographa californica, Bombyx mor, Drosophila melanogaster, Heliothis zea, Spodoptera frugiperda, and Trichoplusia ni [PCT Pub. No. WO 89/046699; Wright, Nature, 321: 718 (1986); Fraser et al., In Vitro Cell Dev. Biol., 25: 225 (1989). Methods of introducing exogenous DNA into insect hosts are well-known in the art. DNA transfection and viral infection procedures usually vary with the insect genus to be transformed. See, for example, Autographa [Carstens et al., Virology, 101: 311 (1980)]; Spodoptera [Kang, "Baculovirus Vectors for Expression of Foreign Genes," in: Advances in Virus Research, 35 (1988)]; and Heliothis (virescens) [PCT Pub. No. WO 88/02030].

A wide variety of other host-cloning vector combinations may be usefully employed in cloning the MN DNA isolated as described herein. For example, useful cloning vehicles may include chromosomal, nonchromosomal and synthetic DNA sequences such as various known bacterial plasmids such as pBR322, other E. coli plasmids and their derivatives and wider host range plasmids such as RP4, phage DNA, such as, the numerous derivatives of phage lambda, e.g., NB989 and vectors derived from combinations of plasmids and phage DNAs such as plasmids which have been modified to employ phage DNA expression control sequences.

Useful hosts may be eukaryotic or prokaryotic and include bacterial hosts such as E. coli and other bacterial strains, yeasts and other fungi, animal or plant hosts such as

animal or plant cells in culture, insect cells and other hosts. Of course, not all hosts may be equally efficient. The particular selection of host-cloning vehicle combination may be made by those of skill in the art after due

5 consideration of the principles set forth herein without departing from the scope of this invention.

The particular site chosen for insertion of the selected DNA fragment into the cloning vehicle to form a recombinant DNA molecule is determined by a variety of
10 factors. These include size and structure of the protein or polypeptide to be expressed, susceptibility of the desired protein or polypeptide to endoenzymatic degradation by the host cell components and contamination by its proteins, expression characteristics such as the location of start and
15 stop codons, and other factors recognized by those of skill in the art.

The recombinant nucleic acid molecule containing the MN gene, fragment thereof, or cDNA therefrom, may be employed to transform a host so as to permit that host (transformant)
20 to express the structural gene or fragment thereof and to produce the protein or polypeptide for which the hybrid DNA encodes. The recombinant nucleic acid molecule may also be employed to transform a host so as to permit that host on replication to produce additional recombinant nucleic acid
25 molecules as a source of MN nucleic acid and fragments thereof. The selection of an appropriate host for either of those uses is controlled by a number of factors recognized in the art. These include, for example, compatibility with the chosen vector, toxicity of the co-products, ease of recovery
30 of the desired protein or polypeptide, expression characteristics, biosafety and costs.

Where the host cell is a procaryote such as E. coli, competent cells which are capable of DNA uptake are prepared from cells harvested after exponential growth phase and
35 subsequently treated by the CaCl_2 method by well known procedures. Transformation can also be performed after forming a protoplast of the host cell.

Where the host used is an eucaryote, transfection methods such as the use of a calcium phosphate-precipitate, electroporation, conventional mechanical procedures such as microinjection, insertion of a plasmid encapsulated in red blood cell ghosts or in liposomes, treatment of cells with agents such as lysophosphatidyl-choline or use of virus vectors, or the like may be used.

The level of production of a protein or polypeptide is governed by three major factors: (1) the number of copies of the gene or DNA sequence encoding for it within the cell; (2) the efficiency with which those gene and sequence copies are transcribed and translated; and (3) the stability of the mRNA. Efficiencies of transcription and translation (which together comprise expression) are in turn dependent upon nucleotide sequences, normally situated ahead of the desired coding sequence. Those nucleotide sequences or expression control sequences define, inter alia, the location at which an RNA polymerase interacts to initiate transcription (the promoter sequence) and at which ribosomes bind and interact with the mRNA (the product of transcription) to initiate translation. Not all such expression control sequences function with equal efficiency. It is thus of advantage to separate the specific coding sequences for the desired protein from their adjacent nucleotide sequences and fuse them instead to known expression control sequences so as to favor higher levels of expression. This having been achieved, the newly engineered DNA fragment may be inserted into a multicopy plasmid or a bacteriophage derivative in order to increase the number of gene or sequence copies within the cell and thereby further improve the yield of expressed protein.

Several expression control sequences may be employed. These include the operator, promoter and ribosome binding and interaction sequences (including sequences such as the Shine-Dalgarno sequences) of the lactose operon of E. coli ("the lac system"), the corresponding sequences of the tryptophan synthetase system of E. coli ("the trp system"), a fusion of the trp and lac promoter ("the tac system"), the major operator and promoter regions of phage lambda ($O_L P_L$ and

O_RP_R), and the control region of the phage fd coat protein. DNA fragments containing these sequences are excised by cleavage with restriction enzymes from the DNA isolated from transducing phages that carry the lac or trp operons, or from the DNA of phage lambda or fd. Those fragments are then manipulated in order to obtain a limited population of molecules such that the essential controlling sequences can be joined very close to, or in juxtaposition with, the initiation codon of the coding sequence.

The fusion product is then inserted into a cloning vehicle for transformation or transfection of the appropriate hosts and the level of antigen production is measured. Cells giving the most efficient expression may be thus selected. Alternatively, cloning vehicles carrying the lac, trp or lambda P_L control system attached to an initiation codon may be employed and fused to a fragment containing a sequence coding for a MN protein or polypeptide such that the gene or sequence is correctly translated from the initiation codon of the cloning vehicle.

The phrase "recombinant nucleic acid molecule" is herein defined to mean a hybrid nucleotide sequence comprising at least two nucleotide sequences, the first sequence not normally being found together in nature with the second.

The phrase "expression control sequence" is herein defined to mean a sequence of nucleotides that controls and regulates expression of structural genes when operatively linked to those genes.

The following are representative examples of genetically engineering MN proteins of this invention. The descriptions are exemplary and not meant to limit the invention in any way.

Expression of MN 20-19 Protein

A representative, recombinantly produced MN protein of this invention is the MN 20-19 protein which, when produced in baculovirus-infected Sf9 cells [Spodoptera frugiperda cells; Clontech; Palo Alto, CA (USA)], is glycosylated. The MN 20-19 protein misses the putative signal peptide (AAs 1-37)

of SEQ. ID. NO.: 6 (Figure 1), has a methionine (Met) at the N-terminus for expression, and a Leu-Glu-His-His-His-His-His [SEQ. ID NO.: 22] added to the C-terminus for purification.

5 In order to insert the portion of the MN coding sequence for the GEX-3X-MN fusion protein into alternate expression systems, a set of primers for PCR was designed. The primers were constructed to provide restriction sites at each end of the coding sequence, as well as in-frame start and
10 stop codons. The sequences of the primers, indicating restriction enzyme cleavage sites and expression landmarks, are shown below.

Primer #20:N-terminus

15 5' GTCGCTAGCTCCATGGGTCATATGCAGAGGTTGCCCCGGATGCAG 3'
NheI NcoI NdeI Translation start MN cDNA #1 [SEQ. ID. NO. 17]

Primer #19:C-terminus

20 5' GAAGATCTCTTACTCGAGCATTCTCCAAGATCCAGCCTCTAGG 3'
BglII XhoI Translation stop MN cDNA [SEQ. ID. NO. 18]

The SEQ. ID. NOS.: 17 and 18 primers were used to amplify the MN coding sequence present in the GEX-3X-MN vector using standard PCR techniques. The resulting PCR product (termed MN 20-19) was electrophoresed on a 0.5% agarose/1X TBE gel; the
25 1.3 kb band was excised; and the DNA recovered using the Gene Clean II kit according to the manufacturer's instructions [Bio101; LaJolla, CA (USA)].

30 MN 20-19 and plasmid pET-22b were cleaved with the restriction enzymes NdeI and XhoI, phenol-chloroform extracted, and the appropriate bands recovered by agarose gel electrophoresis as above. The isolated fragments were ethanol co-precipitated at a vector:insert ratio of 1:4. After

resuspension, the fragments were ligated using T4 DNA ligase. The resulting product was used to transform competent Novablue E. coli cells [Novagen, Inc.]. Plasmid mini-preps [Magic Minipreps; Promega] from the resultant ampicillin resistant colonies were screened for the presence of the correct insert by restriction mapping. Insertion of the gene fragment into the pET-22b plasmid using the NdeI and XhoI sites added a 6-histidine tail to the protein that could be used for affinity isolation.

To prepare MN 20-19 for insertion into the baculovirus expression system, the MN 20-19 gene fragment was excised from pET-22b using the restriction endonucleases XbaI and PvuI. The baculovirus shuttle vector pBacPAK8 [Clontech] was cleaved with XbaI and PacI. The desired fragments (1.3 kb for MN 20-19 and 5.5 kb for pBacPAK8) were isolated by agarose gel electrophoresis, recovered using Gene Clean II, and co-precipitated at an insert:vector ratio of 2.4:1.

After ligation with T4 DNA ligase, the DNA was used to transform competent NM522 E. coli cells (Stratagene).

Plasmid mini-preps from resultant ampicillin resistant colonies were screened for the presence of the correct insert by restriction mapping. Plasmid DNA from an appropriate colony and linearized BacPAK6 baculovirus DNA [Clontech] were used to transform Sf9 cells by standard techniques.

Recombination produced BacPAK viruses carrying the MN 20-19 sequence. Those viruses were plated onto Sf9 cells and overlaid with agar.

Plaques were picked and plated onto Sf9 cells. The conditioned media and cells were collected. A small aliquot of the conditioned media was set aside for testing. The cells were extracted with PBS with 1% Triton X100.

The conditioned media and the cell extracts were dot blotted onto nitrocellulose paper. The blot was blocked with 5% non-fat dried milk in PBS. Mab M75 were used to detect the MN 20-19 protein in the dot blots. A rabbit anti-mouse Ig-HRP was used to detect bound Mab M75. The blots were developed with TMB/H₂O₂ with a membrane enhancer [KPL; Gaithersburg, MD (USA)]. Two clones producing the strongest reaction on the

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dot blots were selected for expansion. One was used to produce MN 20-19 protein in High Five cells [Invitrogen Corp., San Diego, CA (USA); BTI-TN-5BI-4; derived from Trichoplusia ni egg cell homogenate]. MN 20-19 protein was purified from the conditioned media from the virus infected High Five cells.

The MN 20-19 protein was purified from the conditioned media by immunoaffinity chromatography. 6.5 mg of Mab M75 was coupled to 1 g of Tresyl activated Toyopearl™ [Tosoh, Japan (#14471)]. Approximately 150 ml of the conditioned media was run through the M75-Toyopearl column. The column was washed with PBS, and the MN 20-19 protein was eluted with 1.5 M MgCl. The eluted protein was then dialyzed against PBS.

Fusion Proteins with C-Terminal Part Including Transmembrane Region Replaced by Fc or PA

MN fusion proteins in which the C terminal part including the transmembrane region is replaced by the Fc fragment of human IgG or by Protein A were constructed. Such fusion proteins are useful to identify MN binding protein(s). In such MN chimaeras, the whole N-terminal part of MN is accessible to interaction with heterologous proteins, and the C terminal tag serves for simple detection and purification of protein complexes.

Fusion Protein MN-PA (Protein A)

In a first step, the 3' end of the MN cDNA encoding the transmembrane region of the MN protein was deleted. The plasmid pFLMN (e.g. pBluescript with full length MN cDNA) was cleaved by EcoRI and blunt ended by S1 nuclease. Subsequent cleavage by SacI resulted in the removal of the EcoRI-SacI fragment. The deleted fragment was then replaced by a Protein A coding sequence that was derived from plasmid pEZZ (purchased from Pharmacia), which had been cleaved with RsaI and SacI. The obtained MN-PA construct was subcloned into a eukaryotic expression vector pSG5C (described in Example 3), and was then ready for transfection experiments.

Fusion Protein MN-Fc

The cloning of the fusion protein MN-Fc was rather complicated due to the use of a genomic clone containing the Fc fragment of human IgG which had a complex structure in that it contained an enhancer, a promoter, exons and introns. Moreover, the complete sequence of the clone was not available. Thus, it was necessary to ensure the correct in-phase splicing and fusion of MN to the Fc fragment by the addition of a synthetic splice donor site (SSDS) designed according to the splicing sequences of the MN gene.

The construction procedure was as follows:

1. Plasmid pMH4 (e.g. pSV2gpt containing a genomic clone of the human IgG Fc region) was cleaved by *Bam*HI in order to get a 13 kb fragment encoding Fc. [In pSV2gpt, the *E. coli* xanthine-guanine phosphoribosyl transferase gene (*gpt*) is expressed using the SV40 early promoter (P_E) located in the SV40 origin, the SV40 small T intron, and the SV40 polyadenylation site.]

2. At the same time, plasmid pFLMN (with full length MN cDNA) was cleaved by *Sal*I-*Eco*RI. The released fragment was purified and ligated with a synthetic adapter *Eco*RI-*Bgl*III containing a synthetic splice donor site (SSDS).

3. Simultaneously, the plasmid pBKCMV was cleaved by *Sal*I-*Bam*HI. Then advantage was taken of the fact that the *Bam*HI cohesive ends (of the Fc coding fragment) are compatible with the *Bgl*III ends of the SSDS, and Fc was ligated to MN. The MN-Fc ligation product was then inserted into pBKCMV by directional cloning through the *Sal*I and *Bam*HI sites.

Verification of the correct orientation and in-phase fusion of the obtained MN-Fc chimaeric clones was problematic in that the sequence of Fc was not known. Thus, functional constructs are selected on the basis of results of transient eukaryotic expression analyses.

Synthetic and Biologic Production of MN Proteins and Polypeptides

MN proteins and polypeptides of this invention may be prepared not only by recombinant means but also by

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synthetic and by other biologic means. Synthetic formation of the polypeptide or protein requires chemically synthesizing the desired chain of amino acids by methods well known in the art. Exemplary of other biologic means to prepare the desired
5 polypeptide or protein is to subject to selective proteolysis a longer MN polypeptide or protein containing the desired amino acid sequence; for example, the longer polypeptide or protein can be split with chemical reagents or with enzymes.

Chemical synthesis of a peptide is conventional in
10 the art and can be accomplished, for example, by the Merrifield solid phase synthesis technique [Merrifield, J., Am. Chem. Soc., 85: 2149-2154 (1963); Kent et al., Synthetic Peptides in Biology and Medicine, 29 f.f. eds. Alitalo et al., (Elsevier Science Publishers 1985); and Haug, J.D., "Peptide
15 Synthesis and Protecting Group Strategy", American Biotechnology Laboratory, 5(1): 40-47 (Jan/Feb. 1987)].

Techniques of chemical peptide synthesis include using automatic peptide synthesizers employing commercially available protected amino acids, for example, Biosearch [San
20 Rafael, CA (USA)] Models 9500 and 9600; Applied Biosystems, Inc. [Foster City, CA (USA)] Model 430; Milligen [a division of Millipore Corp.; Bedford, MA (USA)] Model 9050; and Du Pont's RAMP (Rapid Automated Multiple Peptide Synthesis) [Du Pont Compass, Wilmington, DE (USA)].

25 Regulation of MN Expression and MN Promoter

MN appears to be a novel regulatory protein that is directly involved in the control of cell proliferation and in cellular transformation. In HeLa cells, the expression of MN is positively regulated by cell density. Its level is
30 increased by persistent infection with LCMV. In hybrid cells between HeLa and normal fibroblasts, MN expression correlates with tumorigenicity. The fact that MN is not present in nontumorigenic hybrid cells (CGL1), but is expressed in a tumorigenic segregant lacking chromosome 11, indicates that MN
35 is negatively regulated by a putative suppressor in chromosome 11.

Evidence supporting the regulatory role of MN protein was found in the generation of stable transfectants of NIH 3T3 cells that constitutively express MN protein as described in Example 3. As a consequence of MN expression, the NIH 3T3 cells acquired features associated with a transformed phenotype: altered morphology, increased saturation density, proliferative advantage in serum-reduced media, enhanced DNA synthesis and capacity for anchorage-independent growth. Further, as shown in Example 4, flow cytometric analyses of asynchronous cell populations indicated that the expression of MN protein leads to accelerated progression of cells through G1 phase, reduction of cell size and the loss of capacity for growth arrest under inappropriate conditions. Also, Example 4 shows that MN expressing cells display a decreased sensitivity to the DNA damaging drug mitomycin C.

Nontumorigenic human cells, CGL1 cells, were also transfected with the full-length MN cDNA. The same pSG5C-MN construct in combination with pSV2neo plasmid as used to transfect the NIH 3T3 cells (Example 3) was used. Also the protocol was the same except that the G418 concentration was increased to 1000 $\mu\text{g/ml}$.

Out of 15 MN-positive clones (tested by SP-RIA and Western blotting), 3 were chosen for further analysis. Two MN-negative clones isolated from CGL1 cells transfected with empty plasmid were added as controls. Initial analysis indicates that the morphology and growth habits of MN-transfected CGL1 cells are not changed dramatically, but their proliferation rate and plating efficiency is increased.

MN cDNA and promoter. When the promoter region from the MN genomic clone, isolated as described above, was linked to MN cDNA and transfected into CGL1 hybrid cells, expression of MN protein was detectable immediately after selection. However, then it gradually ceased, indicating thus an action of a feedback regulator. The putative regulatory element appeared to be acting via the MN promoter, because when the full-length cDNA (not containing the promoter) was used for transfection, no similar effect was observed.

An "antisense" MN cDNA/MN promoter construct was used to transfect CGL3 cells. The effect was the opposite of that of the CGL1 cells transfected with the "sense" construct. Whereas the transfected CGL1 cells formed colonies several
5 times larger than the control CGL1, the transfected CGL3 cells formed colonies much smaller than the control CGL3 cells.

For those experiments, the part of the promoter region that was linked to the MN cDNA through a BamHI site was derived from a NcoI - BamHI fragment of the MN genomic clone
10 [Bd3] and represents a region a few hundred bp upstream from the transcription initiation site. After the ligation, the joint DNA was inserted into a pBK-CMV expression vector [Stratagene]. The required orientation of the inserted
15 sequence was ensured by directional cloning and subsequently verified by restriction analysis. The tranfection procedure was the same as used in transfecting the NIH 3T3 cells (Example 3), but co-transfection with the pSV2neo plasmid was not necessary since the neo selection marker was already included in the pBK-CMV vector.

20 After two weeks of selection in a medium containing G418, remarkable differences between the numbers and sizes of the colonies grown were evident as noted above. Immediately following the selection and cloning, the MN-transfected CGL1 and CGL3 cells were tested by SP-RIA for expression and
25 repression of MN, respectively. The isolated transfected CGL1 clones were MN positive (although the level was lower than obtained with the full-length cDNA), whereas MN protein was almost absent from the transfected CGL3 clones. However, in subsequent passages, the expression of MN in transfected CGL1
30 cells started to cease, and was then blocked perhaps evidencing a control feedback mechanism.

As a result of the very much lowered proliferation of the transfected CGL3 cells, it was difficult to expand the majority of cloned cells (according to SP-RIA, those with the
35 lowest levels of MN), and they were lost during passaging. However, some clones overcame that problem and again expressed MN. It is possible that once those cells reached a higher quantity, that the level of endogenously produced MN mRNA

increased over the amount of ectopically expressed antisense mRNA.

Transformation and Reversion

As illustrated in Examples 3 and 4, vertebrate cells
5 transfected with MN cDNA in suitable vectors show striking
morphologic transformation. Transformed cells may be very
small, densely packed, slowly growing, with basophilic
cytoplasm and enlarged Golgi apparatus. However, it has been
found that transformed clones revert over time, for example,
10 within 3-4 weeks, to nearly normal morphology, even though the
cells may be producing MN protein at high levels. MN protein
is biologically active even in yeast cells; depending upon the
level of its expression, it stimulates or retards their growth
and induces morphologic alterations.

15 Full-length MN cDNA was inserted into pGD, a MLV-
derived vector, which together with standard competent MLV
(murine leukemia virus), forms an infectious, transmissible
complex [pGD-MN + MLV]. That complex also transforms
vertebrate cells, such as, NIH 3T3 cells and mouse embryo
20 fibroblasts BALB/c, which also revert to nearly normal
morphology. Such revertants again contain MN protein and
produce the [pGD-MN + MLV] artificial virus complex, which
retains its transforming capacity. Thus, reversion of MN-
transformed cells is apparently not due to a loss, silencing
25 or mutation of MN cDNA, but may be the result of the
activation of suppressor gene(s).

Nucleic Acid Probes and Test Kits

Nucleic acid probes of this invention are those
comprising sequences that are complementary or substantially
30 complementary to the MN cDNA sequence shown in Figure 1 or to
other MN gene sequences, such as, the complete genomic
sequence of Figure 3a-d [SEQ. ID. NO.: 5] and the putative
promoter sequence [SEQ. ID. NO.: 27 of Figure 6]. The phrase
"substantially complementary" is defined herein to have the
35 meaning as it is well understood in the art and, thus, used in
the context of standard hybridization conditions. The

stringency of hybridization conditions can be adjusted to control the precision of complementarity. Two nucleic acids are, for example, substantially complementary to each other, if they hybridize to each other under stringent hybridization conditions.

Stringent hybridization conditions are considered herein to conform to standard hybridization conditions understood in the art to be stringent. For example, it is generally understood that stringent conditions encompass relatively low salt and/or high temperature conditions, such as provided by 0.02 M to 0.15 M NaCl at temperatures of 50°C to 70°C. Less stringent conditions, such as, 0.15 M to 0.9 M salt at temperatures ranging from 20°C to 55°C can be made more stringent by adding increasing amounts of formamide, which serves to destabilize hybrid duplexes as does increased temperature.

Exemplary stringent hybridization conditions are described in Sambrook et al., Molecular Cloning: A Laboratory Manual, pages 1.91 and 9.47-9.51 (Second Edition, Cold Spring Harbor Laboratory Press; Cold Spring Harbor, NY; 1989); Maniatis et al., Molecular Cloning: A Laboratory Manual, pages 387-389 (Cold Spring Harbor Laboratory; Cold Spring Harbor, NY; 1982); Tsuchiya et al., Oral Surgery, Oral Medicine, Oral Pathology, 71(6): 721-725 (June 1991).

Preferred nucleic acid probes of this invention are fragments of the isolated nucleic acid sequences that encode MN proteins or polypeptides according to this invention. Preferably those probes are composed of at least twenty-nine nucleotides, more preferably, fifty nucleotides.

Nucleic acid probes of this invention need not hybridize to a coding region of MN. For example, nucleic acid probes of this invention may hybridize partially or wholly to a non-coding region of the genomic sequence shown in Figure 3a-d [SEQ. ID. NO.: 5]. Conventional technology can be used to determine whether fragments of SEQ. ID. NO.: 5 or related nucleic acids are useful to identify MN nucleic acid sequences. [See, for example, Benton and Davis, supra and Fuscoe et al., supra.]

Areas of homology of the MN nt sequence to other non-MN nt sequences are indicated above. In general, nucleotide sequences that are not in the Alu or LTR-like regions, of preferably 29 bases or more, or still more preferably of 50 bases or more, can be routinely tested and screened and found to hybridize under stringent conditions to only MN nucleotide sequences. Further, not all homologies within the Alu-like MN genomic sequences are so close to Alu repeats as to give a hybridization signal under stringent hybridization conditions. The percent of homology between MN Alu-like regions and a standard Alu-J sequence are indicated as follows:

15	<u>Region of Homology within</u>	<u>SEQ.</u>	<u>% Homology to</u>
	<u>MN Genomic Sequence</u>		
	<u>[SEQ. ID. NO.: 5;</u>	<u>ID.</u>	<u>Entire Alu-J</u>
	<u>Figure 3a-d]</u>	<u>NOS.</u>	<u>Sequence</u>
	921-1212	59	89.1%
	2370-2631	60	78.6%
	4587-4880	61	90.1%
20	6463-6738	62	85.4%
	7651-7939	63	91.0%
	9020-9317	64	69.8%
			<u>% Homology to</u>
			<u>One Half of</u>
			<u>Alu-J Sequence</u>
	8301-8405	65	88.8%
	10040-10122	66	73.2%.

25 Nucleic acid probes of this invention can be used to detect MN DNA and/or RNA, and thus can be used to test for the presence or absence of MN genes, and amplification(s), mutation(s) or genetic rearrangements of MN genes in the cells of a patient. For example, overexpression of an MN gene may
30 be detected by Northern blotting and RNase protection analysis using probes of this invention. Gene alterations, as amplifications, translocations, inversions, and deletions among others, can be detected by using probes of this

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invention for in situ hybridization to chromosomes from a patient's cells, whether in metaphase spreads or interphase nuclei. Southern blotting could also be used with the probes of this invention to detect amplifications or deletions of MN genes. Restriction Fragment Length Polymorphism (RFLP) analysis using said probes is a preferred method of detecting gene alterations, mutations and deletions. Said probes can also be used to identify MN proteins and/or polypeptides as well as homologs or near homologs thereto by their hybridization to various mRNAs transcribed from MN genes in different tissues.

Probes of this invention thus can be useful diagnostically/prognostically. Said probes can be embodied in test kits, preferably with appropriate means to enable said probes when hybridized to an appropriate MN gene or MN mRNA target to be visualized. Such samples include tissue specimens including smears, body fluids and tissue and cell extracts.

PCR Assays

To detect relatively large genetic rearrangements, hybridization tests can be used. To detect relatively small genetic rearrangements, as, for example, small deletions or amplifications, or point mutations, PCR would preferably be used. [U.S. Patent Nos. 4,800,159; 4,683,195; 4,683,202; and Chapter 14 of Sambrook et al., Molecular Cloning: A Laboratory Manual, supra]

An exemplary assay would use cellular DNA from normal and cancerous cells, which DNA would be isolated and amplified employing appropriate PCR primers. The PCR products would be compared, preferably initially, on a sizing gel to detect size changes indicative of certain genetic rearrangements. If no differences in sizes are noted, further comparisons can be made, preferably using, for example, PCR-single-strand conformation polymorphism (PCR-SSCP) assay or a denaturing gradient gel electrophoretic assay. [See, for example, Hayashi, K., "PCR-SSCP: A Simple and Sensitive Method for Detection of Mutations in the Genomic DNA," in PCR

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Methods and Applications, 1: 34-38 (1991); and Meyers et al., "Detection and Localization of Single Base Changes by Denaturing Gradient Gel Electrophoresis," Methods in Enzymology, 155: 501 (1987).]

5

Assays

Assays according to this invention are provided to detect and/or quantitate MN antigen or MN-specific antibodies in vertebrate samples, preferably mammalian samples, more preferably human samples. Such samples include tissue
10 specimens, body fluids, tissue extracts and cell extracts. MN antigen may be detected by immunoassay, immunohistochemical staining, immunoelectron and scanning microscopy using immunogold among other techniques.

Preferred tissue specimens to assay by
15 immunohistochemical staining include cell smears, histological sections from biopsied tissues or organs, and imprint preparations among other tissue samples. Such tissue specimens can be variously maintained, for example, they can be fresh, frozen, or formalin-, alcohol- or acetone- or
20 otherwise fixed and/or paraffin-embedded and deparaffinized. Biopsied tissue samples can be, for example, those samples removed by aspiration, bite, brush, cone, chorionic villus, endoscopic, excisional, incisional, needle, percutaneous punch, and surface biopsies, among other biopsy techniques.

25 Preferred cervical tissue specimens include cervical smears, conization specimens, histologic sections from hysterectomy specimens or other biopsied cervical tissue samples. Preferred means of obtaining cervical smears include routine swab, scraping or cytobrush techniques, among other
30 means. More preferred are cytobrush or swab techniques. Preferably, cell smears are made on microscope slides, fixed, for example, with 55% EtOH or an alcohol based spray fixative and air-dried.

Papanicolaou-stained cervical smears (Pap smears)
35 can be screened by the methods of this invention, for example, for retrospective studies. Preferably, Pap smears would be decolorized and re-stained with labeled antibodies against MN

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antigen. Also archival specimens, for example, matched smears and biopsy and/or tumor specimens, can be used for retrospective studies. Prospective studies can also be done with matched specimens from patients that have a higher than
5 normal risk of exhibiting abnormal cervical cytopathology.

Preferred samples in which to assay MN antigen by, for example, Western blotting or radioimmunoassay, are tissue and/or cell extracts. However, MN antigen may be detected in body fluids, which can include among other fluids: blood,
10 serum, plasma, semen, breast exudate, saliva, tears, sputum, mucous, urine, lymph, cytosols, ascites, pleural effusions, amniotic fluid, bladder washes, bronchioalveolar lavages and cerebrospinal fluid. It is preferred that the MN antigen be concentrated from a larger volume of body fluid before
15 testing. Preferred body fluids to assay would depend on the type of cancer for which one was testing, but in general preferred body fluids would be breast exudate, pleural effusions and ascites.

MN-specific antibodies can be bound by serologically
20 active MN proteins/polypeptides in samples of such body fluids as blood, plasma, serum, lymph, mucous, tears, urine, spinal fluid and saliva; however, such antibodies are found most usually in blood, plasma and serum, preferably in serum. Correlation of the results from the assays to detect and/or
25 quantitate MN antigen and MN-specific antibodies reactive therewith, provides a preferred profile of the disease condition of a patient.

The assays of this invention are both diagnostic and/or prognostic, i.e., diagnostic/prognostic. The term
30 "diagnostic/ prognostic" is herein defined to encompass the following processes either individually or cumulatively depending upon the clinical context: determining the presence of disease, determining the nature of a disease, distinguishing one disease from another, forecasting as to the
35 probable outcome of a disease state, determining the prospect as to recovery from a disease as indicated by the nature and symptoms of a case, monitoring the disease status of a patient, monitoring a patient for recurrence of disease,

and/or determining the preferred therapeutic regimen for a patient. The diagnostic/prognostic methods of this invention are useful, for example, for screening populations for the presence of neoplastic or pre-neoplastic disease, determining
5 the risk of developing neoplastic disease, diagnosing the presence of neoplastic and/or pre-neoplastic disease, monitoring the disease status of patients with neoplastic disease, and/or determining the prognosis for the course of neoplastic disease. For example, it appears that the
10 intensity of the immunostaining with MN-specific antibodies may correlate with the severity of dysplasia present in samples tested.

The present invention is useful for screening for the presence of a wide variety of neoplastic diseases as
15 indicated above. The invention provides methods and compositions for evaluating the probability of the presence of malignant or pre-malignant cells, for example, in a group of cells freshly removed from a host. Such an assay can be used to detect tumors, quantitate their growth, and help in the
20 diagnosis and prognosis of disease. The assays can also be used to detect the presence of cancer metastasis, as well as confirm the absence or removal of all tumor tissue following surgery, cancer chemotherapy and/or radiation therapy. It can further be used to monitor cancer chemotherapy and tumor
25 reappearance.

The presence of MN antigen or antibodies can be detected and/or quantitated using a number of well-defined diagnostic assays. Those in the art can adapt any of the conventional immunoassay formats to detect and/or quantitate
30 MN antigen and/or antibodies.

Many formats for detection of MN antigen and MN-specific antibodies are, of course available. Those can be Western blots, ELISAs, RIAs, competitive EIA or dual antibody sandwich assays, immunohistochemical staining, among other
35 assays all commonly used in the diagnostic industry. In such immunoassays, the interpretation of the results is based on the assumption that the antibody or antibody combination will

not cross-react with other proteins and protein fragments present in the sample that are unrelated to MN.

Representative of one type of ELISA test for MN antigen is a format wherein a microtiter plate is coated with antibodies made to MN proteins/polypeptides or antibodies made to whole cells expressing MN proteins, and to this is added a patient sample, for example, a tissue or cell extract. After a period of incubation permitting any antigen to bind to the antibodies, the plate is washed and another set of anti-MN antibodies which are linked to an enzyme is added, incubated to allow reaction to take place, and the plate is then rewashed. Thereafter, enzyme substrate is added to the microtiter plate and incubated for a period of time to allow the enzyme to work on the substrate, and the adsorbance of the final preparation is measured. A large change in absorbance indicates a positive result.

It is also apparent to one skilled in the art of immunoassays that MN proteins and/or polypeptides can be used to detect and/or quantitate the presence of MN antigen in the body fluids, tissues and/or cells of patients. In one such embodiment, a competition immunoassay is used, wherein the MN protein/polypeptide is labeled and a body fluid is added to compete the binding of the labeled MN protein/polypeptide to antibodies specific to MN protein/polypeptide.

In another embodiment, an immunometric assay may be used wherein a labeled antibody made to a MN protein or polypeptide is used. In such an assay, the amount of labeled antibody which complexes with the antigen-bound antibody is directly proportional to the amount of MN antigen in the sample.

A representative assay to detect MN-specific antibodies is a competition assay in which labeled MN protein/polypeptide is precipitated by antibodies in a sample, for example, in combination with monoclonal antibodies recognizing MN proteins/polypeptides. One skilled in the art could adapt any of the conventional immunoassay formats to detect and/or quantitate MN-specific antibodies. Detection of the binding of said antibodies to said MN protein/polypeptide

could be by many ways known to those in the art, e.g., in humans with the use of anti-human labeled IgG.

An exemplary immunoassay method of this invention to detect and/or quantitate MN antigen in a vertebrate sample
5 comprises the steps of:

a) incubating said vertebrate sample with one or more sets of antibodies (an antibody or antibodies) that bind to MN antigen wherein one set is labeled or otherwise detectable;

10 b) examining the incubated sample for the presence of immune complexes comprising MN antigen and said antibodies.

Another exemplary immunoassay method according to this invention is that wherein a competition immunoassay is used to detect and/or quantitate MN antigen in a vertebrate
15 sample and wherein said method comprises the steps of:

a) incubating a vertebrate sample with one or more sets of MN-specific antibodies and a certain amount of a labeled or otherwise detectable MN protein/polypeptide wherein said MN protein/ polypeptide competes for binding to said
20 antibodies with MN antigen present in the sample;

b) examining the incubated sample to determine the amount of labeled/detectable MN protein/polypeptide bound to said antibodies; and

c) determining from the results of the examination
25 in step b) whether MN antigen is present in said sample and/or the amount of MN antigen present in said sample.

Once antibodies (including biologically active antibody fragments) having suitable specificity have been prepared, a wide variety of immunological assay methods are
30 available for determining the formation of specific antibody-antigen complexes. Numerous competitive and non-competitive protein binding assays have been described in the scientific and patent literature, and a large number of such assays are commercially available. Exemplary
35 immunoassays which are suitable for detecting a serum antigen include those described in U.S. Patent Nos. 3,984,533; 3,996,345; 4,034,074; and 4,098,876.

Antibodies employed in assays may be labeled or unlabeled. Unlabeled antibodies may be employed in agglutination; labeled antibodies may be employed in a wide variety of assays, employing a wide variety of labels.

5 Suitable detection means include the use of labels such as radionuclides, enzymes, coenzymes, fluorescers, chemiluminescers, chromogens, enzyme substrates or co-factors, enzyme inhibitors, free radicals, particles, dyes and the like. Such labeled reagents may be used in a variety of well
10 known assays, such as radioimmunoassays, enzyme immunoassays, e.g., ELISA, fluorescent immunoassays, and the like. See for example, U.S. Patent Nos. 3,766,162; 3,791,932; 3,817,837; and 4,233,402.

Immunoassay Test Kits

15 The above outlined assays can be embodied in test kits to detect and/or quantitate MN antigen and/or MN-specific antibodies (including biologically active antibody fragments). Kits to detect and/or quantitate MN antigen can comprise MN
20 protein(s)/polypeptides(s) and/or MN-specific antibodies, polyclonal and/or monoclonal. Such diagnostic/prognostic test kits can comprise one or more sets of antibodies, polyclonal and/or monoclonal, for a sandwich format wherein antibodies recognize epitopes on the MN antigen, and one set is
appropriately labeled or is otherwise detectable.

25 Test kits for an assay format wherein there is competition between a labeled (or otherwise detectable) MN protein/polypeptide and MN antigen in the sample, for binding to an antibody, can comprise the combination of the labeled
30 protein/polypeptide and the antibody in amounts which provide for optimum sensitivity and accuracy.

 Test kits for MN-specific antibodies preferably comprise labeled/detectable MN proteins(s) and/or
polypeptides(s), and may comprise other components as
35 necessary, such as, controls, buffers, diluents and detergents. Such test kits can have other appropriate formats for conventional assays.

A kit for use in an enzyme-immunoassay typically includes an enzyme-labelled reagent and a substrate for the enzyme. The enzyme can, for example, bind either an MN-specific antibody of this invention or to an antibody to such an MN-specific antibody.

Preparation of MN-Specific Antibodies

The term "antibodies" is defined herein to include not only whole antibodies but also biologically active fragments of antibodies, preferably fragments containing the antigen binding regions. Such antibodies may be prepared by conventional methodology and/or by genetic engineering. Antibody fragments may be genetically engineered, preferably from the variable regions of the light and/or heavy chains (V_H and V_L), including the hypervariable regions, and still more preferably from both the V_H and V_L regions. For example, the term "antibodies" as used herein comprehends polyclonal and monoclonal antibodies and biologically active fragments thereof including among other possibilities "univalent" antibodies [Glennie et al., Nature, 295: 712 (1982)]; Fab proteins including Fab' and $F(ab')_2$ fragments whether covalently or non-covalently aggregated; light or heavy chains alone, preferably variable heavy and light chain regions (V_H and V_L regions), and more preferably including the hypervariable regions [otherwise known as the complementarity determining regions (CDRs) of said V_H and V_L regions]; F_c proteins; "hybrid" antibodies capable of binding more than one antigen; constant-variable region chimeras; "composite" immunoglobulins with heavy and light chains of different origins; "altered" antibodies with improved specificity and other characteristics as prepared by standard recombinant techniques and also by oligonucleotide-directed mutagenesis techniques [Dalbadie-McFarland et al., PNAS (USA), 79: 6409 (1982)].

It may be preferred for therapeutic and/or imaging uses that the antibodies be biologically active antibody fragments, preferably genetically engineered fragments, more preferably genetically engineered fragments from the V_H and/or

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V_L regions, and still more preferably comprising the hypervariable regions thereof.

There are conventional techniques for making polyclonal and monoclonal antibodies well-known in the immunoassay art. Immunogens to prepare MN-specific antibodies include MN proteins and/or polypeptides, preferably purified, and MX-infected tumor line cells, for example, MX-infected HeLa cells, among other immunogens.

Anti-peptide antibodies are also made by conventional methods in the art as described in European Patent Publication No. 44,710 (published Jan. 27, 1982). Briefly, such anti-peptide antibodies are prepared by selecting a peptide from an MN amino acid sequence as from Figure 1, chemically synthesizing it, conjugating it to an appropriate immunogenic protein and injecting it into an appropriate animal, usually a rabbit or a mouse; then, either polyclonal or monoclonal antibodies are made, the latter by a Kohler-Milstein procedure, for example.

Besides conventional hybridoma technology, newer technologies can be used to produce antibodies according to this invention. For example, the use of the PCR to clone and express antibody V-genes and phage display technology to select antibody genes encoding fragments with binding activities has resulted in the isolation of antibody fragments from repertoires of PCR amplified V-genes using immunized mice or humans. [Marks et al., BioTechnology, 10: 779 (July 1992) for references; Chiang et al., BioTechniques, 7(4): 360 (1989); Ward et al., Nature, 341: 544 (Oct. 12, 1989); Marks et al., J. Mol. Biol., 222: 581 (1991); Clackson et al., Nature, 352: (15 August 1991); and Mullinax et al., PNAS (USA), 87: 8095 (Oct. 1990).]

Descriptions of preparing antibodies, which term is herein defined to include biologically active antibody fragments, by recombinant techniques can be found in U.S. Patent No. 4,816,567 (issued March 28, 1989); European Patent Application Publication Number (EP) 338,745 (published Oct. 25, 1989); EP 368,684 (published June 16, 1990); EP 239,400 (published September 30, 1987); WO 90/14424 (published Nov.

29, 1990); WO 90/14430 (published May 16, 1990); Huse et al., Science, 246: 1275 (Dec. 8, 1989); Marks et al., BioTechnology, 10: 779 (July 1992); La Sastry et al., PNAS (USA), 86: 5728 (August 1989); Chiang et al., BioTechniques, 7(40): 360 (1989); Orlandi et al., PNAS (USA), 86: 3833 (May 1989); Ward et al. Nature, 341: 544 (October 12, 1989); Marks et al., J. Mol. Biol., 222: 581 (1991); and Hoogenboom et al., Nucleic Acids Res., 19(15): 4133 (1991).

Representative Mabs

10 Monoclonal antibodies for use in the assays of this invention may be obtained by methods well known in the art for example, Galfre and Milstein, "Preparation of Monoclonal Antibodies: Strategies and Procedures," in Methods in Enzymology: Immunochemical Techniques, 73: 1-46 [Langone and 15 Vanatis (eds); Academic Press (1981)]; and in the classic reference, Milstein and Kohler, Nature, 256: 495-497 (1975).]

 Although representative hybridomas of this invention are formed by the fusion of murine cell lines, human/human hybridomas [Olsson et al., PNAS (USA), 77: 5429 (1980)] and 20 human/murine hybridomas [Schlom et al., PNAS (USA), 77: 6841 (1980); Shearman et al. J. Immunol., 146: 928-935 (1991); and Gorman et al., PNAS (USA), 88: 4181-4185 (1991)] can also be prepared among other possibilities. Such humanized monoclonal antibodies would be preferred monoclonal antibodies for 25 therapeutic and imaging uses.

 Monoclonal antibodies specific for this invention can be prepared by immunizing appropriate mammals, preferably rodents, more preferably rabbits or mice, with an appropriate immunogen, for example, MaTu-infected HeLa cells, MN fusion 30 proteins, or MN proteins/polypeptides attached to a carrier protein if necessary. Exemplary methods of producing antibodies of this invention are described below.

 The monoclonal antibodies useful according to this invention to identify MN proteins/polypeptides can be labeled 35 in any conventional manner, for example, with enzymes such as horseradish peroxidase (HRP), fluorescent compounds, or with radioactive isotopes such as, ¹²⁵I, among other labels. A

preferred label, according to this invention is ^{125}I , and a preferred method of labeling the antibodies is by using chloramine-T [Hunter, W.M., "Radioimmunoassay," In: Handbook of Experimental Immunology, pp. 14.1-14.40 (D.W. Weir ed.; Blackwell, Oxford/London/Edinburgh/Melbourne; 1978)].

Representative mabs of this invention include Mabs M75, MN9, MN12 and MN7 described below. Monoclonal antibodies of this invention serve to identify MN proteins/polypeptides in various laboratory diagnostic tests, for example, in tumor cell cultures or in clinical samples.

Mabs Prepared Against HeLa Cells

MAb M75. Monoclonal antibody M75 (MAb M75) is produced by mouse lymphocytic hybridoma VU-M75, which was initially deposited in the Collection of Hybridomas at the Institute of Virology, Slovak Academy of Sciences (Bratislava, Czechoslovakia) and was deposited under ATCC Designation HB 11128 on September 17, 1992 at the American Type Culture Collection (ATCC) in Rockville, MD (USA). The production of hybridoma VU-M75 is described in Zavada et al., WO 93/18152.

Mab M75 recognizes both the nonglycosylated GEX-3X-MN fusion protein and native MN protein as expressed in CGL3 cells equally well. Mab M75 was shown by epitope mapping to be reactive with the epitope represented by the amino acid sequence from AA 62 to AA 67 [SEQ. ID. NO.: 10] of the MN protein shown in Figure 1.

Mabs Prepared Against Fusion Protein GEX-3X-MN

Monoclonal antibodies of this invention were also prepared against the MN glutathione S-transferase fusion protein (GEX-3X-MN). BALB/C mice were immunized intraperitoneally according to standard procedures with the GEX-3X-MN fusion protein in Freund's adjuvant. Spleen cells of the mice were fused with SP/20 myeloma cells [Milstein and Kohler, supra].

Tissue culture media from the hybridomas were screened against CGL3 and CGL1 membrane extracts in an ELISA employing HRP labelled-rabbit anti-mouse. The membrane

extracts were coated onto microtiter plates. Selected were antibodies reacted with the CGL3 membrane extract. Selected hybridomas were cloned twice by limiting dilution.

The mabs prepared by the just described method were characterized by Western blots of the GEX-3X-MN fusion protein, and with membrane extracts from the CGL1 and CGL3 cells. Representative of the mabs prepared are Mabs MN9, MN12 and MN7.

Mab MN9. Monoclonal antibody MN9 (Mab MN9) reacts to the same epitope as Mab M75, represented by the sequence from AA 62 to AA 67 [SEQ. ID. NO.: 10] of the Figure 1 MN protein. As Mab M75, Mab MN9 recognizes both the GEX-3X-MN fusion protein and native MN protein equally well.

Mabs corresponding to Mab MN9 can be prepared reproducibly by screening a series of mabs prepared against an MN protein/polypeptide, such as, the GEX-3X-MN fusion protein, against the peptide representing the epitope for Mabs M75 and MN9, that is, SEQ. ID. NO.: 10. Alternatively, the Novatope system [Novagen] or competition with the deposited Mab M75 could be used to select mabs comparable to Mabs M75 and MN9.

Mab MN12. Monoclonal antibody MN12 (Mab MN12) is produced by the mouse lymphocytic hybridoma MN 12.2.2 which was deposited under ATCC Designation HB 11647 on June 9, 1994 at the American Type Culture Collection (ATCC) at 12301 Parklawn Drive, Rockville, MD 20852 (USA). Antibodies corresponding to Mab MN12 can also be made, analogously to the method outlined above for Mab MN9, by screening a series of antibodies prepared against an MN protein/polypeptide, against the peptide representing the epitope for Mab MN12. That peptide is AA 55 - AA 60 of Figure 1 [SEQ. ID. NO.: 11]. The Novatope system could also be used to find antibodies specific for said epitope.

Mab MN7. Monoclonal antibody MN7 (Mab MN7) was selected from mabs prepared against nonglycosylated GEX-3X-MN as described above. It recognizes the epitope on MN represented by the amino acid sequence from AA 127 to AA 147 [SEQ. ID. NO.: 12] of the Figure 1 MN protein. Analogously to methods described above for Mabs MN9 and MN12, mabs

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corresponding to Mab MN7 can be prepared by selecting mabs prepared against an MN protein/polypeptide that are reactive with the peptide having SEQ. ID. NO.: 12, or by the stated alternative means.

5 Epitope Mapping

Epitope mapping was performed by the Novatope system, a kit for which is commercially available from Novagen, Inc. [See, for analogous example, Li et al., Nature, 363: 85-88 (6 May 1993).] In brief, the MN cDNA was cut into
10 overlapping short fragments of approximately 60 base pairs. The fragments were expressed in E. coli, and the E. coli colonies were transferred onto nitrocellulose paper, lysed and probed with the mab of interest. The MN cDNA of clones
15 reactive with the mab of interest was sequenced, and the epitopes of the mabs were deduced from the overlapping polypeptides found to be reactive with each mab.

Therapeutic Use of MN-Specific Antibodies

The MN-specific antibodies of this invention, monoclonal and/or polyclonal, preferably monoclonal, and as
20 outlined above, may be used therapeutically in the treatment of neoplastic and/or pre-neoplastic disease, either alone or in combination with chemotherapeutic drugs or toxic agents, such as ricin A. Further preferred for therapeutic use would be biologically active antibody fragments as described herein.
25 Also preferred MN-specific antibodies for such therapeutic uses would be humanized monoclonal antibodies.

The MN-specific antibodies can be administered in a therapeutically effective amount, preferably dispersed in a physiologically acceptable, nontoxic liquid vehicle.

30 Imaging Use of Antibodies

Further, the MN-specific antibodies of this invention when linked to an imaging agent, such as a radionuclide, can be used for imaging. Biologically active antibody fragments or humanized monoclonal antibodies, may be
35 preferred for imaging use.

A patient's neoplastic tissue can be identified as, for example, sites of transformed stem cells, of tumors and locations of any metastases. Antibodies, appropriately labeled or linked to an imaging agent, can be injected in a physiologically acceptable carrier into a patient, and the binding of the antibodies can be detected by a method appropriate to the label or imaging agent, for example, by scintigraphy.

Antisense MN Nucleic Acid Sequences

10 MN genes are herein considered putative oncogenes and the encoded proteins thereby are considered to be putative oncoproteins. Antisense nucleic acid sequences substantially complementary to mRNA transcribed from MN genes, as represented by the antisense oligodeoxynucleotides ODN1 and
15 ODN2 [SEQ. ID. NOS.: 3 and 4] can be used to reduce or prevent expression of the MN gene. [Zamecnick, P.C., "Introduction: Oligonucleotide Base Hybridization as a Modulator of Genetic Message Readout," pp. 1-6, Prospects for Antisense Nucleic Acid Therapy of Cancer and AIDS, (Wiley-
20 Liss, Inc., New York, NY, USA; 1991); Wickstrom, E., "Antisense DNA Treatment of HL-60 Promyelocytic Leukemia Cells: Terminal Differentiation and Dependence on Target Sequence," pp. 7-24, id.; Leserman et al., "Targeting and Intracellular Delivery of Antisense Oligonucleotides
25 Interfering with Oncogene Expression," pp. 25-34, id.; Yokoyama, K., "Transcriptional Regulation of c-myc Proto-oncogene by Antisense RNA," pp. 35-52, id.; van den Berg et al., "Antisense fos Oligodeoxyribonucleotides Suppress the Generation of Chromosomal Aberrations," pp. 63-70, id.;
30 Mercola, D., "Antisense fos and fun RNA," pp. 83-114, id.; Inouye, Gene, 72: 25-34 (1988); Miller and Ts'o, Ann. Reports Med. Chem., 23: 295-304 (1988); Stein and Cohen, Cancer Res., 48: 2659-2668 (1988); Stevenson and Inversen, J. Gen. Virol., 70: 2673-2682 (1989); Goodchild, "Inhibition of Gene
35 Expression by Oligonucleotides," pp. 53-77, Oligodeoxynucleotides: Antisense Inhibitors of Gene Expression (Cohen, J.S., ed; CRC Press, Boca Raton, Florida,

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USA; 1989); Dervan et al., "Oligonucleotide Recognition of Double-helical DNA by Triple-helix Formation," pp. 197-210, id.; Neckers, L.M., "Antisense Oligodeoxynucleotides as a Tool for Studying Cell Regulation: Mechanisms of Uptake and Application to the Study of Oncogene Function," pp. 211-232, id.; Leitner et al., PNAS (USA), 87: 3430-3434 (1990); Bevilacqua et al., PNAS (USA), 85: 831-835 (1988); Loke et al. Curr. Top. Microbiol. Immunol., 141: 282-288 (1988); Sarin et al., PNAS (USA), 85: 7448-7451 (1988); Agrawal et al., "Antisense Oligonucleotides: A Possible Approach for Chemotherapy and AIDS," International Union of Biochemistry Conference on Nucleic Acid Therapeutics (Jan. 13-17, 1991; Clearwater Beach, Florida, USA); Armstrong, L., Ber. Week, pp. 88-89 (March 5, 1990); and Weintraub et al., Trends, 1: 22-25 (1985).] Such antisense nucleic acid sequences, preferably oligonucleotides, by hybridizing to the MN mRNA, particularly in the vicinity of the ribosome binding site and translation initiation point, inhibits translation of the mRNA. Thus, the use of such antisense nucleic acid sequences may be considered to be a form of cancer therapy.

Preferred antisense oligonucleotides according to this invention are gene-specific ODNs or oligonucleotides complementary to the 5' end of MN mRNA. Particularly preferred are the 29-mer ODN1 and 19-mer ODN2 [SEQ. ID. NOS.: 3 and 4]. Those antisense ODNs are representative of the many antisense nucleic acid sequences that can function to inhibit MN gene expression. Ones of ordinary skill in the art could determine appropriate antisense nucleic acid sequences, preferably antisense oligonucleotides, from the nucleic acid sequences of Figures 1 and 3a-d.

Also, as described above, CGL3 cells transfected with an "antisense" MN cDNA/promoter construct formed colonies much smaller than control CGL3 cells.

Vaccines

It will be readily appreciated that MN proteins and polypeptides of this invention can be incorporated into vaccines capable of inducing protective immunity against

neoplastic disease and a dampening effect upon tumorigenic activity. Efficacy of a representative MN fusion protein GEX-3X-MN as a vaccine in a rat model is shown in Example 2.

5 MN proteins and/or polypeptides may be synthesized or prepared recombinantly or otherwise biologically, to comprise one or more amino acid sequences corresponding to one or more epitopes of the MN proteins either in monomeric or multimeric form. Those proteins and/or polypeptides may then be incorporated into vaccines capable of inducing protective
10 immunity. Techniques for enhancing the antigenicity of such polypeptides include incorporation into a multimeric structure, binding to a highly immunogenic protein carrier, for example, keyhole limpet hemocyanin (KLH), or diphtheria toxoid, and administration in combination with adjuvants or
15 any other enhancers of immune response.

Preferred MN proteins/polypeptides to be used in a vaccine according to this invention would be genetically engineered MN proteins. Preferred recombinant MN protein are the GEX-3X-MN, MN 20-19, MN-Fc and MN-PA proteins.

20 Other exemplary vaccines include vaccinia-MN (live vaccinia virus with full-length MN cDNA), and baculovirus-MN (full length MN cDNA inserted into baculovirus vector, e.g. in suspension of infected insect cells). Different vaccines may be combined and vaccination periods can be prolonged.

25 A preferred exemplary use of such a vaccine of this invention would be its administration to patients whose MN-carrying primary cancer had been surgically removed. The vaccine may induce active immunity in the patients and prevent recidivism or metastasis.

30 It will further be appreciated that anti-idiotypic antibodies to antibodies to MN proteins/polypeptides are also useful as vaccines and can be similarly formulated.

An amino acid sequence corresponding to an epitope of an MN protein/polypeptide either in monomeric or multimeric
35 form may also be obtained by chemical synthetic means or by purification from biological sources including genetically modified microorganisms or their culture media. [See Lerner, "Synthetic Vaccines", Sci. Am. 248(2): 66-74 (1983).] The

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protein/polypeptide may be combined in an amino acid sequence with other proteins/polypeptides including fragments of other proteins, as for example, when synthesized as a fusion protein, or linked to other antigenic or non-antigenic polypeptides of synthetic or biological origin. In some instances, it may be desirable to fuse a MN protein or polypeptide to an immunogenic and/or antigenic protein or polypeptide, for example, to stimulate efficacy of a MN-based vaccine.

The term "corresponding to an epitope of an MN protein/polypeptide" will be understood to include the practical possibility that, in some instances, amino acid sequence variations of a naturally occurring protein or polypeptide may be antigenic and confer protective immunity against neoplastic disease and/or anti-tumorigenic effects. Possible sequence variations include, without limitation, amino acid substitutions, extensions, deletions, truncations, interpolations and combinations thereof. Such variations fall within the contemplated scope of the invention provided the protein or polypeptide containing them is immunogenic and antibodies elicited by such a polypeptide or protein cross-react with naturally occurring MN proteins and polypeptides to a sufficient extent to provide protective immunity and/or anti-tumorigenic activity when administered as a vaccine.

Such vaccine compositions will be combined with a physiologically acceptable medium, including immunologically acceptable diluents and carriers as well as commonly employed adjuvants such as Freund's Complete Adjuvant, saponin, alum, and the like. Administration would be in immunologically effective amounts of the MN proteins or polypeptides, preferably in quantities providing unit doses of from 0.01 to 10.0 micrograms of immunologically active MN protein and/or polypeptide per kilogram of the recipient's body weight. Total protective doses may range from 0.1 to about 100 micrograms of antigen. Routes of administration, antigen dose, number and frequency of injections are all matters of optimization within the scope of the ordinary skill in the art.

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The following examples are for purposes of illustration only and not meant to limit the invention in any way.

Example 1

5 Immunohistochemical Staining of Tissue Specimens

To study and evaluate the tissue distribution range and expression of MN proteins, the monoclonal antibody M75 was used to stain immunohistochemically a variety of human tissue specimens. The primary antibody used in these
10 immunohistochemical staining experiments was the M75 monoclonal antibody. A biotinylated second antibody and streptavidin-peroxidase were used to detect the M75 reactivity in sections of formalin-fixed, paraffin-embedded tissue samples. A commercially available amplification kit,
15 specifically the DAKO LSAB™ kit [DAKO Corp., Carpinteria, CA (USA)] which provides matched, ready made blocking reagent, secondary antibody and streptavidin-horseradish peroxidase was used in these experiments.

M75 immunoreactivity was tested according to the
20 methods of this invention in multiple-tissue sections of breast, colon, cervical, lung and normal tissues. Such multiple-tissue sections were cut from paraffin blocks of tissues called "sausages" that were purchased from the City of Hope [Duarte, CA (USA)]. Combined in such a multiple-tissue
25 section were normal, benign and malignant specimens of a given tissue; for example, about a score of tissue samples of breast cancers from different patients, a similar number of benign breast tissue samples, and normal breast tissue samples would be combined in one such multiple-breast-tissue section. The
30 normal multiple-tissue sections contained only normal tissues from various organs, for example, liver, spleen, lung, kidney, adrenal gland, brain, prostate, pancreas, thyroid, ovary, and testis.

Also screened for MN gene expression were multiple
35 individual specimens from cervical cancers, bladder cancers, renal cell cancers, and head and neck cancers. Such specimens were obtained from U.C. Davis Medical Center in Sacramento, CA

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and from Dr. Shu Y. Liao [Department of Pathology; St. Joseph Hospital; Orange, CA (USA)].

Controls used in these experiments were the cell lines CGL3 (H/F-T hybrid cells) and CGL1 (H/F-N hybrid cells) which are known to stain respectively, positively and negatively with the M75 monoclonal antibody. The M75 monoclonal antibody was diluted to a 1:5000 dilution wherein the diluent was either PBS [0.05 M phosphate buffered saline (0.15 M NaCl), pH 7.2-7.4] or PBS containing 1% protease-free BSA as a protein stabilizer.

Immunohistochemical Staining Protocol

The immunohistochemical staining protocol was followed according to the manufacturer's instructions for the DAKO LSAB™ kit. In brief, the sections were dewaxed, rehydrated and blocked to remove non-specific reactivity as well as endogenous peroxidase activity. Each section was then incubated with dilutions of the M75 monoclonal antibody. After the unbound M75 was removed by rinsing the section, the section was sequentially reacted with a biotinylated antimouse IgG antibody and streptavidin conjugated to horseradish peroxidase; a rinsing step was included between those two reactions and after the second reaction. Following the last rinse, the antibody-enzyme complexes were detected by reaction with an insoluble chromogen (diaminobenzidine) and hydrogen peroxide. A positive result was indicated by the formation of an insoluble reddish-brown precipitate at the site of the primary antibody reaction. The sections were then rinsed, counterstained with hematoxylin, dehydrated and cover slipped. Then the sections were examined using standard light microscopy.

Interpretation. A deposit of a reddish brown precipitate over the plasma membrane was taken as evidence that the M75 antibody had bound to a MN antigen in the tissue. The known positive control (CGL3) had to be stained to validate the assay. Section thickness was taken into consideration to compare staining intensities, as thicker

sections produce greater staining intensity independently of other assay parameters.

Results

Preliminary examination of cervical specimens showed
5 that 62 of 68 squamous cell carcinoma specimens (91.2%)
stained positively with M75. Additionally, 2 of 6
adenocarcinomas and 2 of 2 adenosquamous cancers of the cervix
also stained positively. In early studies, 55.6% (10 of 18)
of cervical dysplasias stained positively. A total of 9
10 specimens including both cervical dysplasias and tumors,
exhibited some MN expression in normal appearing areas of the
endocervical glandular epithelium, usually at the basal layer.
In some specimens, whereas morphologically normal-looking
areas showed expression of MN antigen, areas exhibiting
15 dysplasia and/or malignancy did not show MN expression.

M75 positive immunoreactivity was most often
localized to the plasma membrane of cells, with the most
apparent stain being present at the junctions between adjacent
cells. Cytoplasmic staining was also evident in some cells;
20 however, plasma membrane staining was most often used as the
main criterion of positivity.

M75 positive cells tended to be near areas showing
keratin differentiation in cervical specimens. In some
specimens, positive staining cells were located in the center
25 of nests of non-staining cells. Often, there was very little,
if any, obvious morphological difference between staining
cells and non-staining cells. In some specimens, the positive
staining cells were associated with adjacent areas of
necrosis.

30 In most of the squamous cell carcinomas of the
cervix, the M75 immunoreactivity was focal in distribution,
i.e., only certain areas of the specimen stained. Although
the distribution of positive reactivity within a given
specimen was rather sporadic, the intensity of the reactivity
35 was usually very strong. In most of the adenocarcinomas of
the cervix, the staining pattern was more homogeneous, with
the majority of the specimen staining positively.

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Among the normal tissue samples, intense, positive and specific M75 immunoreactivity was observed only in normal stomach tissues, with diminishing reactivity in the small intestine, appendix and colon. No other normal tissue stained extensively positively for M75. Occasionally, however, foci of intensely staining cells were observed in normal intestine samples (usually at the base of the crypts) or were sometimes seen in morphologically normal appearing areas of the epithelium of cervical specimens exhibiting dysplasia and/or malignancy. In such, normal appearing areas of cervical specimens, positive staining was seen in focal areas of the basal layer of the ectocervical epithelium or in the basal layer of endocervical glandular epithelium. In one normal specimen of human skin, cytoplasmic MN staining was observed in the basal layer. The basal layers of these epithelia are usually areas of proliferation, suggesting the MN expression may be involved in cellular growth. In a few cervical biopsied specimens, MN positivity was observed in the morphologically normal appearing stratified squamous epithelium, sometimes associated with cells undergoing koilocytic changes.

Some colon adenomas (4 of 11) and adenocarcinomas (9 of 15) were positively stained. One normal colon specimen was positive at the base of the crypts. Of 15 colon cancer specimens, 4 adenocarcinomas and 5 metastatic lesions were MN positive. Fewer malignant breast cancers (3 of 25) and ovarian cancer specimens (3 of 15) were positively stained. Of 4 head and neck cancers, 3 stained very intensely with M75.

Although normal stomach tissue was routinely positive, 4 adenocarcinomas of the stomach were MN negative. Of 3 bladder cancer specimens (1 adenocarcinoma, 1 non-papillary transitional cell carcinoma, and 1 squamous cell carcinoma), only the squamous cell carcinoma was MN positive. Approximately 40% (12 of 30) of lung cancer specimens were positive; 2 of 4 undifferentiated carcinomas; 3 of 8 adenocarcinomas; 2 of 8 oat cell carcinomas; and, 5 of 10 squamous cell carcinomas. One hundred percent (4 of 4) of the renal cell carcinomas were MN positive.

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In summary, MN antigen, as detected by M75 and immunohistochemistry in the experiments described above, was shown to be prevalent in tumor cells, most notably in tissues of cervical cancers. MN antigen was also found in some cells of normal tissues, and sometimes in morphologically normal appearing areas of specimens exhibiting dysplasia and/or malignancy. However, MN is not usually extensively expressed in most normal tissues, except for stomach tissues where it is extensively expressed and in the tissues of the lower gastrointestinal tract where it is less extensively expressed. MN expression is most often localized to the cellular plasma membrane of tumor cells and may play a role in intercellular communication or cell adhesion. Representative results of experiments performed as described above are tabulated in Table 2.

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TABLE 2
Immunoreactivity of M75 in Various Tissues

	<u>TISSUE</u>	<u>TYPE</u>	<u>POS/NEG</u> <u>(#pos/#tested)</u>
5	liver, spleen, lung, kidney, adrenal gland, brain, prostate, pancreas, thyroid, ovary, testis	normal	NEG (all)
10	skin	normal	POS (in basal layer) (1/1)
	stomach	normal	POS
	small intestine	normal	POS
	colon	normal	POS
	breast	normal	NEG (0/10)
15	cervix	normal	NEG (0/2)
	breast	benign	NEG (0/17)
	colon	benign	POS (4/11)
	cervix	benign	POS (10/18)
	breast	malignant	POS (3/25)
20	colon	malignant	POS (9/15)
	ovarian	malignant	POS (3/15)
	lung	malignant	POS (12/30)
	bladder	malignant	POS (1/3)
	head & neck	malignant	POS (3/4)
25	kidney	malignant	POS (4/4)
	stomach	malignant	NEG (0/4)
	cervix	malignant	POS (62/68)

The results recorded in this example indicate that the presence of MN proteins in a tissue sample from a patient may, in general, depending upon the tissue involved, be a marker signaling that a pre-neoplastic or neoplastic process is occurring. Thus, one may conclude from these results that diagnostic/prognostic methods that detect MN antigen may be particularly useful for screening patient samples for a number of cancers which can thereby be detected at a pre-neoplastic stage or at an early stage prior to obvious morphologic

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changes associated with dysplasia and/or malignancy being evident or being evident on a widespread basis.

Example 2

Vaccine -- Rat Model

5 As shown above in Example 7 of WO 93/18152 (International Publication Date: 16 September 1993), in some rat tumors, for example, the XC tumor cell line (cells from a rat rhabdomyosarcoma), a rat MN protein, related to human MN, is expressed. Thus a model was afforded to study antitumor
10 immunity induced by experimental MN-based vaccines. The following representative experiments were performed.

Nine- to eleven-day-old Wistar rats from several families were randomized, injected intraperitoneally with 0.1 ml of either control rat sera (the C group) or with rat serum
15 against the MN fusion protein GEX-3X-MN (the IM group). Simultaneously both groups were injected subcutaneously with 10^6 XC tumor cells.

Four weeks later, the rats were sacrificed, and their tumors weighed. The results are shown in Figure 2.
20 Each point on the graph represents a tumor from one rat. The difference between the two groups -- C and IM -- was significant by Mann-Whitney rank test ($U = 84$, $\alpha < 0.025$). The results indicate that the IM group of baby rats developed tumors about one-half the size of the controls, and 5 of the
25 18 passively immunized rats developed no tumor at all, compared to 1 of 18 controls.

Example 3

Expression of Full-Length MN cDNA in NIH 3T3 Cells

The role of MN in the regulation of cell
30 proliferation was studied by expressing the full-length cDNA in NIH 3T3 cells. That cell line was chosen since it had been used successfully to demonstrate the phenotypic effect of a number of proto-oncogenes [Weinberg, R.A., Cancer Res., 49: 3713 (1989); Hunter, T., Cell, 64: 249 (1991)]. Also, NIH
35 3T3 cells express no endogenous MN-related protein that is detectable by Mab M75.

The full length MN cDNA was obtained by ligation of the two cDNA clones using the unique BamHI site and subcloned from pBluescript into KpnI-SacI sites of the expression vector pSG5C. pSG5C was kindly provided by Dr. Richard Kettman

5 [Department of Molecular Biology, Faculty of Agricultural Sciences, B-5030 Gembloux, Belgium]. pSG5C was derived from pSG5 [Stratagene] by inserting a polylinker consisting of a sequence having several neighboring sites for the following restriction enzymes: EcoRI, XhoI, KpnI, BamHI, SacI, 3 times
10 TAG stop codon and BglII.

The recombinant pSG5C-MN plasmid was co-transfected in a 10:1 ratio (10 μ g : 1 μ g) with the pSV2neo plasmid [Southern and Berg, J. Mol. Appl. Genet., 1: 327 (1982)] which contains the neo gene as a selection marker. The co-
15 transfection was carried out by calcium phosphate precipitation method [Mammalian Transfection Kit; Stratagene] into NIH 3T3 cells plated a day before at a density of 1×10^5 per 60 mm dish. As a control, pSV2neo was co-transfected with empty pSG5C.

20 Transfected cells were cultured in DMEM medium supplemented with 10% FCS and 600 μ g ml⁻¹ of G418 [Gibco BRL] for 14 days. The G418-resistant cells were clonally selected, expanded and analysed for expression of the transfected cDNA by Western blotting using iodinated Mab M75.

25 For an estimation of cell proliferation, the clonal cell lines were plated in triplicates (2×10^4 cells/well) in 24-well plates and cultivated in DMEM with 10% FCS and 1% FCS, respectively. The medium was changed each day, and the cell number was counted using a hemacytometer.

30 To determine the DNA synthesis, the cells were plated in triplicate in 96-well plate at a density of 10^4 /well in DMEM with 10% FCS and allowed to attach overnight. Then the cells were labeled with ³H-thymidine for 24 hours, and the incorporated radioactivity was counted.

35 For the anchorage-independent growth assay, cells (2×10^4) were suspended in a 0.3% agar in DMEM containing 10% FCS and overlaid onto 0.5% agar medium in 60 mm dish. Colonies grown in soft agar were counted two weeks after plating.

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Several clonal cell lines constitutively expressing both 54 and 58 kd forms of MN protein in levels comparable to those found in LCMV-infected HeLa cells were obtained.

Selected MN-positive clones and negative control cells (mock-transfected with an empty pSG5C plasmid) were subjected to further analyses directed to the characterization of their phenotype and growth behavior.

The MN-expressing NIH 3T3 cells displayed spindle-shaped morphology, and increased refractility; they were less adherent to the solid support and smaller in size. The control (mock transfected cells) had a flat morphology, similar to parental NIH 3T3 cells. In contrast to the control cells that were aligned and formed a monolayer with an ordered pattern, the cells expressing MN lost the capacity for growth arrest and grew chaotically on top of one another. Correspondingly, the MN-expressing cells were able to reach significantly higher (more than 2x) saturation densities (Table 3) and were less dependent on growth factors than the control cells.

MN transfectants also showed faster doubling times (by 15%) and enhanced DNA synthesis (by 10%), as determined by the amount of [³H]-thymidine incorporated in comparison to control cells. Finally, NIH 3T3 cells expressing MN protein grew in soft agar. The diameter of colonies grown for 14 days ranged from 0.1 to 0.5 mm; however, the cloning efficiency of MN transfectants was rather low (2.4%). Although that parameter of NIH 3T3 cells seems to be less affected by MN than by conventional oncogenes, all other data are consistent with the idea that MN plays a role in cell growth control.

Table 3

Growth Properties of NIH 3T3 Cells Expressing MN Protein

Transfected DNA	pSG5C/ pSV2neo	pSG5C-MN/ pSV2neo
Doubling time ^a (hours)	27.9 ± 0.5	24.1 ± 1.3

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Saturation density ^b (cells x 10 ⁴ /cm ²)	4.9 ± 0.2	11.4 ± 0.4
Cloning efficiency (%) ^c	< 0.01	2.4 ± 0.2

5

^aFor calculation of the doubling time, the proliferation rate of exponentially growing cells was used. ^bThe saturation cell density was derived from the cell number 4 days after reaching confluency. ^cColonies greater than 0.1 mm in diameter were scored at day 14. Cloning efficiency was estimated as a percentage of colonies per number of cells plated, with correction for cell viability.

Example 4

Acceleration of G1 Transit and Decrease in Mitomycin C Sensitivity Caused by MN Protein

15

For the experiments described in this example, the stable MN transfectants of NIH 3T3 cells generated as described in Example 3 were used. Four selected MN-positive clones and four control mock-transfected clones were either used individually or in pools.

Flow cytometric analyses of asynchronous cell populations. Cells that had been grown in dense culture were plated at 1 x 10⁶ cells per 60 mm dish. Four days later, the cells were collected by trypsinization, washed, resuspended in PBS, fixed by dropwise addition of 70% ethanol and stained by propidium iodine solution containing RNase. Analysis was performed by FACStar using DNA cell cycle analysis software [Becton Dickinson; Franklin Lakes, NJ (USA)].

Exponentially growing cells were plated at 5 x 10⁵ cells per 60 mm dish and analysed as above 2 days later. Forward light scatter was used for the analysis of relative cell sizes. The data were evaluated using Kolmogorov-Smirnov test [Young, J. Histochem. Cytochem., 25: 935 (1977)].

The flow cytometric analyses revealed that clonal populations constitutively expressing MN protein showed a

decreased percentage of cells in G1 phase and an increased percentage of cells in G2-M phases. Those differences were more striking in cell populations grown throughout three passages in high density cultures than in exponentially growing subconfluent cells. That observation supports the idea that MN protein has the capacity to perturb contact inhibition.

Also observed was a decrease in the size of MN expressing cells seen in both exponentially proliferating and high density cultures. It is possible that the MN-mediated acceleration of G1 transit is related to the above-noted shorter doubling time (by about 15%) of exponentially proliferating MN-expressing NIH 3T3 cells. Also, MN expressing cells displayed substantially higher saturation density and lower serum requirements than the control cells. Those facts suggest that MN-transfected cells had the capacity to continue to proliferate despite space limitations and diminished levels of serum growth factors, whereas the control cells were arrested in G1 phase.

Limiting conditions. The proliferation of MN-expressing and control cells was studied both in optimal and limiting conditions. Cells were plated at 2×10^4 per well of 24-well plate in DMEM with 10% FCS. The medium was changed at daily intervals until day 4 when confluency was reached, and the medium was no longer renewed. Viable cells were counted in a hemacytometer at appropriate times using trypan blue dye exclusion. The numbers of cells were plotted versus time wherein each plot point represents a mean value of triplicate determination.

The results showed that the proliferation of MN expressing and control cells was similar during the first phase when the medium was renewed daily, but that a big difference in the number of viable cells occurred after the medium was not renewed. More than half of the control cells were not able to withstand the unfavorable growth conditions. In contrast, the MN-expressing cells continued to proliferate even when exposed to increasing competition for nutrients and serum growth factors.

Those results were supported also by flow cytometric analysis of serum starved cells grown for two days in medium containing 1% FCS. While 83% of control cells accumulated in G0-G1 phase (S = 5%, G2-M = 12%), expression of MN protein partially reversed the delay in G1 as indicated by cell cycle distribution of MN tranfectants (G0-G1 = 65%, S = 10%, G2-M = 26%). The results of the above-described experiments suggest that MN protein might function to release the G1/S checkpoint and allow cells to proliferate under unfavorable conditions.

MMC. To test that assumption, unfavorable conditions were simulated by treating cells with the DNA damaging drug mitomycin C (MMC) and then following their proliferation and viability. The mechanism of action of MMC is thought to result from its intracellular activation and subsequent DNA alkylation and crosslinking [Yier and Szybalski, Science, 145: 55 (1964)]. Normally, cells respond to DNA damage by arrest of their cell cycle progression to repair defects and prevent acquisition of genomic instability. Large damage is accompanied by marked cytotoxicity. However, many studies [for example, Peters et al., Int. J. Cancer, 54: 450 (1993)] concern the emergence of drug resistant cells both in tumor cell populations and after the introduction of oncogenes into nontransformed cell lines.

The response of MN-transfected NIH 3T3 cells to increasing concentrations of MMC was determined by continuous [³H]-thymidine labeling. Cells were plated in 96-well microtiter plate concentration of 10⁴ per well and incubated overnight in DMEM with 10% FCS to attach. Then the growth medium was replaced with 100 µl of medium containing increasing concentrations of MMC from 1 µl/ml to 32 µg/ml. All the drug concentrations were tested in three replicate wells. After 5 hours of treatment, the MMC was removed, cells were washed with PBS and fresh growth medium without the drug was added. After overnight recovery, the fractions of cells that were actively participating in proliferation was determined by continuous 24-hr labeling with [³H]-thymidine. The incorporation by the treated cells was compared to that of the control, untreated cells, and the proliferating fractions

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were considered as a percentage of the control's incorporation.

The viability of the treated cells was estimated three days later by a CellTiter 96 AQ Non-Radioactive Cell Proliferation Assay [Promega] which is based on the bio-reduction of methotrexate (MTX) into a water soluble formazan that absorbs light at 490 nm. The percentage of surviving cells was derived from the values of absorbance obtained after subtraction of background.

The control and MN-expressing NIH 3T3 cells showed remarkable differences in their responses to MMC. The sensitivity of the MN-transfected cells appeared considerably lower than the control's in both sections of the above-described experiments. The results suggested that the MN-transfected cells were able to override the negative growth signal mediated by MMC.

ATCC Deposits. The material listed below was deposited with the American Type Culture Collection (ATCC) at 12301 Parklawn Drive, Rockville, MD 20852 (USA). The deposits were made under the provisions of the Budapest Treaty on the International Recognition of Deposited Microorganisms for the Purposes of Patent Procedure and Regulations thereunder (Budapest Treaty). Maintenance of a viable culture is assured for thirty years from the date of deposit. The hybridomas and plasmids will be made available by the ATCC under the terms of the Budapest Treaty, and subject to an agreement between the Applicants and the ATCC which assures unrestricted availability of the deposited hybridomas and plasmids to the public upon the granting of patent from the instant application. Availability of the deposited strain is not to be construed as a license to practice the invention in contravention of the rights granted under the authority of any Government in accordance with its patent laws.

HybridomaDeposit DateATCC #

VU-M75	September 17, 1992	HB 11128
MN 12.2.2	June 9, 1994	HB 11647

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<u>Plasmid</u>	<u>Deposit Date</u>	<u>ATCC #</u>
A4a	June 6, 1995	97199
XE1	June 6, 1995	97200
XE3	June 6, 1995	97198

5 The description of the foregoing embodiments of the
invention have been presented for purposes of illustration and
description. They are not intended to be exhaustive or to
limit the invention to the precise form disclosed, and
obviously many modifications and variations are possible in
10 light of the above teachings. The embodiments were chosen and
described in order to explain the principles of the invention
and its practical application to enable thereby others skilled
in the art to utilize the invention in various embodiments and
with various modifications as are suited to the particular use
15 contemplated.

 All references cited herein are hereby incorporated
by reference.

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CLAIMS

1. An isolated nucleic acid containing at least twenty-seven nucleotides wherein the nucleotide sequence for said nucleic acid is selected from the group consisting of:

5 (a) SEQ. ID. NOS.: 1, 5, and 27-49 and nucleotide sequences complementary to SEQ. ID. NOS.: 1, 5 and 27-49;

(b) nucleotide sequences that hybridize under stringent hybridization conditions to one or more of the following nucleotide sequences: SEQ. ID. NOS.: 1, 5, and 27-
10 49 and the respective complements of SEQ. ID. NOS.: 1, 5 and 27-49; and

(c) nucleotide sequences that differ from the nucleotide sequences of (a) and (b) in codon sequence due to the degeneracy of the genetic code.

15 2. An isolated nucleic acid according to Claim 1 wherein said nucleotide sequence is selected from the group consisting of:

(a) SEQ. ID. NO.: 5 and its complement;

(b) nucleotide sequences that hybridize under
20 stringent conditions to SEQ. ID. NO.: 5 or to its complement;

(c) nucleotide sequences that differ from the nucleotide sequences of (a) or (b) in codon sequence due to the degeneracy of the genetic code.

25 3. An isolated nucleic acid containing at least sixteen nucleotides wherein the nucleotide sequence therefor is selected from the group consisting of:

(a) the MN nucleotide sequences contained in plasmids A4a, XE1 and XE3 which were deposited at the American Type Culture Collection (ATCC) in Rockville, Maryland in the
30 United State of America under the respective ATCC Nos. 97199, 97200, and 97198;

(b) nucleotide sequences that hybridize under stringent conditions to the MN nucleotide sequences of (a);
and

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(c) nucleotide sequences that differ from the nucleotide sequences of (a) or (b) in codon sequence due to the degeneracy of the genetic code.

4. An isolated nucleic acid according to Claim 3
5 which functions as a polymerase chain reaction primer for MN nucleic acid sequences, and which is from 16 to about 50 nucleotides in length.

5. An isolated nucleic acid, containing at least fifty nucleotides, encoding an MN protein or polypeptide that
10 is specifically bound either by monoclonal antibodies designated M75 secreted by the hybridoma VU-M75 deposited at the American Type Culture Collection (ATCC) in Rockville, Maryland in the United States of America under ATCC No. HB 11128, or by monoclonal antibodies designated MN12 secreted by
15 the hybridoma MN 12.2.2 deposited at the ATCC under ATCC No. 11647, or by both of said monoclonal antibodies.

6. An isolated nucleic acid which is operatively linked to an expression control sequence within a vector wherein said nucleic acid is selected from the group
20 consisting of:

(a) SEQ. ID. NO.: 1 and its complement;

(b) nucleic acids that hybridize under stringent hybridization conditions to SEQ. ID. NO.: 1 or to its complement; and

25 (c) nucleic acids that differ from the nucleic acid sequences of (a) and (b) due to the degeneracy of the genetic code.

7. A unicellular host, which is either prokaryotic or eukaryotic, that is transformed or transfected with the
30 isolated nucleic acid operatively linked to an expression control sequence in a vector according to Claim 6.

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8. A method of recombinantly producing an MN protein, MN fusion protein or MN polypeptide comprising the steps of:

- 5 (a) transforming a unicellular host with the isolated nucleic acid operatively linked to an expression control sequence in a vector according to Claim 6;
- (b) culturing said unicellular host so that said MN protein or polypeptide is expressed; and
- 10 (c) extracting and isolating said MN protein or polypeptide.

9. A recombinant nucleic acid encoding a fusion protein, that consists essentially of an MN protein or polypeptide and a non-MN protein or polypeptide, wherein the nucleotide sequence for the portion of the nucleic acid
15 encoding the MN protein or polypeptide is selected from the group consisting of:

- (a) SEQ. ID. NO.: 1;
 - (b) nucleotide sequences that hybridize under stringent conditions to SEQ. ID. NO.: 1 or to its complement;
20 and
 - (c) degenerate variants of SEQ. ID. NO.: 1 and of the nucleotide sequences of (b);
- wherein the nucleic acid encoding said MN protein or polypeptide contains at least twenty-nine nucleotides.

25 10. A method of detecting mutations in an isolated MN gene and/or fragment(s) thereof comprising the steps of:
amplifying one or more fragment(s) of said gene by the polymerase chain reaction (PCR); and
determining whether said one or more fragments
30 contain any mutations.

11. An MN protein, MN fusion protein or MN polypeptide, wherein said MN protein, MN polypeptide or the MN protein portion of said MN fusion protein is encoded by a nucleic acid of at least twenty-nine nucleotides which is
35 selected from the group consisting of:

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(a) SEQ. ID. NO. NO.: 1;

(b) nucleotide sequences that hybridize under stringent hybridization conditions to SEQ. ID. NO.: 1 or to its complement; and

5 (c) nucleotide sequences that differ from SEQ. ID. NO.: 1 or from the sequences of (b) in codon sequence due to the degeneracy of the genetic code.

10 12. An MN protein, MN fusion protein, or MN polypeptide wherein said MN protein or polypeptide has, and wherein said MN fusion protein contains, an amino acid sequence selected from the group consisting of SEQ. ID. NOS.: 2, 6, 10-16, and 50-53.

15 13. A vaccine comprising an immunogenic amount of one or more MN proteins, MN fusion proteins, and/or MN polypeptides according to Claims 11 or 12 dispersed in a physiologically acceptable, nontoxic vehicle, which amount is effective to immunize a vertebrate against a neoplastic disease associated with expression of MN antigen.

20 14. An antibody which specifically binds to an MN protein, an MN fusion protein and/or an MN polypeptide according to Claims 11 or 12.

25 15. A monoclonal antibody according to Claim 14 which is designated MN12 and is secreted by the hybridoma MN 12.2.2 which was deposited at the American Type Culture Collection (ATCC) in Rockville, Maryland in the United States of America under ATCC No. HB 11647.

30 16. An antibody according to Claim 14 which specifically binds to an MN antigen epitope selected from the group of epitopes represented by the following amino acid sequences: SEQ. ID. NOS. 10-16.

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17. An antibody according to Claim 14 which is linked to an imaging agent, to a chemotherapeutic agent or to a toxic agent.

18. A method of imaging pre-neoplastic or
5 neoplastic disease in a patient comprising:

(a) injecting said patient with antibody linked to an imaging agent according to Claim 17; and

(b) detecting the binding of said antibody.

19. A hybridoma designated MN 12.2.2 which produces
10 the monoclonal antibody MN12, and which was deposited at the American Type Culture Collection (ATCC) in Rockville Maryland in the United States of America under ATCC Accession No. HB 11647.

20. A method of delivering a chemotherapeutic agent
15 or toxic agent to a cancer cell which comprises contacting said cell with an antibody linked to a chemotherapeutic agent or to a toxic agent according to Claim 17.

21. A method of treating neoplastic disease in a patient comprising administering to said patient a
20 therapeutically effective amount of a composition comprising antibodies which specifically bind to an MN protein, an MN fusion protein and/or an MN polypeptide according to Claims 11 or 12.

22. A method of detecting and/or quantitating in a
25 vertebrate sample MN antigen comprising the steps of:

(a) contacting said sample with one or more antibodies according to Claims 14, 15 or 16; and

(b) detecting and/or quantitating binding of said antibody in said sample.

30 23. A method according to Claim 22 wherein said vertebrate sample is a human tissue specimen, such as, a cell

smear, a histological section from a biopsied tissue or organ, or an imprint preparation.

24. A method according to Claim 23 wherein said tissue specimen is ovarian, endometrial, or cervical.

5 25. A method of detecting and/or quantitating MN-specific antibodies in a vertebrate sample comprising the steps of:

(a) contacting and incubating the vertebrate sample with an MN protein, an MN fusion protein and/or an MN polypeptide according to Claims 11 or 12; and

10 (b) detecting and/or quantitating binding of said MN protein, MN fusion protein and/or MN polypeptide to antibody in said sample.

15 26. A method of treating neoplastic disease and/or pre-neoplastic disease comprising inhibiting the expression of MN genes by administering one or more antisense nucleic acid sequences that hybridize under stringent conditions to mRNA transcribed from MN genes.

20 27. Vectors containing an MN nucleic acid sequence wherein said MN nucleic acid sequence is selected from the group consisting of:

(a) SEQ. ID. NO.: 5 and its complement;

(b) nucleic acids that hybridize to SEQ. ID. NO.: 5 or to its complement; and

25 (c) nucleic acids that differ from the nucleic acids of (a) or (b) due to the degeneracy of the genetic code;

wherein said nucleic acid is at least twenty-nine nucleotides in length.

30 28. Vectors containing an MN nucleic acid sequence according to Claim 27 selected from the group consisting of the plasmids A4a, XE1 and XE3 which are deposited at the American Type Culture Collection (ATCC) in Rockville, Maryland

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in the United States of America under the respective ATCC accession numbers 97199, 97200 and 97198.

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1	1	ACA	GTC	AGC	CGC	ATG	GCT	CCC	CTG	TGC	CCC	AGC	CCC	TGG	CTC	CCT	CTG	12	48
13	49	L	I	P	A	P	A	P	G	L	T	V	Q	L	L	L	S	28	96
29	97	L	L	L	L	M	P	P	V	H	P	Q	R	L	P	R	Q	44	144
45	145	E	D	S	P	L	G	G	G	S	S	G	E	D	D	P	L	60	192
61	193	G	E	E	D	L	P	S	E	E	D	S	P	R	E	E	D	76	240
77	241	P	P	G	E	E	D	L	P	G	E	E	D	L	P	G	E	92	288
93	289	E	D	L	P	E	V	K	P	K	S	E	E	E	G	S	L	108	336
109	337	K	L	E	D	L	P	T	V	E	A	P	G	D	P	Q	E	124	384
125	385	P	Q	N	N	A	H	R	D	K	E	G	D	D	Q	S	H	140	432
141	433	W	R	Y	G	G	D	P	P	W	P	R	V	S	P	A	C	156	480
157	481	A	G	R	F	Q	S	P	V	D	I	R	P	Q	L	A	A	172	528

FIG.-1A

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173	F	C	P	A	L	R	P	L	E	L	L	G	F	Q	L	P	188
529	TTC	TGC	CCG	GCC	CTG	CGC	CCC	CTG	GAA	CTC	CTG	GGC	TTC	CAG	CTC	CCG	576
189	P	L	P	E	L	R	L	R	N	N	G	H	S	V	Q	L	204
577	CCG	CTC	CCA	GAA	CTG	CGC	CTG	CGC	AAC	AAT	GGC	CAC	AGT	GTG	CAA	CTG	624
205	T	L	P	P	G	L	E	M	A	L	G	P	G	R	E	Y	220
625	ACC	CTG	CCT	CCT	GGG	CTA	GAG	ATG	GCT	CTG	GGT	CCC	GGG	CGG	GAG	TAC	672
221	R	A	L	Q	L	H	L	H	W	G	A	A	G	R	P	G	236
673	CGG	GCT	CTG	CAG	CTG	CAT	CTG	CAC	TGG	GGG	GCT	GCA	GGT	CGT	CCG	GGC	720
237	S	E	H	T	V	E	G	H	R	F	P	A	E	I	H	V	252
721	TCG	GAG	CAC	ACT	GTG	GAA	GGC	CAC	CGT	TTC	CCT	GCC	GAG	ATC	CAC	GTG	768
253	V	H	L	S	T	A	F	A	R	V	D	E	A	L	G	R	268
769	GTT	CAC	CTC	AGC	ACC	GCC	TTT	GCC	AGA	GTT	GAC	GAG	GCC	TTG	GGG	CGC	816
269	P	G	G	L	A	V	L	A	A	F	L	E	E	G	P	E	284
817	CCG	GGA	GGC	CTG	GCC	GTG	TTG	GCC	GCC	TTT	CTG	GAG	GAG	GGC	CCG	GAA	864
285	E	N	S	A	Y	E	Q	L	L	S	R	L	E	E	I	A	300
865	GAA	AAC	AGT	GCC	TAT	GAG	CAG	TTG	CTG	TCT	CGC	TTG	GAA	GAA	ATC	GCT	912
301	E	E	G	S	E	T	Q	V	P	G	L	D	I	S	A	L	316
913	GAG	GAA	GGC	TCA	GAG	ACT	CAG	GTC	CCA	GGA	CTG	GAC	ATA	TCT	GCA	CTC	960
317	L	P	S	D	F	S	R	Y	F	Q	Y	E	G	S	L	T	332
961	CTG	CCC	TCT	GAC	TTC	AGC	CGC	TAC	TTC	CAA	TAT	GAG	GGG	TCT	CTG	ACT	1008
333	T	P	P	C	A	Q	G	V	I	W	T	V	F	N	Q	T	348
1009	ACA	CCG	CCC	TGT	GCC	CAG	GGT	GTC	ATC	TGG	ACT	GTG	TTT	AAC	CAG	ACA	1056

FIG.-1B

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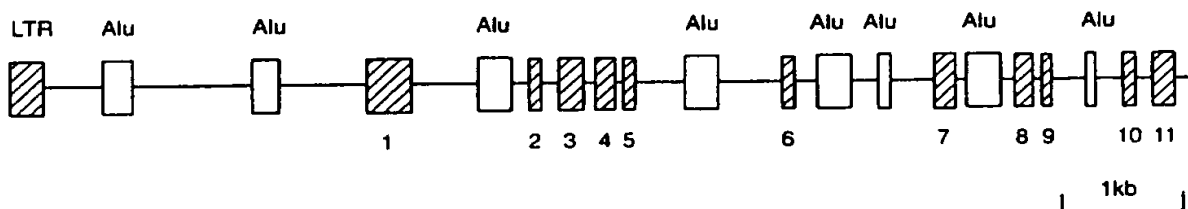
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(57) Abstract

A complete genomic sequence including a full-length cDNA sequence for the MN gene, a putative oncogene, is disclosed, as well as proteins/polypeptides encoded thereby. Recombinant nucleic acid molecules for expressing MN proteins/polypeptides and recombinant proteins are also provided. Expression of the MN gene is disclosed as being associated with tumorigenicity, and the invention concerns methods and compositions for detecting and/or quantitating MN antigen and/or MN-specific antibodies in vertebrate samples that are diagnostic/prognostic for neoplastic and preneoplastic disease. Test kits embodying the immunoassays of this invention are provided. MN-specific antibodies are disclosed that can be used diagnostically/prognostically, therapeutically, for imaging, and/or for affinity purification of MN proteins/polypeptides. Also provided are nucleic acid probes for the MN gene as well as test kits comprising said probes. The invention also concerns vaccines comprising MN proteins/polypeptides which are effective to immunize a vertebrate against neoplastic diseases associated with the expression of MN proteins. The invention still further concerns antisense nucleic acid sequences that can be used to inhibit MN gene expression, and polymerase chain reaction (PCR) assays to detect genetic rearrangements in MN genes.

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MN GENE AND PROTEINFIELD OF THE INVENTION

The present invention is in the general area of medical genetics and in the fields of biochemical engineering and immunochemistry. More specifically, it relates to the identification of a new gene--the MN gene--a cellular gene coding for the MN protein. The inventors hereof found MN proteins to be associated with tumorigenicity. Evidence indicates that the MN protein appears to represent a potentially novel type of oncoprotein. Identification of MN antigen as well as antibodies specific therefor in patient samples provides the basis for diagnostic/prognostic assays for cancer.

BACKGROUND OF THE INVENTION

A novel quasi-viral agent having rather unusual properties was detected by its capacity to complement mutants of vesicular stomatitis virus (VSV) with heat-labile surface G protein in HeLa cells (cell line derived from human cervical adenocarcinoma), which had been cocultivated with human breast carcinoma cells. [Zavada et al., Nature New Biol., 240: 124 (1972); Zavada et al., J. Gen. Virol., 24: 327 (1974); Zavada, J., Arch. Virol., 50: 1 (1976); Zavada, J., J. Gen. Virol., 63: 15-24 (1982); Zavada and Zavadova, Arch. Virol., 118: 189 (1991).] The quasi viral agent was called MaTu as it was presumably derived from a human mammary tumor.

There was significant medical interest in studying and characterizing MaTu as it appeared to be an entirely new type of molecular parasite of living cells, and possibly originated from a human tumor. Zavada et al., International Publication Number WO 93/18152 (published 1 September 1993), describes the elucidation of the biological and molecular nature of MaTu which resulted in the discovery of the MN gene and protein. MaTu was found by the inventors to be a two-

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component system, having an exogenous transmissible component, MX, and an endogenous cellular component, MN. The MN component was found to be a cellular gene, showing only very little homology with known DNA sequences. The MN gene was
5 found to be present in the chromosomal DNA of all vertebrates tested, and its expression was found to be strongly correlated with tumorigenicity.

The exogenous MaTu-MX transmissible agent was identified as lymphocytic choriomeningitis virus (LCMV) which
10 persistently infects HeLa cells. The inventors discovered that the MN expression in HeLa cells is positively regulated by cell density, and also its expression level is increased by persistent infection with LCMV.

Research results provided herein show that cells
15 transfected with MN cDNA undergo changes indicative of malignant transformation. Further research findings indicate that the disruption of cell cycle control is one of the mechanisms by which MN may contribute to the complex process of tumor development.

Described herein is the cloning and sequencing of
20 the MN gene and the recombinant production of MN proteins. The full-length MN cDNA sequence [SEQ. ID. NO.: 1], the amino acid sequence deduced therefrom [SEQ. ID. NO.: 2], a full-length genomic sequence for MN [SEQ. ID. NO.: 5] including a
25 proposed promoter sequence [SEQ. ID. NO.: 27] are provided. Eleven exons [SEQ. ID. NOS. 28-38] and ten introns [SEQ. ID. NOS.: 39-48] are comprised by the MN gene. Also a 1.4 kilobase region [SEQ. ID. NO. 49] within the middle of the MN genomic sequence is described herein, which has the character
30 of a typical CpG-rich island, and which contains multiple putative binding sites for transcription factors AP2 and Sp1.

Also described are antibodies prepared against proteins/polypeptides. MN proteins/ polypeptides can be used in serological assays according to this invention to detect
35 MN-specific antibodies. Further, MN proteins/polypeptides and/or antibodies reactive with MN antigen can be used in immunoassays according to this invention to detect and/or

quantitate MN antigen. Such assays may be diagnostic and/or prognostic for neoplastic/pre-neoplastic disease.

SUMMARY OF THE INVENTION

This invention is directed to the MN gene, fragments thereof and the related cDNA which are useful, for example, as follows: 1) to produce MN proteins/ polypeptides by biochemical engineering; 2) to prepare nucleic acid probes to test for the presence of the MN gene in cells of a subject; 3) to prepare appropriate polymerase chain reaction (PCR) primers for use, for example, in PCR-based assays or to produce nucleic acid probes; 4) to identify MN proteins and polypeptides as well as homologs or near homologs thereto; 5) to identify various mRNAs transcribed from MN genes in various tissues and cell lines, preferably human; and 6) to identify mutations in MN genes. The invention further concerns purified and isolated DNA molecules comprising the MN gene or fragments thereof, or the related cDNA or fragments thereof.

Thus, this invention in one aspect concerns isolated nucleic acid sequences that encode MN proteins or polypeptides wherein the nucleotide sequences for said nucleic acids are selected from the group consisting of:

- (a) SEQ. ID. NO.: 1;
 - (b) nucleotide sequences that hybridize under stringent conditions to SEQ. ID. NO.: 1 or to its complement;
 - (c) nucleotide sequences that differ from SEQ. ID. NO.: 1 or from the nucleotide sequences of (b) in codon sequence because of the degeneracy of the genetic code.
- Further, such nucleic acid sequences are selected from nucleotide sequences that but for the degeneracy of the genetic code would hybridize to SEQ. ID. NO.: 1 or to its complement under stringent hybridization conditions.

Further, such isolated nucleic acids that encode MN proteins or polypeptides can also include the MN nucleic acids of the genomic sequence shown in Figure 3a-d, that is, SEQ. ID. NO.: 5, as well as sequences that hybridize to it or its complement under stringent conditions, or would hybridize to SEQ. ID. NO.: 5 or to its complement under such conditions,

but for the degeneracy of the genetic code. Degenerate variants of SEQ. ID. NOS.: 1 and 5 are within the scope of the invention.

Further, this invention concerns nucleic acid probes which are fragments of the isolated nucleic acids that encode MN proteins or polypeptides as described above. Preferably said nucleic acid probes are comprised of at least 29 nucleotides, more preferably of at least 50 nucleotides, still more preferably at least 100 nucleotides, and even more preferably at least 150 nucleotides.

Still further, this invention is directed to isolated nucleic acids containing at least twenty-seven nucleotides selected from the group consisting of:

(a) SEQ. ID. NOS.: 1, 5 and 27-49 and that are complementary to SEQ. ID. NOS.: 1, 5 and 27-49;

(b) nucleotide sequences that hybridize under standard stringent hybridization conditions to one or more of the following nucleotide sequences: SEQ. ID. NOS.: 1, 5, and 27-49 and the respective complements of SEQ. ID. NOS.: 1, 5 and 27-49; and

(c) nucleotide sequences that differ from the nucleotide sequences of (a) and (b) in codon sequence because of the degeneracy of the genetic code. The invention also concerns nucleic acids that but for the degeneracy of the genetic code would hybridize to the nucleic acids of (a) and (b) under standard stringent hybridization conditions. Further this invention concerns nucleic acids of (b) and (c) that hybridize partially or wholly to the non-coding regions of SEQ. ID. NO.: 5 or its complement as, for example, sequences that function as nucleic acid probes to identify MN nucleic acid sequences. Conventional technology can be used to determine whether the nucleic acids of (b) and (c) or of fragments of SEQ. ID. NO.: 5 are useful to identify MN nucleic acid sequences, for example, as outlined in Benton and Davis, Science, 196: 180 (1977) and Fuscoe et al. Genomics, 5: 100 (1989). In general, such nucleic acids are preferably at least 29 nucleotides, most preferably at least 50 nucleotides and still more preferably at least 100

nucleotides. An exemplary and preferred nucleic acid probe is SEQ. ID. NO.: 55 (a 470 bp probe useful in RNase protection assays).

Test kits of this invention can comprise the nucleic acid probes of the invention which are useful diagnostically/prognostically for neoplastic and/or pre-neoplastic disease. Preferred test kits comprise means for detecting or measuring the hybridization of said probes to the MN gene or to the mRNA product of the MN gene, such as a visualizing means.

Fragments of the isolated nucleic acids of the invention, can also be used as PCR primers to amplify segments of MN genes, and may be useful in identifying mutations in MN genes. Typically, said PCR primers are oligonucleotides, preferably at least 16 nucleotides, but they may be considerably longer. Exemplary primers may be from about 16 nucleotides to about 50 nucleotides, preferably from about 19 nucleotides to about 45 nucleotides.

Further, the invention concerns the use of such PCR primers in methods to detect mutations in an isolated MN gene and/or fragment(s) thereof. For example, such methods can comprise amplifying one or more fragment(s) of an MN gene by PCR, and determining whether any of said one or more fragments contain mutations, by, for example, comparing the size of the amplified fragments to those of similarly amplified corresponding fragments of MN genes known to be normal, by using a PCR-single-strand conformation polymorphism assay or a denaturing gradient gel electrophoretic assay.

This invention also concerns nucleic acids which encode MN proteins or polypeptides that are specifically bound by monoclonal antibodies designated M75 that are produced by the hybridoma VU-M75 deposited at the American Type Culture Collection (ATCC) at 12301 Parklawn Drive in Rockville, Maryland 20852 (USA) under ATCC No. HB 11128, and/or by monoclonal antibodies designated MN12 produced by the hybridoma MN 12.2.2 deposited at the ATCC under ATCC No. HB 11647.

This invention further concerns isolated nucleic acids containing at least sixteen nucleotides, preferably at least twenty-nine nucleotides, more preferably at least fifty nucleotides, wherein said nucleic acid is selected from the group consisting of:

(a) the MN nucleic acids contained in plasmids A4a, XE1 and XE3 which were deposited at the American Type Culture Collection (ATCC) in Rockville, Maryland in the United States of America under the respective ATCC Nos. 97199, 97200, and 97198;

(b) nucleic acids that hybridize under stringent conditions to the MN nucleic acids of (a); and

(c) nucleic acids that differ from the nucleic acids of (a) or (b) in codon sequence due to the degeneracy of the genetic code. Such isolated nucleic acids, for example, can be polymerase chain reaction (PCR) primers.

The invention further concerns isolated nucleic acids that code for an MN protein, MN fusion protein or MN polypeptide that is operatively linked to an expression control sequence within a vector; unicellular hosts, prokaryotic or eukaryotic, that are transformed or transfected therewith; and methods of recombinantly producing MN proteins, MN fusion proteins and MN polypeptides comprising transforming or transfecting unicellular hosts with said nucleic acid operatively linked to an expression control sequence, culturing said transformed or transfected unicellular hosts so that said MN proteins, fusion proteins or polypeptides are expressed, and extracting and isolating said MN protein fusion protein or polypeptide.

Recombinant nucleic acids that encode MN fusion proteins are claimed as consisting essentially of an MN protein or MN polypeptide and a non-MN protein or polypeptide wherein the nucleotide sequence for the portion of the nucleic acid encoding the MN protein or polypeptide is selected from the group consisting of:

(a) SEQ. ID. NO.: 1;

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(b) nucleotide sequences that hybridize under stringent conditions to SEQ. ID. NO.: 1 or to its complement; and

5 (c) degenerate variants of SEQ. ID. NO.: 1, and of the nucleotide sequences of (b);

wherein the nucleic acid encoding said MN protein or polypeptide contains at least twenty-nine nucleotides.

Said non-MN protein or polypeptide may preferably be nonimmunogenic to humans and not typically reactive to
10 antibodies in human body fluids. Examples of such a DNA sequence is the alpha-peptide coding region of beta-galactosidase and a sequence coding for glutathione S-transferase or a fragment thereof. However, in some instances, a non-MN protein or polypeptide that is
15 serologically active, immunogenic and/or antigenic may be preferred as a fusion partner to a MN antigen. Further, claimed herein are such recombinant fusion proteins/polypeptides which are substantially pure and non-naturally occurring. Exemplary fusion proteins of this invention are
20 GEX-3X-MN, MN-Fc and MN-PA, described infra.

In HeLa and in tumorigenic HeLa x fibroblast hybrid (H/F-T) cells, MN protein is manifested as a "twin" protein p54/58N; it is glycosylated and forms disulfide-linked oligomers. As determined by electrophoresis upon reducing
25 gels, MN proteins have molecular weights in the range of from about 40 kd to about 70 kd, preferably from about 45 kd to about 65 kd, more preferably from about 48 kd to about 58 kd. Upon non-reducing gels, MN proteins in the form of oligomers have molecular weights in the range of from about 145 kd to
30 about 160 kd, preferably from about 150 to about 155 kd, still more preferably from about 152 to about 154 kd. A predicted amino acid sequence for a preferred MN protein of this invention is shown in Figure 1 [SEQ. ID. NO. 2].

Other particular MN proteins or polypeptides are
35 exemplified by the putative MN signal peptide shown as the first thirty-seven amino acids in Figure 1 [SEQ. ID. NO.: 6], preferred MN antigen epitopes [SEQ. ID. NOS.: 10-16], and domains of the MN protein represented in Figure 1 as amino

acids 38-135 [SEQ. IS. NO.: 50], 136-391 [SEQ. ID. NO.: 51], 414-433 [SEQ. ID. NO.: 52], and 434-459 [SEQ. ID. NO.: 53].

The discovery of the MN gene and protein and thus, of substantially complementary MN genes and proteins encoded thereby, led to the finding that the expression of MN proteins was associated with tumorigenicity. That finding resulted in the creation of methods that are diagnostic/ prognostic for cancer and precancerous conditions. Methods and compositions are provided for identifying the onset and presence of neoplastic disease by detecting and/or quantitating MN antigen in patient samples, including tissue sections and smears, cell and tissue extracts from vertebrates, preferably mammals and more preferably humans. Such MN antigen may also be found in body fluids.

MN proteins and genes are of use in research concerning the molecular mechanisms of oncogenesis, in cancer diagnostics/prognostics, and may be of use in cancer immunotherapy. The present invention is useful for detecting a wide variety of neoplastic and/or pre-neoplastic diseases. Exemplary neoplastic diseases include carcinomas, such as mammary, bladder, ovarian, uterine, cervical, endometrial, squamous cell and adenosquamous carcinomas; and head and neck cancers; mesodermal tumors, such as neuroblastomas and retinoblastomas; sarcomas, such as osteosarcomas and Ewing's sarcoma; and melanomas. Of particular interest are head and neck cancers, gynecologic cancers including ovarian, cervical, vaginal, endometrial and vulval cancers; gastrointestinal cancer, such as, stomach, colon and esophageal cancers; urinary tract cancer, such as, bladder and kidney cancers; skin cancer; liver cancer; prostate cancer; lung cancer; and breast cancer. Of still further particular interest are gynecologic cancers; breast cancer; urinary tract cancers, especially bladder cancer; lung cancer; and liver cancer. Even further of particular interest are gynecologic cancers and breast cancer. Gynecologic cancers of particular interest are carcinomas of the uterine cervix, endometrium and ovaries; more particularly such gynecologic cancers include cervical squamous cell carcinomas, adenosquamous carcinomas,

adenocarcinomas as well as gynecologic precancerous conditions, such as metaplastic cervical tissues and condylomas.

The invention further relates to the biochemical engineering of the MN gene, fragments thereof or related cDNA. For example, said gene or a fragment thereof or related cDNA can be inserted into a suitable expression vector, wherein it is operatively linked to an expression control sequence; host cells, preferably unicellular, can be transformed or transfected with such an expression vector; and an MN protein/polypeptide, preferably an MN protein, is expressed therein. Such a recombinant protein or polypeptide can be glycosylated or nonglycosylated, preferably glycosylated, and can be purified to substantial purity. The invention further concerns MN proteins/polypeptides which are synthetically or otherwise biologically prepared.

Said MN proteins/polypeptides can be used in assays to detect MN antigen in patient samples and in serological assays to test for MN-specific antibodies. MN proteins/polypeptides of this invention are serologically active, immunogenic and/or antigenic. They can further be used as immunogens to produce MN-specific antibodies, polyclonal and/or monoclonal, as well as an immune T-cell response.

The invention further is directed to MN-specific antibodies, which can be used diagnostically/prognostically and may be used therapeutically. Preferred according to this invention are MN-specific antibodies reactive with the epitopes represented respectively by the amino acid sequences of the MN protein shown in Figure 1 as follows: from AA 62 to AA 67 [SEQ. ID. NO.: 10]; from AA 55 to AA 60 [SEQ. ID. NO.: 11]; from AA 127 to AA 147 [SEQ. ID. NO.: 12]; from AA 36 to AA 51 [SEQ. ID. NO.: 13]; from AA 68 to AA 91 [SEQ. ID. NO.: 14]; from AA 279 to AA 291 [SEQ. ID. NO.: 15]; and from AA 435 to AA 450 [SEQ. ID. NO.: 16]. More preferred are antibodies reactive with epitopes represented by SEQ. ID. NOS.: 10, 11 and 12. Still more preferred are antibodies reactive with the epitopes represented by SEQ. ID NOS: 10 and

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11, as for example, respectively Mabs M75 and MN12. Most preferred are monoclonal antibodies reactive with the epitope represented by SEQ. ID. NO.: 10.

Also preferred according to this invention are
5 antibodies prepared against recombinantly produced MN proteins as, for example, GEX-3X-MN, MN 20-19, MN-Fc and MN-PA. Also preferred are MN-specific antibodies prepared against glycosylated MN proteins, such as, MN 20-19 expressed in baculovirus infected Sf9 cells.

10 A hybridoma that produces a representative MN-specific antibody, the monoclonal antibody M75 (Mab M75), was deposited at the under ATCC Number HB 11128 as indicated above. The M75 antibody was used to discover and identify the MN protein and can be used to identify readily MN antigen in
15 Western blots, in radioimmunoassays and immunohistochemically, for example, in tissue samples that are fresh, frozen, or formalin-, alcohol-, acetone- or otherwise fixed and/or paraffin-embedded and deparaffinized. Another representative MN-specific antibody, Mab MN12, is secreted by the hybridoma
20 MN 12.2.2, which was deposited at the ATCC under the designation HB 11647.

MN-specific antibodies can be used, for example, in laboratory diagnostics, using immunofluorescence microscopy or immunohistochemical staining; as a component in immunoassays
25 for detecting and/or quantitating MN antigen in, for example, clinical samples; as probes for immunoblotting to detect MN antigen; in immunoelectron microscopy with colloid gold beads for localization of MN proteins and/or polypeptides in cells; and in genetic engineering for cloning the MN gene or
30 fragments thereof, or related cDNA. Such MN-specific antibodies can be used as components of diagnostic/prognostic kits, for example, for in vitro use on histological sections; such antibodies can also and used for in vivo diagnostics/prognostics, for example, such antibodies can be labeled
35 appropriately, as with a suitable radioactive isotope, and used in vivo to locate metastases by scintigraphy. Further such antibodies may be used in vivo therapeutically to treat cancer patients with or without toxic and/or cytostatic agents

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attached thereto. Further, such antibodies can be used in vivo to detect the presence of neoplastic and/or pre-neoplastic disease. Still further, such antibodies can be used to affinity purify MN proteins and polypeptides.

5 This invention also concerns methods of treating neoplastic disease and/or pre-neoplastic disease comprising inhibiting the expression of MN genes by administering antisense nucleic acid sequences that are substantially complementary to mRNA transcribed from MN genes. Said
10 antisense nucleic acid sequences are those that hybridize to such mRNA under stringent hybridization conditions. Preferred are antisense nucleic acid sequences that are substantially complementary to sequences at the 5' end of the MN cDNA sequence shown in Figure 1. Preferably said antisense nucleic
15 acid sequences are oligonucleotides.

This invention also concerns vaccines comprising an immunogenic amount of one or more substantially pure MN proteins and/or polypeptides dispersed in a physiologically acceptable, nontoxic vehicle, which amount is effective to
20 immunize a vertebrate, preferably a mammal, more preferably a human, against a neoplastic disease associated with the expression of MN proteins. Said proteins can be recombinantly, synthetically or otherwise biologically produced. A particular use of said vaccine would be to
25 prevent recidivism and/or metastasis. For example, it could be administered to a patient who has had an MN-carrying tumor surgically removed, to prevent recurrence of the tumor.

The immunoassays of this invention can be embodied in test kits which comprise MN proteins/polypeptides and/or
30 MN-specific antibodies. Such test kits can be in solid phase formats, but are not limited thereto, and can also be in liquid phase format, and can be based on immunohistochemical assays, ELISAS, particle assays, radiometric or fluorometric assays either unamplified or amplified, using, for example,
35 avidin/biotin technology.

Abbreviations

The following abbreviations are used herein:

-12-

	AA	-	amino acid
	ATCC	-	American Type Culture Collection
	bp	-	base pairs
	BLV	-	bovine leukemia virus
5	BSA	-	bovine serum albumin
	BRL	-	Bethesda Research Laboratories
	CA	-	carbonic anhydrase
	CAT	-	chloramphenicol acetyltransferase
	Ci	-	curie
10	cm	-	centimeter
	CMV	-	cytomegalovirus
	cpm	-	counts per minute
	C-terminus	-	carboxyl-terminus
	°C	-	degrees centigrade
15	DEAE	-	diethylaminoethyl
	DMEM	-	Dulbecco modified Eagle medium
	EDTA	-	ethylenediaminetetraacetate
	EIA	-	enzyme immunoassay
	ELISA	-	enzyme-linked immunosorbent assay
20	F	-	fibroblasts
	FCS	-	fetal calf serum
	FITC	-	fluorescein isothiocyanate
	GEX-3X-MN	-	fusion protein MN glutathione S-transferase
	H	-	HeLa cells
25	HEF	-	human embryo fibroblasts
	HeLa K	-	standard type of HeLa cells
	HeLa S	-	Stanbridge's mutant HeLa D98/AH.2
	H/F-T	-	hybrid HeLa fibroblast cells that are tumorigenic; derived from HeLa D98/AH.2
30	H/F-N	-	hybrid HeLa fibroblast cells that are nontumorigenic; derived from HeLa D98/AH.2
	HRP	-	horseradish peroxidase
	Inr	-	initiator
	IPTG	-	isopropyl-Beta-D-thiogalacto-pyranoside
35	kb	-	kilobase
	kbp	-	kilobase pairs
	kd	-	kilodaltons
	LCMV	-	lymphocytic choriomeningitis virus

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	LTR	-	long terminal repeat
	M	-	molar
	mA	-	milliamperere
	MAB	-	monoclonal antibody
5	ME	-	mercaptoethanol
	MEM	-	minimal essential medium
	min.	-	minute(s)
	mg	-	milligram
	ml	-	milliliter
10	mM	-	millimolar
	MMC	-	mitomycin C
	MLV	-	murine leukemia virus
	N	-	normal concentration
	NEG	-	negative
15	ng	-	nanogram
	nt	-	nucleotide
	N-terminus	-	amino-terminus
	ODN	-	oligodeoxynucleotide
	ORF	-	open reading frame
20	PA	-	Protein A
	PBS	-	phosphate buffered saline
	PCR	-	polymerase chain reaction
	PEST	-	combination of one-letter abbreviations for proline, glutamic acid, serine, threonine
25	pI	-	isoelectric point
	PMA	-	phorbol 12-myristate 13-acetate
	POS	-	positive
	Py	-	pyrimidine
	RIA	-	radioimmunoassay
30	RIP	-	radioimmunoprecipitation
	RIPA	-	radioimmunoprecipitation assay
	RNP	-	RNase protection assay
	SDRE	-	serum dose response element
	SDS	-	sodium dodecyl sulfate
35	SDS-PAGE	-	sodium dodecyl sulfate-polyacrylamide gel electrophoresis
	SINE	-	short interspersed repeated sequence
	SSDF	-	synthetic splice donor site

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	SP-RIA	-	solid-phase radioimmunoassay
	SSDS	-	synthetic splice donor site
	SSPE	-	NaCl (0.18 M), sodium phosphate (0.01 M), EDTA (0.001 M)
5	TBE	-	Tris-borate/EDTA electrophoresis buffer
	TCA	-	trichloroacetic acid
	TC media	-	tissue culture media
	TMB	-	tetramethylbenzidine
	Tris	-	tris (hydroxymethyl) aminomethane
10	μ Ci	-	microcurie
	μ g	-	microgram
	μ l	-	microliter
	μ M	-	micromolar
	VSV	-	vesicular stomatitis virus
15	X-MLV	-	xenotropic murine leukemia virus

Cell Lines

	HeLa K	--	standard type of HeLa cells; aneuploid, epithelial-like cell line isolated from a human cervical adenocarcinoma [Gey et al., <u>Cancer Res.</u> , 12: 264 (1952); Jones et al., <u>Obstet. Gynecol.</u> , 38: 945-949 (1971)] obtained from Professor B. Korych, [Institute of Medical Microbiology and Immunology, Charles University; Prague, Czech Republic]
20			
25	HeLa D98/AH.2 (also HeLa S)	--	Mutant HeLa clone that is hypoxanthine guanine phosphoribosyl transferase-deficient (HGPRT ⁻) kindly provided by Eric J. Stanbridge [Department of Microbiology, College of Medicine, University of California, Irvine, CA (USA)] and reported in Stanbridge et al., <u>Science</u> , 215: 252-259 (15 Jan. 1982); parent of hybrid cells H/F-N and H/F-T, also obtained from E.J. Stanbridge.
30			
35	NIH-3T3	--	murine fibroblast cell line reported in Aaronson, <u>Science</u> , 237: 178 (1987).

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XC -- cells derived from a rat rhabdomyosarcoma
 induced with Rous sarcoma virus-induced rat
 sarcoma [Svoboda, J., Natl. Cancer Center
 Institute Monograph No. 17, IN:

5 "International Conference on Avian Tumor
 Viruses" (J.W. Beard ed.), pp. 277-298
 (1964)], kindly provided by Jan Svoboda
 [Institute of Molecular Genetics,
10 Czechoslovak Academy of Sciences; Prague,
 Czech Republic]; and

CGL1 -- H/F-N hybrid cells (HeLa D98/AH.2 derivative)

CGL2 -- H/F-N hybrid cells (HeLa D98/AH.2 derivative)

CGL3 -- H/F-T hybrid cells (HeLa D98/AH.2 derivative)

CGL4 -- H/F-T hybrid cells (HeLa D98/Ah.2 derivative)

15 Nucleotide and Amino Acid Sequence Symbols

 The following symbols are used to represent
nucleotides herein:

	<u>Base Symbol</u>	<u>Meaning</u>
20	A	adenine
	C	cytosine
	G	guanine
	T	thymine
	U	uracil
25	I	inosine
	M	A or C
	R	A or G
	W	A or T/U
	S	C or G
30	Y	C or T/U
	K	G or T/U
	V	A or C or G
	H	A or C or T/U

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D	A or G or T/U
B	C or G or T/U
N/X	A or C or G or T/U

There are twenty main amino acids, each of which is
 5 specified by a different arrangement of three adjacent
 nucleotides (triplet code or codon), and which are linked
 together in a specific order to form a characteristic protein.
 A three-letter or one-letter convention is used herein to
 identify said amino acids, as, for example, in Figure 1 as
 10 follows:

	<u>Amino acid name</u>	3 Ltr. <u>Abbrev.</u>	1 Ltr. <u>Abbrev.</u>
	Alanine	Ala	A
	Arginine	Arg	R
15	Asparagine	Asn	N
	Aspartic Acid	Asp	D
	Cysteine	Cys	C
	Glutamic Acid	Glu	E
	Glutamine	Gln	Q
20	Glycine	Gly	G
	Histidine	His	H
	Isoleucine	Ile	I
	Leucine	Leu	L
	Lysine	Lys	K
25	Methionine	Met	M
	Phenylalanine	Phe	F
	Proline	Pro	P
	Serine	Ser	S
	Threonine	Thr	T
30	Tryptophan	Trp	W
	Tyrosine	Tyr	Y
	Valine	Val	V
	Unknown or other		X

BRIEF DESCRIPTION OF THE FIGURES

35 Figure 1 provides the nucleotide sequence for a
 full-length MN cDNA [SEQ. ID. NO.: 1] clone isolated as

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described herein. Figure 1 also sets forth the predicted amino acid sequence [SEQ. ID. NO.: 2] encoded by the cDNA.

Figure 2 compares the results of immunizing baby rats to XC tumor cells with rat serum prepared against the fusion protein MN glutathione S-transferase (GEX-3X-MN) (the IM group) with the results of immunizing baby rats with control rat sera (the C group). Each point on the graph represents the tumor weight of a tumor from one rat. Example 2 details those experiments.

Figure 3a-d provides a 10,898 bp complete genomic sequence of MN [SEQ. ID. NO.: 5]. The base count is as follows: 2654 A; 2739 C; 2645 G; and 2859 T. The 11 exons are shown in capital letters.

Figure 4 is a restriction map of the full-length MN cDNA. The open reading frame is shown as an open box. The thick lines below the restriction map illustrate the sizes and positions of two overlapping cDNA clones. The horizontal arrows indicate the positions of primers R1 [SEQ. ID. NO.: 7] and R2 [SEQ. ID. NO.: 8] used for the 5' end RACE. Relevant restriction sites are BamHI (B), EcoRV (V), EcoRI (E), PstI (Ps), PvuII (Pv).

Figure 5 is a map of the human MN gene. The numbered cross-hatched boxes represent exons. The box designated LTR denotes a region of homology to HERV-K LTR. The empty boxes are Alu-related sequences.

Figure 6 is a nucleotide sequence for the proposed promoter of the human MN gene [SEQ. ID. No.: 27]. The nucleotides are numbered from the transcription initiation site according to RNase protection assay. Potential regulatory elements are overlined. Transcription start sites are indicated by asterisks (RNase protection) and dots (RACE). The sequence of the 1st exon begins under the asterisks.

Figure 7 provides a schematic of the alignment of MN genomic clones according to their position related to the transcription initiation site. All the genomic fragments except Bd3 were isolated from a lambda FIX III genomic library derived from HeLa cells. Clone Bd3 was derived from a human fetal brain library.

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Figure 8 shows the construction and cloning of a series of 5' deletion mutants of MN's putative promoter region linked to the bacterial CAT gene.

DETAILED DESCRIPTION

5 The MN gene is shown herein to be organized into 11 exons and 10 introns. Described herein is the cloning and sequencing of the MN cDNA and genomic sequences, and the genetic engineering of MN proteins -- such as the GEX-3X-MN, MN-PA, MN-Fc and MN 20-19 proteins. The recombinant MN
10 proteins can be conveniently purified by affinity chromatography.

 MN is manifested in HeLa cells by a twin protein, p54/58N. Immunoblots using a monoclonal antibody reactive with p54/58N (MAb M75) revealed two bands at 54 kd and 58 kd.
15 Those two bands may correspond to one type of protein that differs by glycosylation pattern or by how it is processed. Herein, the phrase "twin protein" indicates p54/58N.

 The expression of MN proteins appears to be diagnostic/prognostic for neoplastic disease. The MN twin
20 protein, p54/58N, was found to be expressed in HeLa cells and in Stanbridge's tumorigenic (H/F-T) hybrid cells [Stanbridge et al., Somatic Cell Genet., 7: 699-712 (1981); and Stanbridge et al., Science, 215: 252-259 (1982)] but not in fibroblasts or in non-tumorigenic (H/F-N) hybrid cells [Stanbridge et al.,
25 id.]. In early studies reported in Zavada et al. WO 93/18152, supra, MN proteins were found in immunoblots prepared from human ovarian, endometrial and uterine cervical carcinomas, and in some benign neoplasias (as mammary papilloma) but not from normal ovarian, endometrial, uterine or placental
30 tissues. Example 1 herein details further research on MN gene expression wherein MN antigen, as detected by immunohistochemical staining, was found to be prevalent in tumor cells of a number of cancers, including cervical, bladder, head and neck, and renal cell carcinomas among
35 others. Further, the immunohistochemical staining experiments of Example 1 show that among normal tissues tested, only normal stomach tissues showed routinely and extensively the

presence of MN antigen. MN antigen is further shown herein to be present sometimes in morphologically normal-appearing areas of tissue specimens exhibiting dysplasia and/or malignancy.

MN Gene--Cloning and Sequencing

5 Figure 1 provides the nucleotide sequence for a full-length MN cDNA clone isolated as described below [SEQ. ID. NO.: 1]. Figure 3a-d provides a complete MN genomic sequence [SEQ. ID. NO.: 5]. Figure 6 shows the nucleotide sequence for a proposed MN promoter [SEQ. ID. NO.: 27].

10 It is understood that because of the degeneracy of the genetic code, that is, that more than one codon will code for one amino acid [for example, the codons TTA, TTG, CTT, CTC, CTA and CTG each code for the amino acid leucine (leu)], that variations of the nucleotide sequences in, for example,
15 SEQ. ID. NOS.: 1 and 5 wherein one codon is substituted for another, would produce a substantially equivalent protein or polypeptide according to this invention. All such variations in the nucleotide sequences of the MN cDNA and complementary nucleic acid sequences are included within the scope of this
20 invention.

It is further understood that the nucleotide sequences herein described and shown in Figures 1, 3a-d and 6, represent only the precise structures of the cDNA, genomic and promoter nucleotide sequences isolated and described herein.

25 It is expected that slightly modified nucleotide sequences will be found or can be modified by techniques known in the art to code for substantially similar or homologous MN proteins and polypeptides, for example, those having similar epitopes, and such nucleotide sequences and proteins/

30 polypeptides are considered to be equivalents for the purpose of this invention. DNA or RNA having equivalent codons is considered within the scope of the invention, as are synthetic nucleic acid sequences that encode proteins/polypeptides homologous or substantially homologous to MN

35 proteins/polypeptides, as well as those nucleic acid sequences that would hybridize to said exemplary sequences [SEQ. ID. NOS. 1, 5 and 27] under stringent conditions, or that, but for

the degeneracy of the genetic code would hybridize to said cDNA nucleotide sequences under stringent hybridization conditions. Modifications and variations of nucleic acid sequences as indicated herein are considered to result in
5 sequences that are substantially the same as the exemplary MN sequences and fragments thereof.

Partial cDNA clone

In Zavada et al., *id.*, the isolation of a partial MN cDNA clone of 1397 bp in length was described. A lambda gt11
10 cDNA library of LMCV-infected HeLa cells was prepared and subjected to immunoscreening with Mab M75 in combination with goat anti-mouse antibodies conjugated with alkaline phosphatase. One positive clone was picked and subcloned into the NotI site of pBluescript KS [Stratagen; La Jolla, CA
15 (USA)] thereby creating pBluscript-MN.

Two oppositely oriented nested deletions were made using Erase-a-Base™ kit [Promega; Madison, WI (USA)] and sequenced by dideoxy method with a T7 sequencing kit [Pharmacia; Piscataway, NJ (USA)]. The sequencing showed a
20 partial cDNA clone, the insert being 1397 bp long. The sequence comprises a large 1290 bp open reading frame and 107 bp 3' untranslated region containing a polyadenylation signal (AATAAA). However, the sequence surrounding the first ATG codon in the open reading frame (ORF) did not fit the
25 definition of a translational start site. In addition, as followed from a comparison of the size of the MN clone with that of the corresponding mRNA in a Northern blot, the cDNA was shown to be missing about 100 bp from the 5' end of its sequence.

30 Full-Length cDNA Clone

Attempts to isolate a full-length clone from the original cDNA library failed. Therefore, the inventors performed a rapid amplification of cDNA ends (RACE) using MN-specific primers, R1 and R2 [SEQ. ID. NOS.: 7 and 8], derived
35 from the 5' region of the original cDNA clone. The RACE product was inserted into pBluescript, and the entire

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population of recombinant plasmids was sequenced with an MN-specific primer ODN1 [SEQ. ID. NO.: 3]. In that way, a reliable sequence at the very 5' end of the MN cDNA as shown in Figure 1 [SEQ. ID. NO.: 1] was obtained.

5 Specifically, RACE was performed using 5' RACE System [GIBCO BRL; Gaithersburg, MD (USA)] as follows. 1 µg of mRNA (the same as above) was used as a template for the first strand cDNA synthesis which was primed by the MN-specific antisense oligonucleotide, R1 (5'-
10 TGGGGTTCTTGAGGATCTCCAGGAG-3') [SEQ. ID. NO.: 7]. The first strand product was precipitated twice in the presence of ammonium acetate and a homopolymeric C tail was attached to its 3' end by TdT. Tailed cDNA was then amplified by PCR using a nested primer, R2 (5'-CTCTAACTTCAGGGAGCCCTCTTCTT-3')
15 [SEQ. ID. NO.: 8] and an anchor primer that anneals to the homopolymeric tail (5'-CUACUACUACUAGGCCACGCGTCGACTAGTACGGGI
IGGGIIGGGIIG-3') [SEQ. ID. NO.: 9]. The amplified product was digested with BamHI and SalI restriction enzymes and cloned into pBluescript II KS plasmid. After transformation,
20 plasmid DNA was purified from the whole population of transformed cells and used as a template for sequencing with the MN-specific primer ODN1 [SEQ. ID. NO.: 3; a 29-mer 5'
CGCCCAGTGGGTCATCTTCCCCAGAAGAG 3'].

Based upon results of the RACE analysis, the full-
25 length MN cDNA sequence was seen to contain a single ORF starting at position 12, with an ATG codon that is in a good context (GCGCATGG) with the rule proposed for translation initiation [Kozak, J. Cell. Biol., 108: 229-241 (1989)].
[See below under Mapping of MN Gene Transcription Initiation
30 Site for fine mapping of the 5' end of the MN gene.] The AT rich 3' untranslated region contains a polyadenylation signal (AATAAA) preceding the end of the cDNA by 10 bp.
Surprisingly, the sequence from the original clone as well as from four additional clones obtained from the same cDNA
35 library did not reveal any poly(A) tail. Moreover, just downstream of the poly(A) signal, an ATTTA motif that is thought to contribute to mRNA instability [Shaw and Kamen, Cell, 46: 659-667 (1986)] was found. That fact raised the

possibility that the poly (A) tail is missing due to the specific degradation of the MN mRNA.

Genomic clones

To study MN regulation, MN genomic clones were isolated. One MN genomic clone (Bd3) was isolated from a human cosmid library prepared from fetal brain using both MN cDNA as a probe and the MN-specific primers derived from the 5' end of the cDNA ODN1 [SEQ. ID. NO.: 3, supra] and ODN2 [SEQ. ID NO.: 4; 19-mer (5' GGAATCCTCCTGCATCCGG 3')]. Sequence analysis revealed that that genomic clone covered a region upstream from a MN transcription start site and ending with the BamHI restriction site localized inside the MN cDNA. Other MN genomic clones can be similarly isolated.

In order to identify the complete genomic region of MN, the human genomic library in Lambda FIX II vector (Stratagene) was prepared from HeLa chromosomal DNA and screened by plaque hybridization using MN cDNA as described below. Several independent MN recombinant phages were identified, isolated and characterized by restriction mapping and hybridization analyses. Four overlapping recombinants covering the whole genomic region of MN were selected, digested and subcloned into pBluescript. The subclones were then subjected to bidirectional nested deletions and sequencing. DNA sequences were compiled and analyzed by computer using the DNASIS software package.

The details of isolating genomic clones covering the complete genomic region for MN are provided below. Figure 7 provides a schematic of the alignment of MN genomic clones according to the transcription initiation site. Plasmids containing the A4a clone and the XE1 and XE3 subclones were deposited at the American Type Culture Collection (ATCC) at 12301 Parklawn Drive, Rockville, MD 20852 (USA) on June 6, 1995, respectively under ATCC Deposit Nos. 97199, 97200, and 97198.

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Isolation of Genomic DNA Clones

The Sau3AI human HeLa genomic library was prepared in Lambda FIX II vector [Stratagene; La Jolla, CA (USA)] according to manufacturer's protocol. Human fetal brain cosmid library in SuperCos cosmid was from Stratagene. Recombinant phages or bacteria were plated at 1×10^5 plaque forming units on 22x22 cm Nunc plates or 5×10^4 cells on 150 mm Petri dishes, and plaques or colonies were transferred to Hybond N membranes (Amersham). Hybridization was carried out with the full-length MN cDNA labeled with [P^{32}]PdCTP by the Multiprime DNA labeling method (Amersham) at 65°C in 6 x SSC, 0.5% SDS, 10 x Denhardt's and 0.2 mg/1 ml salmon sperm DNA. Filters were washed twice in 2 x SSC, 0.1% SDS at 65°C for 20 min. The dried filters were exposed to X-ray films, and positive clones were picked up. Phages and bacteria were isolated by 3-4 sequential rounds of screening.

Subcloning and DNA Sequencing

Genomic DNA fragments were subcloned into a pBluescript KS and templates for sequencing were generated by serial nested deletions using the Erase-a-Base system. Sequencing was performed by the dideoxynucleotide chain termination method using T7 sequencing kit (Pharmacia). Nucleotide sequence alignments and analyses were carried out using the DNASIS software package (Hitachi Software Engineering).

Exon-Intron Structure of Complete MN Genomic Region

The complete sequence of the overlapping clones contains 10,898 bp (SEQ. ID. NO.: 5). Figure 5 depicts the organization of the human MN gene, showing the location of all 11 exons as well as the 2 upstream and 6 intronic Alu repeat elements. All the exons are small, ranging from 27 to 191 bp, with the exception of the first exon which is 415 bp. The intron sizes range from 89 to 1400 bp.

Table 1 below lists the splice donor and acceptor sequences that conform to consensus splice sequences including

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the AG-GT motif [Mount, "A catalogue of splice junction sequences," Nucleic Acids Res. 10: 459-472 (1982)].

TABLE 1
Exon-Intron Structure of the Human MN Gene

	Exon	Size	Genomic Position**	SEQ ID NO	5'splice donor	SEQ ID No
5	1	415	*3537-3951	28	AGAAG gtaagt	67
	2	30	5126-5155	29	TGGAG gtgaga	68
	3	171	5349-5519	30	CAGTC gtgagg	69
10	4	143	5651-5793	31	CCGAG gtgagc	70
	5	93	5883-5975	32	TGGAG gtacca	71
	6	67	7376-7442	33	GGAAG gtcagt	72
	7	158	8777-8934	34	AGCAG gtgggc	73
	8	145	9447-9591	35	GCCAG gtacag	74
15	9	27	9706-9732	36	TGCTG gtgagt	75
	10	82	10350-10431	37	CACAG gtatta	76
	11	191	10562-10752	38	ATAAT end	

	Intron	Size	Genomic Position**	SEQ ID NO	3'splice acceptor	SEQ ID NO
	1	1174	3952-5125	39	atacag GGGAT	77
20	2	193	5156-5348	40	ccccag GCGAC	78
	3	131	5520-5650	41	acgcag TGCAA	79
	4	89	5794-5882	42	tttcag ATCCA	80
	5	1400	5976-7375	43	ccccag GAGGG	81
	6	1334	7443-8776	44	tcacag GCTCA	82
25	7	512	8935-9446	45	ccctag CTCCA	83
	8	114	9592-9705	46	ctccag TCCAG	84
	9	617	9733-10349	47	tcgcag GTGACA	85
	10	130	10432-10561	48	acacag AAGGG	86

** positions are related to nt numbering in whole genomic sequence including the 5' flanking region [Figure 3a-d]

* number corresponds to transcription initiation site determined below by RNase protection assay

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A search for sequences related to MN gene in the EMBL Data Library did not reveal any specific homology except for 6 complete and 2 partial Alu-type repeats with homology to Alu sequences ranging from 69.8% to 91% [Jurka and
5 Milosavljevic, "Reconstruction and analysis of human Alu genes," J. Mol. Evol. 32: 105-121 (1991)]. Below under the Characterization of the 5' Flanking Region, also a 222 bp sequence proximal to the 5' end of the genomic region is shown to be closely homologous to a region of the HERV-K LTR.

10 Mapping of MN Gene Transcription Initiation Site

In the earlier attempt to localize the site of transcription initiation of the MN gene by RACE (above), the obtained a major PCR fragment whose sequence placed the start site 12 bp upstream from the first codon of the ORF. That
15 result was obtained probably due to a preferential amplification of the shortest form of mRNA. Therefore, the inventors used an RNase protection assay (RNP) for fine mapping of the 5' end of the MN gene. The probe was a uniformly labeled 470 nucleotide copy RNA (nt -205 to +265)
20 [SEQ. ID. NO.: 55], which was hybridized to total RNA from MN-expressing HeLa and CGL3 cells and analyzed on a sequencing gel. That analysis has shown that the MN gene transcription initiates at multiple sites, the 5' end of the longest MN transcript being 30 nt longer than that previously
25 characterized by RACE.

RNase Protection Assay

³²P-labeled RNA probes were prepared with an RNA Transcription kit (Stratagene). In vitro transcription reactions were carried out using 1 µg of the linearized
30 plasmid as a template, 50 µCi of [P³²P]rUTP (800 Ci/mmol), 10 U of either T3 or T7 RNA polymerase and other components of the Transcription Kit following instructions of the supplier. For mapping of the 5' end of MN mRNA, the 470 bp NcoI-BamHI fragment (NcoI filled in by Klenow enzyme) of Bd3 clone (nt -
35 205 to +265 related to transcription start) was subcloned to EcoRV-BamHI sites of pBluescript SK+, linearized with HindIII

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and labeled with T3 RNA polymerase. For the 3' end mRNA analysis, probe, that was prepared using T7 RNA polymerase on KS-dXE3-16 template (one of the nested deletion clones of MN genomic region XE3 subclone) digested with Sau3AI (which cuts
5 exon 11 at position 10,629), was used. Approximately 3×10^5 cpm of RNA probe were used per one RNase protection assay reaction.

RNase protection assays (RNP) were performed using Lysate RNase Protection Kit (USB/Amersham) according to
10 protocols of the supplier. Briefly, cells were lysed using Lysis Solution at concentration of approximately 10^7 cells/ml, and 45 μ l of the cell homogenate were used in RNA/RNA hybridization reactions with 32 P-labeled RNA probes prepared as described above. Following overnight hybridizations at 42°C,
15 homogenates were treated for 30 min at 37°C with RNase cocktail mix. Protected RNA duplexes were run on polyacrylamide/urea denaturing sequencing gels. Fixed and dried gels were exposed to X-ray film for 24 - 72 hours.

Mapping of MN Gene Transcription Termination Site

20 An RNase protection assay, as described above, was also used to verify also the 3' end of the MN cDNA. That was important with respect to our previous finding that the cDNA contains a poly(A) signal but lacks a poly(A) tail, which could be lost during the proposed degradation of MN mRNA due
25 to the presence of an instability motif in its 3' untranslated region. RNP analysis of MN mRNA with the fragment of the genomic clone XE3 covering the region of interest corroborated our data from MN cDNA sequencing, since the 3' end of the protected fragment corresponded to the last base of MN cDNA
30 (position 10,752 of the genomic sequence). That site also meets the requirement for the presence of a second signal in the genomic sequence that is needed for transcription termination and polyadenylation [McLauchlan et al., Nucleic Acids Res., 13: 1347 (1985)]. Motif TGTGTTAGT (nt 10,759-
35 10,767) corresponds well to both the consensus sequence and the position of that signal within 22 bp downstream from the polyA signal (nt 10,737-10,742).

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Characterization of the 5' Flanking Region

The Bd3 genomic clone isolated from human fetal brain cosmid library was found to cover a region of 3.5 kb upstream from the transcription start site of the MN gene. It contains no significant coding region. Two Alu repeats are situated at positions -2587 to -2296 [SEQ. ID. NO.: 56] and -1138 to -877 [SEQ. ID. NO.: 57] (with respect to the transcription start determined by RNP). The sequence proximal to the 5' end is strongly homologous (91.4% identity) to the U3 region of long terminal repeats of human endogenous retroviruses HERV-K [Ono, M., "Molecular cloning and long terminal repeat sequences of human endogenous retrovirus genes related to types A and B retrovirus genes," J. Virol. 58: 937-944 (1986)]. The LTR-like fragment is 222 bp long with an A-rich tail at its 3' end. Most probably, it represents part of SINE (short interspersed repeated sequence) type nonviral retroposon derived from HERV-K [Ono et al., "A novel human nonviral retroposon derived from an endogenous retrovirus," Nucleic Acids Res., 15: 8725-8373 (1987)]. There are no sequences corresponding to regulatory elements in this fragment, since the 3' part of U3, and the entire R and U5 regions of LTR are absent from the Bd3 genomic clone, and the glucocorticoid responsive element as well as the enhancer core sequences are beyond its 5' border.

However, two keratinocyte-dependent enhancers were identified in the sequence downstream from the LTR-like fragment at positions -3010 and -2814. Those elements are involved in transcriptional regulation of the E6-E7 oncogenes of human papillomaviruses and are thought to account for their tissue specificity [Cripe et al., "Transcriptional regulation of the human papilloma-virus-16 E6-E7 promoter by a keratinocyte-dependent enhancer, and by viral E2 trans-activator and repressor gene products: implications for cervical carcinogenesis," EMBO J., 6: 3745-3753 (1987)].

Nucleotide sequence analysis of the DNA 5' to the transcription start (from nt -507) revealed no recognizable TATA box within the expected distance from the beginning of the first exon (Figure 6). However, the presence of potential

binding sites for transcription factors suggests that this region might contain a promoter for the MN gene. There are several consensus sequences for transcription factors AP1 and AP2 as well as for other regulatory elements, including a p53 binding site [Locker and Buzard, "A dictionary of transcription control sequences," J. DNA Sequencing and Mapping, 1: 3-11 (1990); Imagawa et al., "Transcription factor AP-2 mediates induction by two different signal-transduction pathways: protein kinase C and cAMP," Cell, 51: 251-260 (1987); El Deiry et al., "Human genomic DNA sequences define a consensus binding site for p53," Nat. Genet., 1: 44-49 (1992)]. Although the putative promoter region contains 59.3% C+G, it does not have additional attributes of CpG-rich islands that are typical for TATA-less promoters of housekeeping genes [Bird, "CpG-rich islands and the function of DNA methylation," Nature, 321: 209-213 (1986)]. Another class of genes lacking TATA box utilizes the initiator (Inr) element as a promoter. Many of these genes are not constitutively active, but they are rather regulated during differentiation or development. The Inr has a consensus sequence of PyPyPyCAPyPyPyPyPy [SEQ. ID. NO.: 23] and encompasses the transcription start site [Smale and Baltimore, "The 'initiator' as a transcription control element," Cell, 57: 103-113 (1989)]. There are two such consensus sequences in the MN putative promoter; however, they do not overlap the transcription start (Figure 6).

In the initial experiments, the inventors were unable to show promoter activity in human carcinoma cells HeLa and CGL3 that express MN, using the 3.5 kb Bd3 fragment and series of its deletion mutants (from nt -933 to -30) [SEQ. ID. NO.: 58] fused to chloramphenicol acetyl transferase (CAT) gene in a transient system. This might indicate that either the promoter activity of the region 5' to the MN transcription start is below the sensitivity of the CAT assay, or additional regulatory elements not present in our constructs are required for driving the expression of MN gene.

With respect to this fact, an interesting region was found in the middle of the MN gene. The region is about 1.4

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kb in length [nt 4,600-6,000 of the genomic sequence; SEQ. ID. NO.: 49] and spans from the 3' part of the 1st intron to the end of the 5th exon. The region has the character of a typical CpG-rich island, with 62.8% C+G content and 82 CpG: 5 131 GpC dinucleotides. Moreover, there are multiple putative binding sites for transcription factors AP2 and Sp1 [Locker and Buzard, supra; Briggs et al., "Purification and biochemical characterization of the promoter-specific transcription factor Sp-1," Science, 234: 47-52 (1986)] 10 concentrated in the center of this area. Particularly the 3rd intron of 131 bp in length contains three Sp1 and three AP2 consensus sequences. That data indicates the possible involvement of that region in the regulation of MN gene expression. However, functionality of that region, as well as 15 other regulatory elements found in the proposed 5' MN promoter, remains to be determined.

MN Promoter Analysis

To define sequences necessary for MN gene expression, a series of 5' deletion mutants of the putative 20 promoter region were fused to the bacterial chloramphenicol acetyltransferase (CAT) gene. [See Figure 8.] The pMN-CAT deletion constructs were transfected using a DEAE dextran method for transient expression into HeLa and CGL3 cells. Those cells were used since they naturally express MN protein, 25 and thus, should contain all the required transcription factors.

After 48 hours, crude cell lysates were prepared and the activity of the expressed CAT was evaluated according to acetylation of [¹⁴C]chloramphenicol by thin layer 30 chromatography. However, no MN promoter CAT activity was detected in either the HeLa or the CGL3 cells in a transient system. On the other hand, reporter CAT plasmids with viral promoters (e.g. pBLV-LTR + tax transactivator, pRSV CAT and pSV2 CAT), that served as positive controls, gave strong 35 signals on the chromatogram. [pSV2 CAT carries the SV40 origin and expresses CAT from the SV40 early promoter (P_E).

pRSV CAT expresses CAT from the Rous sarcoma virus (RSV) LTR promoter (P_{LTR}).]

No detectable CAT activity was observed in additional experiments using increasing amounts of transfected
5 plasmids (from 2 to 20 g DNA per dish) and prolonged periods of cell incubation after transcription. Increased cell density also did not improve the results (in contrast to the expectations based on density-dependent expression of native MN protein in HeLa cells). Since the inventors had found
10 consensus sequences for transcription factors AP2 and AP1 in the putative MN promoter, they studied the effect of their inducers dexamethasone (1 M) and phorbol ester phorbol 12-myristate 13-acetate (PMA 50 ng/ml) on CAT activity. However, the MN promoter was unresponsive to those compounds.

15 The following provides explanations for the results:
--the putative MN promoter immediately preceding the transcription initiation site is very weak, and its activity is below the sensitivity of a standard CAT assay; --additional sequences (e.g. enhancers) are necessary for MN transcription.

20 To further shed light on the regulation of MN expression at the level of transcription, constructs, analogously prepared to the MN-CAT constructs, are prepared, wherein the MN promoter region is upstream from the neomycin phosphotransferase gene engineered for mammalian expression.
25 Such constructs are then transfected to cells which are subjected to selection with G418. Activity of the promoter is then evaluated on the basis of the number of G418 resistant colonies that result. That method has the capacity to detect activity of a promoter that is 50 to 100 times weaker in
30 comparison to promoters detectable by a CAT assay.

Deduced Amino Acid Sequence

The ORF of the MN cDNA shown in Figure 1 has the coding capacity for a 459 amino acid protein with a calculated molecular weight of 49.7 kd. MN protein has an estimated pI
35 of about 4. As assessed by amino acid sequence analysis, the deduced primary structure of the MN protein can be divided into four distinct regions. The initial hydrophobic region of

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37 amino acids (AA) corresponds to a signal peptide. The mature protein has an N-terminal part of 377 AA, a hydrophobic transmembrane segment of 20 AA and a C-terminal region of 25 AA. Alternatively, the MN protein can be viewed as having five domains as follows: (1) a signal peptide [amino acids (AA) 1-37; SEQ. ID. NO.: 6]; (2) a region of homology to collagen alpha1 chain (AA 38-135; SEQ. ID. NO.: 50); (3) a carbonic anhydrase domain (AA 136-391; SEQ. ID. NO.: 51); (4) a transmembrane region (AA 414-433; SEQ. ID. NO.: 52); and (5) an intracellular C terminus (AA 434-459; SEQ. ID. NO.: 53). (The AA numbers are keyed to Figure 1.)

More detailed insight into MN protein primary structure disclosed the presence of several consensus sequences. One potential N-glycosylation site was found at position 346 of Figure 1. That feature, together with a predicted membrane-spanning region are consistent with the results, in which MN was shown to be an N-glycosylated protein localized in the plasma membrane. MN protein sequence deduced from cDNA was also found to contain seven S/TPXX sequence elements [SEQ. ID. NOS.: 25 AND 26] (one of them is in the signal peptide) defined by Suzuki, J. Mol. Biol., 207: 61-84 (1989) as motifs frequently found in gene regulatory proteins. However, only two of them are composed of the suggested consensus amino acids.

Experiments have shown that the MN protein is able to bind zinc cations, as shown by affinity chromatography using Zn-charged chelating sepharose. MN protein immunoprecipitated from HeLa cells by Mab M75 was found to have weak catalytic activity of CA. The CA-like domain of MN has a structural predisposition to serve as a binding site for small soluble domains. Thus, MN protein could mediate some kind of signal transduction.

MN protein from LCMV-infected HeLa cells was shown by using DNA cellulose affinity chromatography to bind to immobilized double-stranded salmon sperm DNA. The binding activity required both the presence of zinc cations and the absence of a reducing agent in the binding buffer.

Sequence Similarities

Computer analysis of the MN cDNA sequence was carried out using DNASIS and PROSID (Pharmacia Software packages). GenBank, EMBL, Protein Identification Resource and
5 SWISS-PROT databases were searched for all possible sequence similarities. In addition, a search for proteins sharing sequence similarities with MN was performed in the MIPS databank with the FastA program [Pearson and Lipman, PNAS (USA), 85: 2444 (1988)].

10 The MN gene was found to clearly be a novel sequence derived from the human genome. Searches for amino acid sequence similarities in protein databases revealed as the closest homology a level of sequence identity (38.9% in 256 AA or 44% in an 170 AA overlap) between the central part of the
15 MN protein [AAs 136-391 (SEQ. ID. NO: 51)] or 221-390 [SEQ. ID. NO.: 54] of Figure 1 and carbonic anhydrases (CA). However, the overall sequence homology between the cDNA MN sequence and cDNA sequences encoding different CA isoenzymes is in a homology range of 48-50% which is considered by ones
20 in the art to be low. Therefore, the MN cDNA sequence is not closely related to any CA cDNA sequences.

Only very closely related nt sequences having a homology of at least 80-90% would hybridize to each other under stringent conditions. A sequence comparison of the MN
25 cDNA sequence shown in Figure 1 and a corresponding cDNA of the human carbonic anhydrase II (CA II) showed that there are no stretches of identity between the two sequences that would be long enough to allow for a segment of the CA II cDNA sequence having 50 or more nucleotides to hybridize under
30 stringent hybridization conditions to the MN cDNA or vice versa.

Although MN deduced amino acid sequences show some homology to known carbonic anhydrases, they differ from them in several respects. Seven carbonic anhydrases are known
35 [Dodgson et al. (eds.), The Carbonic Anhydrases, (Plenum Press; New York/London (1991))]. All of the known carbonic anhydrases are proteins of about 30 kd, smaller than the p54/58N-related products of the MN gene. Further, the

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carbonic anhydrases do not form oligomers as do the MN-related proteins.

The N-terminal part of the MN protein (AA 38-135; SEQ. ID. NO.: 50) shows a 27-30% identity with human collagen 5 alpha1 chain, which is an important component of the extracellular matrix.

MN Proteins and/or Polypeptides

The phrase "MN proteins and/or polypeptides" (MN proteins/polypeptides) is herein defined to mean proteins 10 and/or polypeptides encoded by an MN gene or fragments thereof. An exemplary and preferred MN protein according to this invention has the deduced amino acid sequence shown in Figure 1. Preferred MN proteins/polypeptides are those 15 proteins and/or polypeptides that have substantial homology with the MN protein shown in Figure 1. For example, such substantially homologous MN proteins/ polypeptides are those that are reactive with the MN-specific antibodies of this invention, preferably the Mabs M75, MN12, MN9 and MN7 or their equivalents.

20 A "polypeptide" is a chain of amino acids covalently bound by peptide linkages and is herein considered to be composed of 50 or less amino acids. A "protein" is herein defined to be a polypeptide composed of more than 50 amino acids.

25 MN proteins exhibit several interesting features: cell membrane localization, cell density dependent expression in HeLa cells, correlation with the tumorigenic phenotype of HeLa x fibroblast somatic cell hybrids, and expression in several human carcinomas among other tissues. As demonstrated 30 herein, for example, in Example 1, MN protein can be found directly in tumor tissue sections but not in general in counterpart normal tissues (exceptions noted *infra* in Example 1 as in normal stomach tissues). MN is also expressed sometimes in morphologically normal appearing areas of tissue 35 specimens exhibiting dysplasia and/or malignancy. Taken together, these features suggest a possible involvement of MN

in the regulation of cell proliferation, differentiation and/or transformation.

It can be appreciated that a protein or polypeptide produced by a neoplastic cell in vivo could be altered in sequence from that produced by a tumor cell in cell culture or by a transformed cell. Thus, MN proteins and/or polypeptides which have varying amino acid sequences including without limitation, amino acid substitutions, extensions, deletions, truncations and combinations thereof, fall within the scope of this invention. It can also be appreciated that a protein extant within body fluids is subject to degradative processes, such as, proteolytic processes; thus, MN proteins that are significantly truncated and MN polypeptides may be found in body fluids, such as, sera. The phrase "MN antigen" is used herein to encompass MN proteins and/or polypeptides.

It will further be appreciated that the amino acid sequence of MN proteins and polypeptides can be modified by genetic techniques. One or more amino acids can be deleted or substituted. Such amino acid changes may not cause any measurable change in the biological activity of the protein or polypeptide and result in proteins or polypeptides which are within the scope of this invention, as well as, MN muteins.

The MN proteins and polypeptides of this invention can be prepared in a variety of ways according to this invention, for example, recombinantly, synthetically or otherwise biologically, that is, by cleaving longer proteins and polypeptides enzymatically and/or chemically. A preferred method to prepare MN proteins is by a recombinant means. Particularly preferred methods of recombinantly producing MN proteins are described below for the GEX-3X-MN, MN 20-19, MN-Fc and MN-PA proteins.

Recombinant Production of MN Proteins and Polypeptides

A representative method to prepare the MN proteins shown in Figure 1 or fragments thereof would be to insert the full-length or an appropriate fragment of MN cDNA into an appropriate expression vector as exemplified below. In Zavada et al., WO 93/18152, supra, production of a fusion protein

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GEX-3X-MN using the partial cDNA clone (described above) in the vector pGEX-3X (Pharmacia) is described. Nonglycosylated GEX-3X-MN (the Mn fusion protein MN glutathione S-transferase) from XL1-Blue cells. Herein described is the recombinant production of both a glycosylated MN protein expressed from insect cells and a nonglycosylated MN protein expressed from E. coli using the expression plasmid pEt-22b [Novagen Inc.; Madison, WI (USA)].

Baculovirus Expression Systems. Recombinant baculovirus express vectors have been developed for infection into several types of insect cells. For example, recombinant baculoviruses have been developed for among others: Aedes aegypti, Autographa californica, Bombyx mor, Drosophila melanogaster, Heliothis zea, Spodoptera frugiperda, and Trichoplusia ni [PCT Pub. No. WO 89/046699; Wright, Nature, 321: 718 (1986); Fraser et al., In Vitro Cell Dev. Biol., 25: 225 (1989). Methods of introducing exogenous DNA into insect hosts are well-known in the art. DNA transfection and viral infection procedures usually vary with the insect genus to be transformed. See, for example, Autographa [Carstens et al., Virology, 101: 311 (1980)]; Spodoptera [Kang, "Baculovirus Vectors for Expression of Foreign Genes," in: Advances in Virus Research, 35 (1988)]; and Heliothis (virescens) [PCT Pub. No. WO 88/02030].

A wide variety of other host-cloning vector combinations may be usefully employed in cloning the MN DNA isolated as described herein. For example, useful cloning vehicles may include chromosomal, nonchromosomal and synthetic DNA sequences such as various known bacterial plasmids such as pBR322, other E. coli plasmids and their derivatives and wider host range plasmids such as RP4, phage DNA, such as, the numerous derivatives of phage lambda, e.g., NB989 and vectors derived from combinations of plasmids and phage DNAs such as plasmids which have been modified to employ phage DNA expression control sequences.

Useful hosts may be eukaryotic or prokaryotic and include bacterial hosts such as E. coli and other bacterial strains, yeasts and other fungi, animal or plant hosts such as

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animal or plant cells in culture, insect cells and other hosts. Of course, not all hosts may be equally efficient. The particular selection of host-cloning vehicle combination may be made by those of skill in the art after due
5 consideration of the principles set forth herein without departing from the scope of this invention.

The particular site chosen for insertion of the selected DNA fragment into the cloning vehicle to form a recombinant DNA molecule is determined by a variety of
10 factors. These include size and structure of the protein or polypeptide to be expressed, susceptibility of the desired protein or polypeptide to endoenzymatic degradation by the host cell components and contamination by its proteins, expression characteristics such as the location of start and
15 stop codons, and other factors recognized by those of skill in the art.

The recombinant nucleic acid molecule containing the MN gene, fragment thereof, or cDNA therefrom, may be employed to transform a host so as to permit that host (transformant)
20 to express the structural gene or fragment thereof and to produce the protein or polypeptide for which the hybrid DNA encodes. The recombinant nucleic acid molecule may also be employed to transform a host so as to permit that host on replication to produce additional recombinant nucleic acid
25 molecules as a source of MN nucleic acid and fragments thereof. The selection of an appropriate host for either of those uses is controlled by a number of factors recognized in the art. These include, for example, compatibility with the chosen vector, toxicity of the co-products, ease of recovery
30 of the desired protein or polypeptide, expression characteristics, biosafety and costs.

Where the host cell is a procaryote such as E. coli, competent cells which are capable of DNA uptake are prepared from cells harvested after exponential growth phase and
35 subsequently treated by the CaCl_2 method by well known procedures. Transformation can also be performed after forming a protoplast of the host cell.

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Where the host used is an eucaryote, transfection methods such as the use of a calcium phosphate-precipitate, electroporation, conventional mechanical procedures such as microinjection, insertion of a plasmid encapsulated in red blood cell ghosts or in liposomes, treatment of cells with agents such as lysophosphatidyl-choline or use of virus vectors, or the like may be used.

The level of production of a protein or polypeptide is governed by three major factors: (1) the number of copies of the gene or DNA sequence encoding for it within the cell; (2) the efficiency with which those gene and sequence copies are transcribed and translated; and (3) the stability of the mRNA. Efficiencies of transcription and translation (which together comprise expression) are in turn dependent upon nucleotide sequences, normally situated ahead of the desired coding sequence. Those nucleotide sequences or expression control sequences define, inter alia, the location at which an RNA polymerase interacts to initiate transcription (the promoter sequence) and at which ribosomes bind and interact with the mRNA (the product of transcription) to initiate translation. Not all such expression control sequences function with equal efficiency. It is thus of advantage to separate the specific coding sequences for the desired protein from their adjacent nucleotide sequences and fuse them instead to known expression control sequences so as to favor higher levels of expression. This having been achieved, the newly engineered DNA fragment may be inserted into a multicopy plasmid or a bacteriophage derivative in order to increase the number of gene or sequence copies within the cell and thereby further improve the yield of expressed protein.

Several expression control sequences may be employed. These include the operator, promoter and ribosome binding and interaction sequences (including sequences such as the Shine-Dalgarno sequences) of the lactose operon of E. coli ("the lac system"), the corresponding sequences of the tryptophan synthetase system of E. coli ("the trp system"), a fusion of the trp and lac promoter ("the tac system"), the major operator and promoter regions of phage lambda ($O_L P_L$ and

O_RP_R), and the control region of the phage fd coat protein. DNA fragments containing these sequences are excised by cleavage with restriction enzymes from the DNA isolated from transducing phages that carry the lac or trp operons, or from
5 the DNA of phage lambda or fd. Those fragments are then manipulated in order to obtain a limited population of molecules such that the essential controlling sequences can be joined very close to, or in juxtaposition with, the initiation codon of the coding sequence.

10 The fusion product is then inserted into a cloning vehicle for transformation or transfection of the appropriate hosts and the level of antigen production is measured. Cells giving the most efficient expression may be thus selected. Alternatively, cloning vehicles carrying the lac, trp or
15 lambda P_L control system attached to an initiation codon may be employed and fused to a fragment containing a sequence coding for a MN protein or polypeptide such that the gene or sequence is correctly translated from the initiation codon of the cloning vehicle.

20 The phrase "recombinant nucleic acid molecule" is herein defined to mean a hybrid nucleotide sequence comprising at least two nucleotide sequences, the first sequence not normally being found together in nature with the second.

The phrase "expression control sequence" is herein
25 defined to mean a sequence of nucleotides that controls and regulates expression of structural genes when operatively linked to those genes.

The following are representative examples of genetically engineering MN proteins of this invention. The
30 descriptions are exemplary and not meant to limit the invention in any way.

Expression of MN 20-19 Protein

A representative, recombinantly produced MN protein of this invention is the MN 20-19 protein which, when produced
35 in baculovirus-infected Sf9 cells [*Spodoptera frugiperda* cells; Clontech; Palo Alto, CA (USA)], is glycosylated. The MN 20-19 protein misses the putative signal peptide (AAs 1-37)

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of SEQ. ID. NO.: 6 (Figure 1), has a methionine (Met) at the N-terminus for expression, and a Leu-Glu-His-His-His-His-His [SEQ. ID NO.: 22] added to the C-terminus for purification.

5 In order to insert the portion of the MN coding sequence for the GEX-3X-MN fusion protein into alternate expression systems, a set of primers for PCR was designed. The primers were constructed to provide restriction sites at each end of the coding sequence, as well as in-frame start and
10 stop codons. The sequences of the primers, indicating restriction enzyme cleavage sites and expression landmarks, are shown below.

Primer #20:N-terminus

15 5' GTCGCTAGCTCCATGGGTCATATGCAGAGGTTGCCCCGGATGCAG 3'
NheI NcoI NdeI Translation start MN cDNA #1 [SEQ. ID. NO. 17]

Primer #19:C-terminus

20 5' GAAGATCTCTTACTCGAGCATTCTCCAAGATCCAGCCTCTAGG 3'
BglII XhoI Translation stop MN cDNA [SEQ. ID. NO. 18]

The SEQ. ID. NOS.: 17 and 18 primers were used to amplify the MN coding sequence present in the GEX-3X-MN vector using standard PCR techniques. The resulting PCR product (termed MN 20-19) was electrophoresed on a 0.5% agarose/1X TBE gel; the
25 1.3 kb band was excised; and the DNA recovered using the Gene Clean II kit according to the manufacturer's instructions [Bio101; LaJolla, CA (USA)].

30 MN 20-19 and plasmid pET-22b were cleaved with the restriction enzymes NdeI and XhoI, phenol-chloroform extracted, and the appropriate bands recovered by agarose gel electrophoresis as above. The isolated fragments were ethanol co-precipitated at a vector:insert ratio of 1:4. After

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resuspension, the fragments were ligated using T4 DNA ligase. The resulting product was used to transform competent Novablu E. coli cells [Novagen, Inc.]. Plasmid mini-preps [Magic Minipreps; Promega] from the resultant ampicillin resistant colonies were screened for the presence of the correct insert by restriction mapping. Insertion of the gene fragment into the pET-22b plasmid using the NdeI and XhoI sites added a 6-histidine tail to the protein that could be used for affinity isolation.

10 To prepare MN 20-19 for insertion into the baculovirus expression system, the MN 20-19 gene fragment was excised from pET-22b using the restriction endonucleases XbaI and PvuI. The baculovirus shuttle vector pBacPAK8 [Clontech] was cleaved with XbaI and PacI. The desired fragments (1.3 kb
15 for MN 20-19 and 5.5 kb for pBacPAK8) were isolated by agarose gel electrophoresis, recovered using Gene Clean II, and co-precipitated at an insert:vector ratio of 2.4:1.

After ligation with T4 DNA ligase, the DNA was used to transform competent NM522 E. coli cells (Stratagene).

20 Plasmid mini-preps from resultant ampicillin resistant colonies were screened for the presence of the correct insert by restriction mapping. Plasmid DNA from an appropriate colony and linearized BacPAK6 baculovirus DNA [Clontech] were used to transform Sf9 cells by standard techniques.
25 Recombination produced BacPAK viruses carrying the MN 20-19 sequence. Those viruses were plated onto Sf9 cells and overlaid with agar.

Plaques were picked and plated onto Sf9 cells. The conditioned media and cells were collected. A small aliquot
30 of the conditioned media was set aside for testing. The cells were extracted with PBS with 1% Triton X100.

The conditioned media and the cell extracts were dot blotted onto nitrocellulose paper. The blot was blocked with 5% non-fat dried milk in PBS. Mab M75 were used to detect the
35 MN 20-19 protein in the dot blots. A rabbit anti-mouse Ig-HRP was used to detect bound Mab M75. The blots were developed with TMB/H₂O₂ with a membrane enhancer [KPL; Gaithersburg, MD (USA)]. Two clones producing the strongest reaction on the

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dot blots were selected for expansion. One was used to produce MN 20-19 protein in High Five cells [Invitrogen Corp., San Diego, CA (USA); BTI-TN-5BI-4; derived from Trichoplusia ni egg cell homogenate]. MN 20-19 protein was purified from the conditioned media from the virus infected High Five cells.

The MN 20-19 protein was purified from the conditioned media by immunoaffinity chromatography. 6.5 mg of Mab M75 was coupled to 1 g of Tresyl activated Toyopearl™ [Tosoh, Japan (#14471)]. Approximately 150 ml of the conditioned media was run through the M75-Toyopearl column. The column was washed with PBS, and the MN 20-19 protein was eluted with 1.5 M MgCl. The eluted protein was then dialyzed against PBS.

Fusion Proteins with C-Terminal Part Including Transmembrane Region Replaced by Fc or PA

MN fusion proteins in which the C terminal part including the transmembrane region is replaced by the Fc fragment of human IgG or by Protein A were constructed. Such fusion proteins are useful to identify MN binding protein(s). In such MN chimaeras, the whole N-terminal part of MN is accessible to interaction with heterologous proteins, and the C terminal tag serves for simple detection and purification of protein complexes.

Fusion Protein MN-PA (Protein A)

In a first step, the 3' end of the MN cDNA encoding the transmembrane region of the MN protein was deleted. The plasmid pFLMN (e.g. pBluescript with full length MN cDNA) was cleaved by *EcoRI* and blunt ended by *S1* nuclease. Subsequent cleavage by *SacI* resulted in the removal of the *EcoRI-SacI* fragment. The deleted fragment was then replaced by a Protein A coding sequence that was derived from plasmid pEZZ (purchased from Pharmacia), which had been cleaved with *RsaI* and *SacI*. The obtained MN-PA construct was subcloned into a eukaryotic expression vector pSG5C (described in Example 3), and was then ready for transfection experiments.

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Fusion Protein MN-Fc

The cloning of the fusion protein MN-Fc was rather complicated due to the use of a genomic clone containing the Fc fragment of human IgG which had a complex structure in that it contained an enhancer, a promoter, exons and introns. Moreover, the complete sequence of the clone was not available. Thus, it was necessary to ensure the correct in-phase splicing and fusion of MN to the Fc fragment by the addition of a synthetic splice donor site (SSDS) designed according to the splicing sequences of the MN gene.

The construction procedure was as follows:

1. Plasmid pMH4 (e.g. pSV2gpt containing a genomic clone of the human IgG Fc region) was cleaved by *Bam*HI in order to get a 13 kb fragment encoding Fc. [In pSV2gpt, the *E. coli* xanthine-guanine phosphoribosyl transferase gene (*gpt*) is expressed using the SV40 early promoter (P_E) located in the SV40 origin, the SV40 small T intron, and the SV40 polyadenylation site.]

2. At the same time, plasmid pFLMN (with full length MN cDNA) was cleaved by *Sal*I-*Eco*RI. The released fragment was purified and ligated with a synthetic adapter *Eco*RI-*Bgl*III containing a synthetic splice donor site (SSDS).

3. Simultaneously, the plasmid pBKCMV was cleaved by *Sal*I-*Bam*HI. Then advantage was taken of the fact that the *Bam*HI cohesive ends (of the Fc coding fragment) are compatible with the *Bgl*III ends of the SSDS, and Fc was ligated to MN. The MN-Fc ligation product was then inserted into pBKCMV by directional cloning through the *Sal*I and *Bam*HI sites.

Verification of the correct orientation and in-phase fusion of the obtained MN-Fc chimaeric clones was problematic in that the sequence of Fc was not known. Thus, functional constructs are selected on the basis of results of transient eukaryotic expression analyses.

Synthetic and Biologic Production of MN Proteins and Polypeptides

MN proteins and polypeptides of this invention may be prepared not only by recombinant means but also by

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synthetic and by other biologic means. Synthetic formation of the polypeptide or protein requires chemically synthesizing the desired chain of amino acids by methods well known in the art. Exemplary of other biologic means to prepare the desired polypeptide or protein is to subject to selective proteolysis a longer MN polypeptide or protein containing the desired amino acid sequence; for example, the longer polypeptide or protein can be split with chemical reagents or with enzymes.

Chemical synthesis of a peptide is conventional in the art and can be accomplished, for example, by the Merrifield solid phase synthesis technique [Merrifield, J., Am. Chem. Soc., 85: 2149-2154 (1963); Kent et al., Synthetic Peptides in Biology and Medicine, 29 f.f. eds. Alitalo et al., (Elsevier Science Publishers 1985); and Haug, J.D., "Peptide Synthesis and Protecting Group Strategy", American Biotechnology Laboratory, 5(1): 40-47 (Jan/Feb. 1987)].

Techniques of chemical peptide synthesis include using automatic peptide synthesizers employing commercially available protected amino acids, for example, Biosearch [San Rafael, CA (USA)] Models 9500 and 9600; Applied Biosystems, Inc. [Foster City, CA (USA)] Model 430; Milligen [a division of Millipore Corp.; Bedford, MA (USA)] Model 9050; and Du Pont's RAMP (Rapid Automated Multiple Peptide Synthesis) [Du Pont Compass, Wilmington, DE (USA)].

25 Regulation of MN Expression and MN Promoter

MN appears to be a novel regulatory protein that is directly involved in the control of cell proliferation and in cellular transformation. In HeLa cells, the expression of MN is positively regulated by cell density. Its level is increased by persistent infection with LCMV. In hybrid cells between HeLa and normal fibroblasts, MN expression correlates with tumorigenicity. The fact that MN is not present in nontumorigenic hybrid cells (CGL1), but is expressed in a tumorigenic segregant lacking chrom some 11, indicates that MN is negatively regulated by a putative suppressor in chromosome 11.

Evidence supporting the regulatory role of MN protein was found in the generation of stable transfectants of NIH 3T3 cells that constitutively express MN protein as described in Example 3. As a consequence of MN expression, the NIH 3T3 cells acquired features associated with a transformed phenotype: altered morphology, increased saturation density, proliferative advantage in serum-reduced media, enhanced DNA synthesis and capacity for anchorage-independent growth. Further, as shown in Example 4, flow cytometric analyses of asynchronous cell populations indicated that the expression of MN protein leads to accelerated progression of cells through G1 phase, reduction of cell size and the loss of capacity for growth arrest under inappropriate conditions. Also, Example 4 shows that MN expressing cells display a decreased sensitivity to the DNA damaging drug mitomycin C.

Nontumorigenic human cells, CGL1 cells, were also transfected with the full-length MN cDNA. The same pSG5C-MN construct in combination with pSV2neo plasmid as used to transfect the NIH 3T3 cells (Example 3) was used. Also the protocol was the same except that the G418 concentration was increased to 1000 μ g/ml.

Out of 15 MN-positive clones (tested by SP-RIA and Western blotting), 3 were chosen for further analysis. Two MN-negative clones isolated from CGL1 cells transfected with empty plasmid were added as controls. Initial analysis indicates that the morphology and growth habits of MN-transfected CGL1 cells are not changed dramatically, but their proliferation rate and plating efficiency is increased.

MN cDNA and promoter. When the promoter region from the MN genomic clone, isolated as described above, was linked to MN cDNA and transfected into CGL1 hybrid cells, expression of MN protein was detectable immediately after selection. However, then it gradually ceased, indicating thus an action of a feedback regulator. The putative regulatory element appeared to be acting via the MN promoter, because when the full-length cDNA (not containing the promoter) was used for transfection, no similar effect was observed.

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An "antisense" MN cDNA/MN promoter construct was used to transfect CGL3 cells. The effect was the opposite of that of the CGL1 cells transfected with the "sense" construct. Whereas the transfected CGL1 cells formed colonies several
5 times larger than the control CGL1, the transfected CGL3 cells formed colonies much smaller than the control CGL3 cells.

For those experiments, the part of the promoter region that was linked to the MN cDNA through a BamHI site was derived from a NcoI - BamHI fragment of the MN genomic clone
10 [Bd3] and represents a region a few hundred bp upstream from the transcription initiation site. After the ligation, the joint DNA was inserted into a pBK-CMV expression vector [Stratagene]. The required orientation of the inserted
15 sequence was ensured by directional cloning and subsequently verified by restriction analysis. The tranfection procedure was the same as used in transfecting the NIH 3T3 cells (Example 3), but co-transfection with the pSV2neo plasmid was not necessary since the neo selection marker was already
included in the pBK-CMV vector.

20 After two weeks of selection in a medium containing G418, remarkable differences between the numbers and sizes of the colonies grown were evident as noted above. Immediately following the selection and cloning, the MN-transfected CGL1 and CGL3 cells were tested by SP-RIA for expression and
25 repression of MN, respectively. The isolated transfected CGL1 clones were MN positive (although the level was lower than obtained with the full-length cDNA), whereas MN protein was almost absent from the transfected CGL3 clones. However, in subsequent passages, the expression of MN in transfected CGL1
30 cells started to cease, and was then blocked perhaps evidencing a control feedback mechanism.

As a result of the very much lowered proliferation of the transfected CGL3 cells, it was difficult to expand the majority of cloned cells (according to SP-RIA, those with the
35 lowest levels of MN), and they were lost during passaging. However, some clones overcame that problem and again expressed MN. It is possible that once those cells reached a higher quantity, that the level of endogenously produced MN mRNA

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increased over the amount of ectopically expressed antisense mRNA.

Transformation and Reversion

As illustrated in Examples 3 and 4, vertebrate cells
5 transfected with MN cDNA in suitable vectors show striking
morphologic transformation. Transformed cells may be very
small, densely packed, slowly growing, with basophilic
cytoplasm and enlarged Golgi apparatus. However, it has been
found that transformed clones revert over time, for example,
10 within 3-4 weeks, to nearly normal morphology, even though the
cells may be producing MN protein at high levels. MN protein
is biologically active even in yeast cells; depending upon the
level of its expression, it stimulates or retards their growth
and induces morphologic alterations.

15 Full-length MN cDNA was inserted into pGD, a MLV-
derived vector, which together with standard competent MLV
(murine leukemia virus), forms an infectious, transmissible
complex [pGD-MN + MLV]. That complex also transforms
vertebrate cells, such as, NIH 3T3 cells and mouse embryo
20 fibroblasts BALB/c, which also revert to nearly normal
morphology. Such revertants again contain MN protein and
produce the [pGD-MN + MLV] artificial virus complex, which
retains its transforming capacity. Thus, reversion of MN-
transformed cells is apparently not due to a loss, silencing
25 or mutation of MN cDNA, but may be the result of the
activation of suppressor gene(s).

Nucleic Acid Probes and Test Kits

Nucleic acid probes of this invention are those
comprising sequences that are complementary or substantially
30 complementary to the MN cDNA sequence shown in Figure 1 or to
other MN gene sequences, such as, the complete genomic
sequence of Figure 3a-d [SEQ. ID. NO.: 5] and the putative
promoter sequence [SEQ. ID. NO.: 27 of Figure 6]. The phrase
"substantially complementary" is defined herein to have the
35 meaning as it is well understood in the art and, thus, used in
the context of standard hybridization conditions. The

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stringency of hybridization conditions can be adjusted to control the precision of complementarity. Two nucleic acids are, for example, substantially complementary to each other, if they hybridize to each other under stringent hybridization conditions.

Stringent hybridization conditions are considered herein to conform to standard hybridization conditions understood in the art to be stringent. For example, it is generally understood that stringent conditions encompass relatively low salt and/or high temperature conditions, such as provided by 0.02 M to 0.15 M NaCl at temperatures of 50°C to 70°C. Less stringent conditions, such as, 0.15 M to 0.9 M salt at temperatures ranging from 20°C to 55°C can be made more stringent by adding increasing amounts of formamide, which serves to destabilize hybrid duplexes as does increased temperature.

Exemplary stringent hybridization conditions are described in Sambrook et al., Molecular Cloning: A Laboratory Manual, pages 1.91 and 9.47-9.51 (Second Edition, Cold Spring Harbor Laboratory Press; Cold Spring Harbor, NY; 1989); Maniatis et al., Molecular Cloning: A Laboratory Manual, pages 387-389 (Cold Spring Harbor Laboratory; Cold Spring Harbor, NY; 1982); Tsuchiya et al., Oral Surgery, Oral Medicine, Oral Pathology, 71(6): 721-725 (June 1991).

Preferred nucleic acid probes of this invention are fragments of the isolated nucleic acid sequences that encode MN proteins or polypeptides according to this invention. Preferably those probes are composed of at least twenty-nine nucleotides, more preferably, fifty nucleotides.

Nucleic acid probes of this invention need not hybridize to a coding region of MN. For example, nucleic acid probes of this invention may hybridize partially or wholly to a non-coding region of the genomic sequence shown in Figure 3a-d [SEQ. ID. NO.: 5]. Conventional technology can be used to determine whether fragments of SEQ. ID. NO.: 5 or related nucleic acids are useful to identify MN nucleic acid sequences. [See, for example, Bentor and Davis, supra and Fuscoe et al., supra.]

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Areas of homology of the MN nt sequence to other non-MN nt sequences are indicated above. In general, nucleotide sequences that are not in the Alu or LTR-like regions, of preferably 29 bases or more, or still more preferably of 50 bases or more, can be routinely tested and screened and found to hybridize under stringent conditions to only MN nucleotide sequences. Further, not all homologies within the Alu-like MN genomic sequences are so close to Alu repeats as to give a hybridization signal under stringent hybridization conditions. The percent of homology between MN Alu-like regions and a standard Alu-J sequence are indicated as follows:

<u>Region of Homology within</u>			
	<u>MN Genomic Sequence</u>	<u>SEO.</u>	<u>% Homology to</u>
15	<u>[SEO. ID. NO.: 5;</u>	<u>ID.</u>	<u>Entire Alu-J</u>
	<u>Figure 3a-d]</u>	<u>NOS.</u>	<u>Sequence</u>
	921-1212	59	89.1%
	2370-2631	60	78.6%
	4587-4880	61	90.1%
20	6463-6738	62	85.4%
	7651-7939	63	91.0%
	9020-9317	64	69.8%
			<u>% Homology to</u>
			<u>One Half of</u>
			<u>Alu-J Sequence</u>
	8301-8405	65	88.8%
	10040-10122	66	73.2%

25 Nucleic acid probes of this invention can be used to detect MN DNA and/or RNA, and thus can be used to test for the presence or absence of MN genes, and amplification(s), mutation(s) or genetic rearrangements of MN genes in the cells of a patient. For example, overexpression of an MN gene may
30 be detected by Northern blotting and RNase protection analysis using probes of this invention. Gene alterations, as amplifications, translocations, inversions, and deletions among others, can be detected by using probes of this

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invention for in situ hybridization to chromosomes from a patient's cells, whether in metaphase spreads or interphase nuclei. Southern blotting could also be used with the probes of this invention to detect amplifications or deletions of MN genes. Restriction Fragment Length Polymorphism (RFLP) analysis using said probes is a preferred method of detecting gene alterations, mutations and deletions. Said probes can also be used to identify MN proteins and/or polypeptides as well as homologs or near homologs thereto by their hybridization to various mRNAs transcribed from MN genes in different tissues.

Probes of this invention thus can be useful diagnostically/prognostically. Said probes can be embodied in test kits, preferably with appropriate means to enable said probes when hybridized to an appropriate MN gene or MN mRNA target to be visualized. Such samples include tissue specimens including smears, body fluids and tissue and cell extracts.

PCR Assays

To detect relatively large genetic rearrangements, hybridization tests can be used. To detect relatively small genetic rearrangements, as, for example, small deletions or amplifications, or point mutations, PCR would preferably be used. [U.S. Patent Nos. 4,800,159; 4,683,195; 4,683,202; and Chapter 14 of Sambrook et al., Molecular Cloning: A Laboratory Manual, supra]

An exemplary assay would use cellular DNA from normal and cancerous cells, which DNA would be isolated and amplified employing appropriate PCR primers. The PCR products would be compared, preferably initially, on a sizing gel to detect size changes indicative of certain genetic rearrangements. If no differences in sizes are noted, further comparisons can be made, preferably using, for example, PCR-single-strand conformation polymorphism (PCR-SSCP) assay or a denaturing gradient gel electrophoretic assay. [See, for example, Hayashi, K., "PCR-SSCP: A Simple and Sensitive Method for Detection of Mutations in the Genomic DNA," in PCR

Methods and Applications, 1: 34-38 (1991); and Meyers et al., "Detection and Localization of Single Base Changes by Denaturing Gradient Gel Electrophoresis," Methods in Enzymology, 155: 501 (1987).]

5

Assays

Assays according to this invention are provided to detect and/or quantitate MN antigen or MN-specific antibodies in vertebrate samples, preferably mammalian samples, more preferably human samples. Such samples include tissue
10 specimens, body fluids, tissue extracts and cell extracts. MN antigen may be detected by immunoassay, immunohistochemical staining, immunoelectron and scanning microscopy using immunogold among other techniques.

Preferred tissue specimens to assay by
15 immunohistochemical staining include cell smears, histological sections from biopsied tissues or organs, and imprint preparations among other tissue samples. Such tissue specimens can be variously maintained, for example, they can be fresh, frozen, or formalin-, alcohol- or acetone- or
20 otherwise fixed and/or paraffin-embedded and deparaffinized. Biopsied tissue samples can be, for example, those samples removed by aspiration, bite, brush, cone, chorionic villus, endoscopic, excisional, incisional, needle, percutaneous punch, and surface biopsies, among other biopsy techniques.

25 Preferred cervical tissue specimens include cervical smears, conization specimens, histologic sections from hysterectomy specimens or other biopsied cervical tissue samples. Preferred means of obtaining cervical smears include routine swab, scraping or cytobrush techniques, among other
30 means. More preferred are cytobrush or swab techniques. Preferably, cell smears are made on microscope slides, fixed, for example, with 55% EtOH or an alcohol based spray fixative and air-dried.

Papanicolaou-stained cervical smears (Pap smears)
35 can be screened by the methods of this invention, for example, for retrospective studies. Preferably, Pap smears would be decolorized and re-stained with labeled antibodies against MN

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antigen. Also archival specimens, for example, matched smears and biopsy and/or tumor specimens, can be used for retrospective studies. Prospective studies can also be done with matched specimens from patients that have a higher than
5 normal risk of exhibiting abnormal cervical cytopathology.

Preferred samples in which to assay MN antigen by, for example, Western blotting or radioimmunoassay, are tissue and/or cell extracts. However, MN antigen may be detected in body fluids, which can include among other fluids: blood,
10 serum, plasma, semen, breast exudate, saliva, tears, sputum, mucous, urine, lymph, cytosols, ascites, pleural effusions, amniotic fluid, bladder washes, bronchioalveolar lavages and cerebrospinal fluid. It is preferred that the MN antigen be concentrated from a larger volume of body fluid before
15 testing. Preferred body fluids to assay would depend on the type of cancer for which one was testing, but in general preferred body fluids would be breast exudate, pleural effusions and ascites.

MN-specific antibodies can be bound by serologically
20 active MN proteins/polypeptides in samples of such body fluids as blood, plasma, serum, lymph, mucous, tears, urine, spinal fluid and saliva; however, such antibodies are found most usually in blood, plasma and serum, preferably in serum. Correlation of the results from the assays to detect and/or
25 quantitate MN antigen and MN-specific antibodies reactive therewith, provides a preferred profile of the disease condition of a patient.

The assays of this invention are both diagnostic and/or prognostic, i.e., diagnostic/prognostic. The term
30 "diagnostic/ prognostic" is herein defined to encompass the following processes either individually or cumulatively depending upon the clinical context: determining the presence of disease, determining the nature of a disease, distinguishing one disease from another, forecasting as to the
35 probable outcome of a disease state, determining the prospect as to recovery from a disease as indicated by the nature and symptoms of a case, monitoring the disease status of a patient, monitoring a patient for recurrence of disease,

and/or determining the preferred therapeutic regimen for a patient. The diagnostic/prognostic methods of this invention are useful, for example, for screening populations for the presence of neoplastic or pre-neoplastic disease, determining
5 the risk of developing neoplastic disease, diagnosing the presence of neoplastic and/or pre-neoplastic disease, monitoring the disease status of patients with neoplastic disease, and/or determining the prognosis for the course of neoplastic disease. For example, it appears that the
10 intensity of the immunostaining with MN-specific antibodies may correlate with the severity of dysplasia present in samples tested.

The present invention is useful for screening for the presence of a wide variety of neoplastic diseases as
15 indicated above. The invention provides methods and compositions for evaluating the probability of the presence of malignant or pre-malignant cells, for example, in a group of cells freshly removed from a host. Such an assay can be used to detect tumors, quantitate their growth, and help in the
20 diagnosis and prognosis of disease. The assays can also be used to detect the presence of cancer metastasis, as well as confirm the absence or removal of all tumor tissue following surgery, cancer chemotherapy and/or radiation therapy. It can further be used to monitor cancer chemotherapy and tumor
25 reappearance.

The presence of MN antigen or antibodies can be detected and/or quantitated using a number of well-defined diagnostic assays. Those in the art can adapt any of the conventional immunoassay formats to detect and/or quantitate
30 MN antigen and/or antibodies.

Many formats for detection of MN antigen and MN-specific antibodies are, of course available. Those can be Western blots, ELISAs, RIAs, competitive EIA or dual antibody sandwich assays, immunohistochemical staining, among other
35 assays all commonly used in the diagnostic industry. In such immunoassays, the interpretation of the results is based on the assumption that the antibody or antibody combination will

not cross-react with other proteins and protein fragments present in the sample that are unrelated to MN.

Representative of one type of ELISA test for MN antigen is a format wherein a microtiter plate is coated with
5 antibodies made to MN proteins/polypeptides or antibodies made to whole cells expressing MN proteins, and to this is added a patient sample, for example, a tissue or cell extract. After a period of incubation permitting any antigen to bind to the antibodies, the plate is washed and another set of anti-MN
10 antibodies which are linked to an enzyme is added, incubated to allow reaction to take place, and the plate is then rewashed. Thereafter, enzyme substrate is added to the microtiter plate and incubated for a period of time to allow the enzyme to work on the substrate, and the adsorbance of the
15 final preparation is measured. A large change in absorbance indicates a positive result.

It is also apparent to one skilled in the art of immunoassays that MN proteins and/or polypeptides can be used to detect and/or quantitate the presence of MN antigen in the
20 body fluids, tissues and/or cells of patients. In one such embodiment, a competition immunoassay is used, wherein the MN protein/polypeptide is labeled and a body fluid is added to compete the binding of the labeled MN protein/polypeptide to antibodies specific to MN protein/polypeptide.

25 In another embodiment, an immunometric assay may be used wherein a labeled antibody made to a MN protein or polypeptide is used. In such an assay, the amount of labeled antibody which complexes with the antigen-bound antibody is directly proportional to the amount of MN antigen in the
30 sample.

A representative assay to detect MN-specific antibodies is a competition assay in which labeled MN protein/polypeptide is precipitated by antibodies in a sample, for example, in combination with monoclonal antibodies
35 recognizing MN proteins/polypeptides. One skilled in the art could adapt any of the conventional immunoassay formats to detect and/or quantitate MN-specific antibodies. Detection of the binding of said antibodies to said MN protein/polypeptide

could be by many ways known to those in the art, e.g., in humans with the use of anti-human labeled IgG.

An exemplary immunoassay method of this invention to detect and/or quantitate MN antigen in a vertebrate sample
5 comprises the steps of:

a) incubating said vertebrate sample with one or more sets of antibodies (an antibody or antibodies) that bind to MN antigen wherein one set is labeled or otherwise detectable;

10 b) examining the incubated sample for the presence of immune complexes comprising MN antigen and said antibodies.

Another exemplary immunoassay method according to this invention is that wherein a competition immunoassay is used to detect and/or quantitate MN antigen in a vertebrate
15 sample and wherein said method comprises the steps of:

a) incubating a vertebrate sample with one or more sets of MN-specific antibodies and a certain amount of a labeled or otherwise detectable MN protein/polypeptide wherein said MN protein/ polypeptide competes for binding to said
20 antibodies with MN antigen present in the sample;

b) examining the incubated sample to determine the amount of labeled/detectable MN protein/polypeptide bound to said antibodies; and

c) determining from the results of the examination
25 in step b) whether MN antigen is present in said sample and/or the amount of MN antigen present in said sample.

Once antibodies (including biologically active antibody fragments) having suitable specificity have been prepared, a wide variety of immunological assay methods are
30 available for determining the formation of specific antibody-antigen complexes. Numerous competitive and non-competitive protein binding assays have been described in the scientific and patent literature, and a large number of such assays are commercially available. Exemplary
35 immunoassays which are suitable for detecting a serum antigen include those described in U.S. Patent Nos. 3,984,533; 3,996,345; 4,034,074; and 4,098,876.

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Antibodies employed in assays may be labeled or unlabeled. Unlabeled antibodies may be employed in agglutination; labeled antibodies may be employed in a wide variety of assays, employing a wide variety of labels.

5 Suitable detection means include the use of labels such as radionuclides, enzymes, coenzymes, fluorescers, chemilumescers, chromogens, enzyme substrates or co-factors, enzyme inhibitors, free radicals, particles, dyes and the like. Such labeled reagents may be used in a variety of well
10 known assays, such as radioimmunoassays, enzyme immunoassays, e.g., ELISA, fluorescent immunoassays, and the like. See for example, U.S. Patent Nos. 3,766,162; 3,791,932; 3,817,837; and 4,233,402.

Immunoassay Test Kits

15 The above outlined assays can be embodied in test kits to detect and/or quantitate MN antigen and/or MN-specific antibodies (including biologically active antibody fragments). Kits to detect and/or quantitate MN antigen can comprise MN
20 protein(s)/polypeptides(s) and/or MN-specific antibodies, polyclonal and/or monoclonal. Such diagnostic/prognostic test kits can comprise one or more sets of antibodies, polyclonal and/or monoclonal, for a sandwich format wherein antibodies recognize epitopes on the MN antigen, and one set is
appropriately labeled or is otherwise detectable.

25 Test kits for an assay format wherein there is competition between a labeled (or otherwise detectable) MN protein/polypeptide and MN antigen in the sample, for binding to an antibody, can comprise the combination of the labeled
protein/polypeptide and the antibody in amounts which provide
30 for optimum sensitivity and accuracy.

 Test kits for MN-specific antibodies preferably comprise labeled/detectable MN proteins(s) and/or
polypeptides(s), and may comprise other components as
necessary, such as, controls, buffers, diluents and
35 detergents. Such test kits can have other appropriate formats for conventional assays.

A kit for use in an enzyme-immunoassay typically includes an enzyme-labelled reagent and a substrate for the enzyme. The enzyme can, for example, bind either an MN-specific antibody of this invention or to an antibody to such an MN-specific antibody.

Preparation of MN-Specific Antibodies

The term "antibodies" is defined herein to include not only whole antibodies but also biologically active fragments of antibodies, preferably fragments containing the antigen binding regions. Such antibodies may be prepared by conventional methodology and/or by genetic engineering. Antibody fragments may be genetically engineered, preferably from the variable regions of the light and/or heavy chains (V_H and V_L), including the hypervariable regions, and still more preferably from both the V_H and V_L regions. For example, the term "antibodies" as used herein comprehends polyclonal and monoclonal antibodies and biologically active fragments thereof including among other possibilities "univalent" antibodies [Glennie et al., Nature, 295: 712 (1982)]; Fab proteins including Fab' and $F(ab')_2$ fragments whether covalently or non-covalently aggregated; light or heavy chains alone, preferably variable heavy and light chain regions (V_H and V_L regions), and more preferably including the hypervariable regions [otherwise known as the complementarity determining regions (CDRs) of said V_H and V_L regions]; F_c proteins; "hybrid" antibodies capable of binding more than one antigen; constant-variable region chimeras; "composite" immunoglobulins with heavy and light chains of different origins; "altered" antibodies with improved specificity and other characteristics as prepared by standard recombinant techniques and also by oligonucleotide-directed mutagenesis techniques [Dalbadie-McFarland et al., PNAS (USA), 79: 6409 (1982)].

It may be preferred for therapeutic and/or imaging uses that the antibodies be biologically active antibody fragments, preferably genetically engineered fragments, more preferably genetically engineered fragments from the V_H and/or

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V_L regions, and still more preferably comprising the hypervariable regions thereof.

There are conventional techniques for making polyclonal and monoclonal antibodies well-known in the immunoassay art. Immunogens to prepare MN-specific antibodies include MN proteins and/or polypeptides, preferably purified, and MX-infected tumor line cells, for example, MX-infected HeLa cells, among other immunogens.

Anti-peptide antibodies are also made by conventional methods in the art as described in European Patent Publication No. 44,710 (published Jan. 27, 1982). Briefly, such anti-peptide antibodies are prepared by selecting a peptide from an MN amino acid sequence as from Figure 1, chemically synthesizing it, conjugating it to an appropriate immunogenic protein and injecting it into an appropriate animal, usually a rabbit or a mouse; then, either polyclonal or monoclonal antibodies are made, the latter by a Kohler-Milstein procedure, for example.

Besides conventional hybridoma technology, newer technologies can be used to produce antibodies according to this invention. For example, the use of the PCR to clone and express antibody V-genes and phage display technology to select antibody genes encoding fragments with binding activities has resulted in the isolation of antibody fragments from repertoires of PCR amplified V-genes using immunized mice or humans. [Marks et al., BioTechnology, 10: 779 (July 1992) for references; Chiang et al., BioTechniques, 7(4): 360 (1989); Ward et al., Nature, 341: 544 (Oct. 12, 1989); Marks et al., J. Mol. Biol., 222: 581 (1991); Clackson et al., Nature, 352: (15 August 1991); and Mullinax et al., PNAS (USA), 87: 8095 (Oct. 1990).]

Descriptions of preparing antibodies, which term is herein defined to include biologically active antibody fragments, by recombinant techniques can be found in U.S. Patent No. 4,816,567 (issued March 28, 1989); European Patent Application Publication Number (EP) 338,745 (published Oct. 25, 1989); EP 368,684 (published June 16, 1990); EP 239,400 (published September 30, 1987); WO 90/14424 (published Nov.

29, 1990); WO 90/14430 (published May 16, 1990); Huse et al., Science, 246: 1275 (Dec. 8, 1989); Marks et al., BioTechnology, 10: 779 (July 1992); La Sastry et al., PNAS (USA), 86: 5728 (August 1989); Chiang et al., BioTechniques, 7(40): 360 (1989); Orlandi et al., PNAS (USA), 86: 3833 (May 1989); Ward et al. Nature, 341: 544 (October 12, 1989); Marks et al., J. Mol. Biol., 222: 581 (1991); and Hoogenboom et al., Nucleic Acids Res., 19(15): 4133 (1991).

Representative Mabs

10 Monoclonal antibodies for use in the assays of this invention may be obtained by methods well known in the art for example, Galfre and Milstein, "Preparation of Monoclonal Antibodies: Strategies and Procedures," in Methods in Enzymology: Immunochemical Techniques, 73: 1-46 [Langone and 15 Vanatis (eds); Academic Press (1981)]; and in the classic reference, Milstein and Kohler, Nature, 256: 495-497 (1975).]

Although representative hybridomas of this invention are formed by the fusion of murine cell lines, human/human hybridomas [Olsson et al., PNAS (USA), 77: 5429 (1980)] and 20 human/murine hybridomas [Schlom et al., PNAS (USA), 77: 6841 (1980); Shearman et al. J. Immunol., 146: 928-935 (1991); and Gorman et al., PNAS (USA), 88: 4181-4185 (1991)] can also be prepared among other possibilities. Such humanized monoclonal antibodies would be preferred monoclonal antibodies for 25 therapeutic and imaging uses.

Monoclonal antibodies specific for this invention can be prepared by immunizing appropriate mammals, preferably rodents, more preferably rabbits or mice, with an appropriate immunogen, for example, MaTu-infected HeLa cells, MN fusion 30 proteins, or MN proteins/polypeptides attached to a carrier protein if necessary. Exemplary methods of producing antibodies of this invention are described below.

The monoclonal antibodies useful according to this invention to identify MN proteins/polypeptides can be labeled 35 in any conventional manner, for example, with enzymes such as horseradish peroxidase (HRP), fluorescent compounds, or with radioactive isotopes such as, ¹²⁵I, among other labels. A

preferred label, according to this invention is ^{125}I , and a preferred method of labeling the antibodies is by using chloramine-T [Hunter, W.M., "Radioimmunoassay," In: Handbook of Experimental Immunology, pp. 14.1-14.40 (D.W. Weir ed.; Blackwell, Oxford/London/Edinburgh/Melbourne; 1978)].

Representative mabs of this invention include Mabs M75, MN9, MN12 and MN7 described below. Monoclonal antibodies of this invention serve to identify MN proteins/polypeptides in various laboratory diagnostic tests, for example, in tumor cell cultures or in clinical samples.

Mabs Prepared Against HeLa Cells

MAb M75. Monoclonal antibody M75 (MAb M75) is produced by mouse lymphocytic hybridoma VU-M75, which was initially deposited in the Collection of Hybridomas at the Institute of Virology, Slovak Academy of Sciences (Bratislava, Czechoslovakia) and was deposited under ATCC Designation HB 11128 on September 17, 1992 at the American Type Culture Collection (ATCC) in Rockville, MD (USA). The production of hybridoma VU-M75 is described in Zavada et al., WO 93/18152.

Mab M75 recognizes both the nonglycosylated GEX-3X-MN fusion protein and native MN protein as expressed in CGL3 cells equally well. Mab M75 was shown by epitope mapping to be reactive with the epitope represented by the amino acid sequence from AA 62 to AA 67 [SEQ. ID. NO.: 10] of the MN protein shown in Figure 1.

Mabs Prepared Against Fusion Protein GEX-3X-MN

Monoclonal antibodies of this invention were also prepared against the MN glutathione S-transferase fusion protein (GEX-3X-MN). BALB/C mice were immunized intraperitoneally according to standard procedures with the GEX-3X-MN fusion protein in Freund's adjuvant. Spleen cells of the mice were fused with SP/20 myeloma cells [Milstein and Kohler, supra].

Tissue culture media from the hybridomas were screened against CGL3 and CGL1 membrane extracts in an ELISA employing HRP labelled rabbit anti-mouse. The membrane

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extracts were coated onto microtiter plates. Selected were antibodies reacted with the CGL3 membrane extract. Selected hybridomas were cloned twice by limiting dilution.

The mabs prepared by the just described method were characterized by Western blots of the GEX-3X-MN fusion protein, and with membrane extracts from the CGL1 and CGL3 cells. Representative of the mabs prepared are Mabs MN9, MN12 and MN7.

Mab MN9. Monoclonal antibody MN9 (Mab MN9) reacts to the same epitope as Mab M75, represented by the sequence from AA 62 to AA 67 [SEQ. ID. NO.: 10] of the Figure 1 MN protein. As Mab M75, Mab MN9 recognizes both the GEX-3X-MN fusion protein and native MN protein equally well.

Mabs corresponding to Mab MN9 can be prepared reproducibly by screening a series of mabs prepared against an MN protein/polypeptide, such as, the GEX-3X-MN fusion protein, against the peptide representing the epitope for Mabs M75 and MN9, that is, SEQ. ID. NO.: 10. Alternatively, the Novatope system [Novagen] or competition with the deposited Mab M75 could be used to select mabs comparable to Mabs M75 and MN9.

Mab MN12. Monoclonal antibody MN12 (Mab MN12) is produced by the mouse lymphocytic hybridoma MN 12.2.2 which was deposited under ATCC Designation HB 11647 on June 9, 1994 at the American Type Culture Collection (ATCC) at 12301 Parklawn Drive, Rockville, MD 20852 (USA). Antibodies corresponding to Mab MN12 can also be made, analogously to the method outlined above for Mab MN9, by screening a series of antibodies prepared against an MN protein/polypeptide, against the peptide representing the epitope for Mab MN12. That peptide is AA 55 - AA 60 of Figure 1 [SEQ. ID. NO.: 11]. The Novatope system could also be used to find antibodies specific for said epitope.

Mab MN7. Monoclonal antibody MN7 (Mab MN7) was selected from mabs prepared against nonglycosylated GEX-3X-MN as described above. It recognizes the epitope on MN represented by the amino acid sequence from AA 127 to AA 147 [SEQ. ID. NO.: 12] of the Figure 1 MN protein. Analogously to methods described above for Mabs MN9 and MN12, mabs

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corresponding to Mab MN7 can be prepared by selecting mabs prepared against an MN protein/polypeptide that are reactive with the peptide having SEQ. ID. NO.: 12, or by the stated alternative means.

5 Epitope Mapping

Epitope mapping was performed by the Novatope system, a kit for which is commercially available from Novagen, Inc. [See, for analogous example, Li et al., Nature, 363: 85-88 (6 May 1993).] In brief, the MN cDNA was cut into
10 overlapping short fragments of approximately 60 base pairs. The fragments were expressed in E. coli, and the E. coli colonies were transferred onto nitrocellulose paper, lysed and probed with the mab of interest. The MN cDNA of clones reactive with the mab of interest was sequenced, and the
15 epitopes of the mabs were deduced from the overlapping polypeptides found to be reactive with each mab.

Therapeutic Use of MN-Specific Antibodies

The MN-specific antibodies of this invention, monoclonal and/or polyclonal, preferably monoclonal, and as
20 outlined above, may be used therapeutically in the treatment of neoplastic and/or pre-neoplastic disease, either alone or in combination with chemotherapeutic drugs or toxic agents, such as ricin A. Further preferred for therapeutic use would be biologically active antibody fragments as described herein.
25 Also preferred MN-specific antibodies for such therapeutic uses would be humanized monoclonal antibodies.

The MN-specific antibodies can be administered in a therapeutically effective amount, preferably dispersed in a physiologically acceptable, nontoxic liquid vehicle.

30 Imaging Use of Antibodies

Further, the MN-specific antibodies of this invention when linked to an imaging agent, such as a radionuclide, can be used for imaging. Biologically active antibody fragments or humanized monoclonal antibodies, may be
35 preferred for imaging use.

A patient's neoplastic tissue can be identified as, for example, sites of transformed stem cells, of tumors and locations of any metastases. Antibodies, appropriately labeled or linked to an imaging agent, can be injected in a physiologically acceptable carrier into a patient, and the binding of the antibodies can be detected by a method appropriate to the label or imaging agent, for example, by scintigraphy.

Antisense MN Nucleic Acid Sequences

MN genes are herein considered putative oncogenes and the encoded proteins thereby are considered to be putative oncoproteins. Antisense nucleic acid sequences substantially complementary to mRNA transcribed from MN genes, as represented by the antisense oligodeoxynucleotides ODN1 and ODN2 [SEQ. ID. NOS.: 3 and 4] can be used to reduce or prevent expression of the MN gene. [Zamecnick, P.C., "Introduction: Oligonucleotide Base Hybridization as a Modulator of Genetic Message Readout," pp. 1-6, Prospects for Antisense Nucleic Acid Therapy of Cancer and AIDS, (Wiley-Liss, Inc., New York, NY, USA; 1991); Wickstrom, E., "Antisense DNA Treatment of HL-60 Promyelocytic Leukemia Cells: Terminal Differentiation and Dependence on Target Sequence," pp. 7-24, id.; Leserman et al., "Targeting and Intracellular Delivery of Antisense Oligonucleotides Interfering with Oncogene Expression," pp. 25-34, id.; Yokoyama, K., "Transcriptional Regulation of c-myc Proto-oncogene by Antisense RNA," pp. 35-52, id.; van den Berg et al., "Antisense fos Oligodeoxyribonucleotides Suppress the Generation of Chromosomal Aberrations," pp. 63-70, id.; Mercola, D., "Antisense fos and fun RNA," pp. 83-114, id.; Inouye, Gene, 72: 25-34 (1988); Miller and Ts'o, Ann. Reports Med. Chem., 23: 295-304 (1988); Stein and Cohen, Cancer Res., 48: 2659-2668 (1988); Stevenson and Inversen, J. Gen. Virol., 70: 2673-2682 (1989); Goodchild, "Inhibition of Gene Expression by Oligonucleotides," pp. 53-77, Oligodeoxynucleotides: Antisense Inhibitors of Gene Expression (Cohen, J.S., ed; CRC Press, Boca Raton, Florida,

USA; 1989); Dervan et al., "Oligonucleotide Recognition of Double-helical DNA by Triple-helix Formation," pp. 197-210, id.; Neckers, L.M., "Antisense Oligodeoxynucleotides as a Tool for Studying Cell Regulation: Mechanisms of Uptake and Application to the Study of Oncogene Function," pp. 211-232, id.; Leitner et al., PNAS (USA), 87: 3430-3434 (1990); Bevilacqua et al., PNAS (USA), 85: 831-835 (1988); Loke et al. Curr. Top. Microbiol. Immunol., 141: 282-288 (1988); Sarin et al., PNAS (USA), 85: 7448-7451 (1988); Agrawal et al., "Antisense Oligonucleotides: A Possible Approach for Chemotherapy and AIDS," International Union of Biochemistry Conference on Nucleic Acid Therapeutics (Jan. 13-17, 1991; Clearwater Beach, Florida, USA); Armstrong, L., Ber. Week, pp. 88-89 (March 5, 1990); and Weintraub et al., Trends, 1: 22-25 (1985).] Such antisense nucleic acid sequences, preferably oligonucleotides, by hybridizing to the MN mRNA, particularly in the vicinity of the ribosome binding site and translation initiation point, inhibits translation of the mRNA. Thus, the use of such antisense nucleic acid sequences may be considered to be a form of cancer therapy.

Preferred antisense oligonucleotides according to this invention are gene-specific ODNs or oligonucleotides complementary to the 5' end of MN mRNA. Particularly preferred are the 29-mer ODN1 and 19-mer ODN2 [SEQ. ID. NOS.: 3 and 4]. Those antisense ODNs are representative of the many antisense nucleic acid sequences that can function to inhibit MN gene expression. Ones of ordinary skill in the art could determine appropriate antisense nucleic acid sequences, preferably antisense oligonucleotides, from the nucleic acid sequences of Figures 1 and 3a-d.

Also, as described above, CGL3 cells transfected with an "antisense" MN cDNA/promoter construct formed colonies much smaller than control CGL3 cells.

Vaccines

It will be readily appreciated that MN proteins and polypeptides of this invention can be incorporated into vaccines capable of inducing protective immunity against

neoplastic disease and a dampening effect upon tumorigenic activity. Efficacy of a representative MN fusion protein GEX-3X-MN as a vaccine in a rat model is shown in Example 2.

MN proteins and/or polypeptides may be synthesized
5 or prepared recombinantly or otherwise biologically, to
comprise one or more amino acid sequences corresponding to one
or more epitopes of the MN proteins either in monomeric or
multimeric form. Those proteins and/or polypeptides may then
be incorporated into vaccines capable of inducing protective
10 immunity. Techniques for enhancing the antigenicity of such
polypeptides include incorporation into a multimeric
structure, binding to a highly immunogenic protein carrier,
for example, keyhole limpet hemocyanin (KLH), or diphtheria
toxoid, and administration in combination with adjuvants or
15 any other enhancers of immune response.

Preferred MN proteins/polypeptides to be used in a
vaccine according to this invention would be genetically
engineered MN proteins. Preferred recombinant MN protein are
the GEX-3X-MN, MN 20-19, MN-Fc and MN-PA proteins.

20 Other exemplary vaccines include vaccinia-MN (live
vaccinia virus with full-length MN cDNA), and baculovirus-MN
(full length MN cDNA inserted into baculovirus vector, e.g. in
suspension of infected insect cells). Different vaccines may
be combined and vaccination periods can be prolonged.

25 A preferred exemplary use of such a vaccine of this
invention would be its administration to patients whose MN-
carrying primary cancer had been surgically removed. The
vaccine may induce active immunity in the patients and prevent
recidivism or metastasis.

30 It will further be appreciated that anti-idiotypic
antibodies to antibodies to MN proteins/polypeptides are also
useful as vaccines and can be similarly formulated.

An amino acid sequence corresponding to an epitope
of an MN protein/polypeptide either in monomeric or multimeric
35 form may also be obtained by chemical synthetic means or by
purification from biological sources including genetically
modified microorganisms or their culture media. [See Lerner,
"Synthetic Vaccines", Sci. Am. 248(2): 66-74 (1983).] The

protein/polypeptide may be combined in an amino acid sequence with other proteins/polypeptides including fragments of other proteins, as for example, when synthesized as a fusion protein, or linked to other antigenic or non-antigenic polypeptides of synthetic or biological origin. In some instances, it may be desirable to fuse a MN protein or polypeptide to an immunogenic and/or antigenic protein or polypeptide, for example, to stimulate efficacy of a MN-based vaccine.

The term "corresponding to an epitope of an MN protein/polypeptide" will be understood to include the practical possibility that, in some instances, amino acid sequence variations of a naturally occurring protein or polypeptide may be antigenic and confer protective immunity against neoplastic disease and/or anti-tumorigenic effects. Possible sequence variations include, without limitation, amino acid substitutions, extensions, deletions, truncations, interpolations and combinations thereof. Such variations fall within the contemplated scope of the invention provided the protein or polypeptide containing them is immunogenic and antibodies elicited by such a polypeptide or protein cross-react with naturally occurring MN proteins and polypeptides to a sufficient extent to provide protective immunity and/or anti-tumorigenic activity when administered as a vaccine.

Such vaccine compositions will be combined with a physiologically acceptable medium, including immunologically acceptable diluents and carriers as well as commonly employed adjuvants such as Freund's Complete Adjuvant, saponin, alum, and the like. Administration would be in immunologically effective amounts of the MN proteins or polypeptides, preferably in quantities providing unit doses of from 0.01 to 10.0 micrograms of immunologically active MN protein and/or polypeptide per kilogram of the recipient's body weight. Total protective doses may range from 0.1 to about 100 micrograms of antigen. Routes of administration, antigen dose, number and frequency of injections are all matters of optimization within the scope of the ordinary skill in the art.

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The following examples are for purposes of illustration only and not meant to limit the invention in any way.

Example 1

5 Immunohistochemical Staining of Tissue Specimens

To study and evaluate the tissue distribution range and expression of MN proteins, the monoclonal antibody M75 was used to stain immunohistochemically a variety of human tissue specimens. The primary antibody used in these
10 immunohistochemical staining experiments was the M75 monoclonal antibody. A biotinylated second antibody and streptavidin-peroxidase were used to detect the M75 reactivity in sections of formalin-fixed, paraffin-embedded tissue samples. A commercially available amplification kit,
15 specifically the DAKO LSAB™ kit [DAKO Corp., Carpinteria, CA (USA)] which provides matched, ready made blocking reagent, secondary antibody and streptavidin-horseradish peroxidase was used in these experiments.

M75 immunoreactivity was tested according to the
20 methods of this invention in multiple-tissue sections of breast, colon, cervical, lung and normal tissues. Such multiple-tissue sections were cut from paraffin blocks of tissues called "sausages" that were purchased from the City of Hope [Duarte, CA (USA)]. Combined in such a multiple-tissue
25 section were normal, benign and malignant specimens of a given tissue; for example, about a score of tissue samples of breast cancers from different patients, a similar number of benign breast tissue samples, and normal breast tissue samples would be combined in one such multiple-breast-tissue section. The
30 normal multiple-tissue sections contained only normal tissues from various organs, for example, liver, spleen, lung, kidney, adrenal gland, brain, prostate, pancreas, thyroid, ovary, and testis.

Also screened for MN gene expression were multiple
35 individual specimens from cervical cancers, bladder cancers, renal cell cancers, and head and neck cancers. Such specimens were obtained from U.C. Davis Medical Center in Sacramento, CA

and from Dr. Shu Y. Liao [Department of Pathology; St. Joseph Hospital; Orange, CA (USA)].

Controls used in these experiments were the cell lines CGL3 (H/F-T-hybrid cells) and CGL1 (H/F-N hybrid cells) which are known to stain respectively, positively and negatively with the M75 monoclonal antibody. The M75 monoclonal antibody was diluted to a 1:5000 dilution wherein the diluent was either PBS [0.05 M phosphate buffered saline (0.15 M NaCl), pH 7.2-7.4] or PBS containing 1% protease-free BSA as a protein stabilizer.

Immunohistochemical Staining Protocol

The immunohistochemical staining protocol was followed according to the manufacturer's instructions for the DAKO LSAB™ kit. In brief, the sections were dewaxed, rehydrated and blocked to remove non-specific reactivity as well as endogenous peroxidase activity. Each section was then incubated with dilutions of the M75 monoclonal antibody. After the unbound M75 was removed by rinsing the section, the section was sequentially reacted with a biotinylated antimouse IgG antibody and streptavidin conjugated to horseradish peroxidase; a rinsing step was included between those two reactions and after the second reaction. Following the last rinse, the antibody-enzyme complexes were detected by reaction with an insoluble chromogen (diaminobenzidine) and hydrogen peroxide. A positive result was indicated by the formation of an insoluble reddish-brown precipitate at the site of the primary antibody reaction. The sections were then rinsed, counterstained with hematoxylin, dehydrated and cover slipped. Then the sections were examined using standard light microscopy.

Interpretation. A deposit of a reddish brown precipitate over the plasma membrane was taken as evidence that the M75 antibody had bound to a MN antigen in the tissue. The known positive control (CGL3) had to be stained to validate the assay. Section thickness was taken into consideration to compare staining intensities, as thicker

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sections produce greater staining intensity independently of other assay parameters.

Results

Preliminary examination of cervical specimens showed
5 that 62 of 68 squamous cell carcinoma specimens (91.2%)
stained positively with M75. Additionally, 2 of 6
adenocarcinomas and 2 of 2 adenosquamous cancers of the cervix
also stained positively. In early studies, 55.6% (10 of 18)
of cervical dysplasias stained positively. A total of 9
10 specimens including both cervical dysplasias and tumors,
exhibited some MN expression in normal appearing areas of the
endocervical glandular epithelium, usually at the basal layer.
In some specimens, whereas morphologically normal-looking
areas showed expression of MN antigen, areas exhibiting
15 dysplasia and/or malignancy did not show MN expression.

M75 positive immunoreactivity was most often
localized to the plasma membrane of cells, with the most
apparent stain being present at the junctions between adjacent
cells. Cytoplasmic staining was also evident in some cells;
20 however, plasma membrane staining was most often used as the
main criterion of positivity.

M75 positive cells tended to be near areas showing
keratin differentiation in cervical specimens. In some
specimens, positive staining cells were located in the center
25 of nests of non-staining cells. Often, there was very little,
if any, obvious morphological difference between staining
cells and non-staining cells. In some specimens, the positive
staining cells were associated with adjacent areas of
necrosis.

30 In most of the squamous cell carcinomas of the
cervix, the M75 immunoreactivity was focal in distribution,
i.e., only certain areas of the specimen stained. Although
the distribution of positive reactivity within a given
specimen was rather sporadic, the intensity of the reactivity
35 was usually very strong. In most of the adenocarcinomas of
the cervix, the staining pattern was more homogeneous, with
the majority of the specimen staining positively.

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Among the normal tissue samples, intense, positive and specific M75 immunoreactivity was observed only in normal stomach tissues, with diminishing reactivity in the small intestine, appendix and colon. No other normal tissue stained extensively positively for M75. Occasionally, however, foci of intensely staining cells were observed in normal intestine samples (usually at the base of the crypts) or were sometimes seen in morphologically normal appearing areas of the epithelium of cervical specimens exhibiting dysplasia and/or malignancy. In such, normal appearing areas of cervical specimens, positive staining was seen in focal areas of the basal layer of the ectocervical epithelium or in the basal layer of endocervical glandular epithelium. In one normal specimen of human skin, cytoplasmic MN staining was observed in the basal layer. The basal layers of these epithelia are usually areas of proliferation, suggesting the MN expression may be involved in cellular growth. In a few cervical biopsied specimens, MN positivity was observed in the morphologically normal appearing stratified squamous epithelium, sometimes associated with cells undergoing koilocytic changes.

Some colon adenomas (4 of 11) and adenocarcinomas (9 of 15) were positively stained. One normal colon specimen was positive at the base of the crypts. Of 15 colon cancer specimens, 4 adenocarcinomas and 5 metastatic lesions were MN positive. Fewer malignant breast cancers (3 of 25) and ovarian cancer specimens (3 of 15) were positively stained. Of 4 head and neck cancers, 3 stained very intensely with M75.

Although normal stomach tissue was routinely positive, 4 adenocarcinomas of the stomach were MN negative. Of 3 bladder cancer specimens (1 adenocarcinoma, 1 non-papillary transitional cell carcinoma, and 1 squamous cell carcinoma), only the squamous cell carcinoma was MN positive. Approximately 40% (12 of 30) of lung cancer specimens were positive; 2 of 4 undifferentiated carcinomas; 3 of 8 adenocarcinomas; 2 of 8 oat cell carcinomas; and, 5 of 10 squamous cell carcinomas. One hundred percent (4 of 4) of the renal cell carcinomas were MN positive.

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In summary, MN antigen, as detected by M75 and immunohistochemistry in the experiments described above, was shown to be prevalent in tumor cells, most notably in tissues of cervical cancers. MN antigen was also found in some cells of normal tissues, and sometimes in morphologically normal appearing areas of specimens exhibiting dysplasia and/or malignancy. However, MN is not usually extensively expressed in most normal tissues, except for stomach tissues where it is extensively expressed and in the tissues of the lower gastrointestinal tract where it is less extensively expressed. MN expression is most often localized to the cellular plasma membrane of tumor cells and may play a role in intercellular communication or cell adhesion. Representative results of experiments performed as described above are tabulated in Table 2.

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TABLE 2
Immunoreactivity of M75 in Various Tissues

	TISSUE	TYPE	POS/NEG (#pos/#tested)
5	liver, spleen, lung, kidney, adrenal gland, brain, prostate, pancreas, thyroid, ovary, testis	normal	NEG (all)
10	skin	normal	POS (in basal layer) (1/1)
	stomach	normal	POS
	small intestine	normal	POS
	colon	normal	POS
	breast	normal	NEG (0/10)
15	cervix	normal	NEG (0/2)
	breast	benign	NEG (0/17)
	colon	benign	POS (4/11)
	cervix	benign	POS (10/18)
	breast	malignant	POS (3/25)
20	colon	malignant	POS (9/15)
	ovarian	malignant	POS (3/15)
	lung	malignant	POS (12/30)
	bladder	malignant	POS (1/3)
	head & neck	malignant	POS (3/4)
25	kidney	malignant	POS (4/4)
	stomach	malignant	NEG (0/4)
	cervix	malignant	POS (62/68)

The results recorded in this example indicate that the presence of MN proteins in a tissue sample from a patient may, in general, depending upon the tissue involved, be a marker signaling that a pre-neoplastic or neoplastic process is occurring. Thus, one may conclude from these results that diagnostic/prognostic methods that detect MN antigen may be particularly useful for screening patient samples for a number of cancers which can thereby be detected at a pre-neoplastic stage or at an early stage prior to obvious morphologic

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changes associated with dysplasia and/or malignancy being evident or being evident on a widespread basis.

Example 2

Vaccine -- Rat Model

5 As shown above in Example 7 of WO 93/18152 (International Publication Date: 16 September 1993), in some rat tumors, for example, the XC tumor cell line (cells from a rat rhabdomyosarcoma), a rat MN protein, related to human MN, is expressed. Thus a model was afforded to study antitumor
10 immunity induced by experimental MN-based vaccines. The following representative experiments were performed.

Nine- to eleven-day-old Wistar rats from several families were randomized, injected intraperitoneally with 0.1 ml of either control rat sera (the C group) or with rat serum
15 against the MN fusion protein GEX-3X-MN (the IM group). Simultaneously both groups were injected subcutaneously with 10^6 XC tumor cells.

Four weeks later, the rats were sacrificed, and their tumors weighed. The results are shown in Figure 2.
20 Each point on the graph represents a tumor from one rat. The difference between the two groups -- C and IM -- was significant by Mann-Whitney rank test ($U = 84$, $\alpha < 0.025$). The results indicate that the IM group of baby rats developed tumors about one-half the size of the controls, and 5 of the
25 18 passively immunized rats developed no tumor at all, compared to 1 of 18 controls.

Example 3

Expression of Full-Length MN cDNA in NIH 3T3 Cells

The role of MN in the regulation of cell
30 proliferation was studied by expressing the full-length cDNA in NIH 3T3 cells. That cell line was chosen since it had been used successfully to demonstrate the phenotypic effect of a number of proto-oncogenes [Weinberg, R.A., Cancer Res., 49: 3713 (1989); Hunter, T., Cell, 64: 249 (1991)]. Also, NIH
35 3T3 cells express no endogenous MN-related protein that is detectable by Mab M75.

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The full length MN cDNA was obtained by ligation of the two cDNA clones using the unique BamHI site and subcloned from pBluescript into KpnI-SacI sites of the expression vector pSG5C. pSG5C was kindly provided by Dr. Richard Kettman [Department of Molecular Biology, Faculty of Agricultural Sciences, B-5030 Gembloux, Belgium]. pSG5C was derived from pSG5 [Stratagene] by inserting a polylinker consisting of a sequence having several neighboring sites for the following restriction enzymes: EcoRI, XhoI, KpnI, BamHI, SacI, 3 times TAG stop codon and BglII.

The recombinant pSG5C-MN plasmid was co-transfected in a 10:1 ratio (10 μ g : 1 μ g) with the pSV2neo plasmid [Southern and Berg, J. Mol. Appl. Genet., 1: 327 (1982)] which contains the neo gene as a selection marker. The co-transfection was carried out by calcium phosphate precipitation method [Mammalian Transfection Kit; Stratagene] into NIH 3T3 cells plated a day before at a density of 1×10^5 per 60 mm dish. As a control, pSV2neo was co-transfected with empty pSG5C.

Transfected cells were cultured in DMEM medium supplemented with 10% FCS and 600 μ g ml⁻¹ of G418 [Gibco BRL] for 14 days. The G418-resistant cells were clonally selected, expanded and analysed for expression of the transfected cDNA by Western blotting using iodinated Mab M75.

For an estimation of cell proliferation, the clonal cell lines were plated in triplicates (2×10^4 cells/well) in 24-well plates and cultivated in DMEM with 10% FCS and 1% FCS, respectively. The medium was changed each day, and the cell number was counted using a hemacytometer.

To determine the DNA synthesis, the cells were plated in triplicate in 96-well plate at a density of 10^4 /well in DMEM with 10% FCS and allowed to attach overnight. Then the cells were labeled with ³H-thymidine for 24 hours, and the incorporated radioactivity was counted.

For the anchorage-independent growth assay, cells (2×10^4) were suspended in a 0.3% agar in DMEM containing 10% FCS and overlaid onto 0.5% agar medium in 60 mm dish. Colonies grown in soft agar were counted two weeks after plating.

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Several clonal cell lines constitutively expressing both 54 and 58 kd forms of MN protein in levels comparable to those found in LCMV-infected HeLa cells were obtained.

Selected MN-positive clones and negative control cells (mock-transfected with an empty pSG5C plasmid) were subjected to further analyses directed to the characterization of their phenotype and growth behavior.

The MN-expressing NIH 3T3 cells displayed spindle-shaped morphology, and increased refractility; they were less adherent to the solid support and smaller in size. The control (mock transfected cells) had a flat morphology, similar to parental NIH 3T3 cells. In contrast to the control cells that were aligned and formed a monolayer with an ordered pattern, the cells expressing MN lost the capacity for growth arrest and grew chaotically on top of one another. Correspondingly, the MN-expressing cells were able to reach significantly higher (more than 2x) saturation densities (Table 3) and were less dependent on growth factors than the control cells.

MN transfectants also showed faster doubling times (by 15%) and enhanced DNA synthesis (by 10%), as determined by the amount of [³H]-thymidine incorporated in comparison to control cells. Finally, NIH 3T3 cells expressing MN protein grew in soft agar. The diameter of colonies grown for 14 days ranged from 0.1 to 0.5 mm; however, the cloning efficiency of MN transfectants was rather low (2.4%). Although that parameter of NIH 3T3 cells seems to be less affected by MN than by conventional oncogenes, all other data are consistent with the idea that MN plays a role in cell growth control.

Table 3

Growth Properties of NIH 3T3 Cells Expressing MN Protein

Transfected DNA	pSG5C/ pSV2neo	pSG5C-MN/ pSV2neo
Doubling time ^a (hours)	27.9 ± 0.5	24.1 ± 1.3

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Saturation density ^b (cells x 10 ⁴ /cm ²)	4.9 ± 0.2	11.4 ± 0.4
Cloning efficiency (%) ^c	< 0.01	2.4 ± 0.2

5

^aFor calculation of the doubling time, the proliferation rate of exponentially growing cells was used. ^bThe saturation cell density was derived from the cell number 4 days after reaching confluency. ^cColonies greater than 0.1 mm in diameter were scored at day 14. Cloning efficiency was estimated as a percentage of colonies per number of cells plated, with correction for cell viability.

Example 4

Acceleration of G1 Transit and Decrease in Mitomycin C Sensitivity Caused by MN Protein

15

For the experiments described in this example, the stable MN transfectants of NIH 3T3 cells generated as described in Example 3 were used. Four selected MN-positive clones and four control mock-transfected clones were either used individually or in pools.

Flow cytometric analyses of asynchronous cell populations. Cells that had been grown in dense culture were plated at 1 x 10⁶ cells per 60 mm dish. Four days later, the cells were collected by trypsinization, washed, resuspended in PBS, fixed by dropwise addition of 70% ethanol and stained by propidium iodine solution containing RNase. Analysis was performed by FACStar using DNA cell cycle analysis software [Becton Dickinson; Franklin Lakes, NJ (USA)].

Exponentially growing cells were plated at 5 x 10⁵ cells per 60 mm dish and analysed as above 2 days later. Forward light scatter was used for the analysis of relative cell sizes. The data were evaluated using Kolmogorov-Smirnov test [Young, J. Histochem. Cytochem., 25: 935 (1977)].

The flow cytometric analyses revealed that clonal populations constitutively expressing MN protein showed a

35

decreased percentage of cells in G1 phase and an increased percentage of cells in G2-M phases. Those differences were more striking in cell populations grown throughout three passages in high density cultures than in exponentially growing subconfluent cells. That observation supports the idea that MN protein has the capacity to perturb contact inhibition.

Also observed was a decrease in the size of MN expressing cells seen in both exponentially proliferating and high density cultures. It is possible that the MN-mediated acceleration of G1 transit is related to the above-noted shorter doubling time (by about 15%) of exponentially proliferating MN-expressing NIH 3T3 cells. Also, MN expressing cells displayed substantially higher saturation density and lower serum requirements than the control cells. Those facts suggest that MN-transfected cells had the capacity to continue to proliferate despite space limitations and diminished levels of serum growth factors, whereas the control cells were arrested in G1 phase.

Limiting conditions. The proliferation of MN-expressing and control cells was studied both in optimal and limiting conditions. Cells were plated at 2×10^4 per well of 24-well plate in DMEM with 10% FCS. The medium was changed at daily intervals until day 4 when confluency was reached, and the medium was no longer renewed. Viable cells were counted in a hemacytometer at appropriate times using trypan blue dye exclusion. The numbers of cells were plotted versus time wherein each plot point represents a mean value of triplicate determination.

The results showed that the proliferation of MN expressing and control cells was similar during the first phase when the medium was renewed daily, but that a big difference in the number of viable cells occurred after the medium was not renewed. More than half of the control cells were not able to withstand the unfavorable growth conditions. In contrast, the MN-expressing cells continued to proliferate even when exposed to increasing competition for nutrients and serum growth factors.

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Those results were supported also by flow cytometric analysis of serum starved cells grown for two days in medium containing 1% FCS. While 83% of control cells accumulated in G0-G1 phase (S = 5%, G2-M = 12%), expression of MN protein partially reversed the delay in G1 as indicated by cell cycle distribution of MN transfectants (G0-G1 = 65%, S = 10%, G2-M = 26%). The results of the above-described experiments suggest that MN protein might function to release the G1/S checkpoint and allow cells to proliferate under unfavorable conditions.

MMC. To test that assumption, unfavorable conditions were simulated by treating cells with the DNA damaging drug mitomycin C (MMC) and then following their proliferation and viability. The mechanism of action of MMC is thought to result from its intracellular activation and subsequent DNA alkylation and crosslinking [Yier and Szybalski, Science, 145: 55 (1964)]. Normally, cells respond to DNA damage by arrest of their cell cycle progression to repair defects and prevent acquisition of genomic instability. Large damage is accompanied by marked cytotoxicity. However, many studies [for example, Peters et al., Int. J. Cancer, 54: 450 (1993)] concern the emergence of drug resistant cells both in tumor cell populations and after the introduction of oncogenes into nontransformed cell lines.

The response of MN-transfected NIH 3T3 cells to increasing concentrations of MMC was determined by continuous [³H]-thymidine labeling. Cells were plated in 96-well microtiter plate concentration of 10⁴ per well and incubated overnight in DMEM with 10% FCS to attach. Then the growth medium was replaced with 100 µl of medium containing increasing concentrations of MMC from 1 µl/ml to 32 µg/ml. All the drug concentrations were tested in three replicate wells. After 5 hours of treatment, the MMC was removed, cells were washed with PBS and fresh growth medium without the drug was added. After overnight recovery, the fractions of cells that were actively participating in proliferation was determined by continuous 24-hr labeling with [³H]-thymidine. The incorporation by the treated cells was compared to that of the control, untreated cells, and the proliferating fractions

were considered as a percentage of the control's incorporation.

The viability of the treated cells was estimated three days later by a CellTiter 96 AQ Non-Radioactive Cell Proliferation Assay [Promega] which is based on the bioreduction of methotrexate (MTX) into a water soluble formazan that absorbs light at 490 nm. The percentage of surviving cells was derived from the values of absorbance obtained after subtraction of background.

The control and MN-expressing NIH 3T3 cells showed remarkable differences in their responses to MMC. The sensitivity of the MN-transfected cells appeared considerably lower than the control's in both sections of the above-described experiments. The results suggested that the MN-transfected cells were able to override the negative growth signal mediated by MMC.

ATCC Deposits. The material listed below was deposited with the American Type Culture Collection (ATCC) at 12301 Parklawn Drive, Rockville, MD 20852 (USA). The deposits were made under the provisions of the Budapest Treaty on the International Recognition of Deposited Microorganisms for the Purposes of Patent Procedure and Regulations thereunder (Budapest Treaty). Maintenance of a viable culture is assured for thirty years from the date of deposit. The hybridomas and plasmids will be made available by the ATCC under the terms of the Budapest Treaty, and subject to an agreement between the Applicants and the ATCC which assures unrestricted availability of the deposited hybridomas and plasmids to the public upon the granting of patent from the instant application. Availability of the deposited strain is not to be construed as a license to practice the invention in contravention of the rights granted under the authority of any Government in accordance with its patent laws.

	<u>Hybridoma</u>	<u>Deposit Date</u>	<u>ATCC #</u>
35	VU-M75	September 17, 1992	HB 11128
	MN 12.2.2	June 9, 1994	HB 11647

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<u>Plasmid</u>	<u>Deposit Date</u>	<u>ATCC #</u>
A4a	June 6, 1995	97199
XE1	June 6, 1995	97200
XE3	June 6, 1995	97198

5 The description of the foregoing embodiments of the
invention have been presented for purposes of illustration and
description. They are not intended to be exhaustive or to
limit the invention to the precise form disclosed, and
obviously many modifications and variations are possible in
10 light of the above teachings. The embodiments were chosen and
described in order to explain the principles of the invention
and its practical application to enable thereby others skilled
in the art to utilize the invention in various embodiments and
with various modifications as are suited to the particular use
15 contemplated.

 All references cited herein are hereby incorporated
by reference.

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CLAIMS

1. An isolated nucleic acid containing at least twenty-seven nucleotides wherein the nucleotide sequence for said nucleic acid is selected from the group consisting of:

- 5 (a) SEQ. ID. NOS.: 1, 5, and 27-49 and nucleotide sequences complementary to SEQ. ID. NOS.: 1, 5 and 27-49;
- (b) nucleotide sequences that hybridize under stringent hybridization conditions to one or more of the following nucleotide sequences: SEQ. ID. NOS.: 1, 5, and 27-
10 49 and the respective complements of SEQ. ID. NOS.: 1, 5 and 27-49; and
- (c) nucleotide sequences that differ from the nucleotide sequences of (a) and (b) in codon sequence due to the degeneracy of the genetic code.

15 2. An isolated nucleic acid according to Claim 1 wherein said nucleotide sequence is selected from the group consisting of:

- (a) SEQ. ID. NO.: 5 and its complement;
- (b) nucleotide sequences that hybridize under
20 stringent conditions to SEQ. ID. NO.: 5 or to its complement;
- (c) nucleotide sequences that differ from the nucleotide sequences of (a) or (b) in codon sequence due to the degeneracy of the genetic code.

25 3. An isolated nucleic acid containing at least sixteen nucleotides wherein the nucleotide sequence therefor is selected from the group consisting of:

- (a) the MN nucleotide sequences contained in plasmids A4a, XE1 and XE3 which were deposited at the American Type Culture Collection (ATCC) in Rockville, Maryland in the
30 United State of America under the respective ATCC Nos. 97199, 97200, and 97198;

(b) nucleotide sequences that hybridize under stringent conditions to the MN nucleotide sequences of (a);
and

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(c) nucleotide sequences that differ from the nucleotide sequences of (a) or (b) in codon sequence due to the degeneracy of the genetic code.

4. An isolated nucleic acid according to Claim 3
5 which functions as a polymerase chain reaction primer for MN nucleic acid sequences, and which is from 16 to about 50 nucleotides in length.

5. An isolated nucleic acid, containing at least fifty nucleotides, encoding an MN protein or polypeptide that
10 is specifically bound either by monoclonal antibodies designated M75 secreted by the hybridoma VU-M75 deposited at the American Type Culture Collection (ATCC) in Rockville, Maryland in the United States of America under ATCC No. HB 11128, or by monoclonal antibodies designated MN12 secreted by
15 the hybridoma MN 12.2.2 deposited at the ATCC under ATCC No. 11647, or by both of said monoclonal antibodies.

6. An isolated nucleic acid which is operatively linked to an expression control sequence within a vector wherein said nucleic acid is selected from the group
20 consisting of:

- (a) SEQ. ID. NO.: 1 and its complement;
- (b) nucleic acids that hybridize under stringent hybridization conditions to SEQ. ID. NO.: 1 or to its complement; and
- 25 (c) nucleic acids that differ from the nucleic acid sequences of (a) and (b) due to the degeneracy of the genetic code.

7. A unicellular host, which is either prokaryotic or eukaryotic, that is transformed or transfected with the
30 isolated nucleic acid operatively linked to an expression control sequence in a vector according to Claim 6.

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8. A method of recombinantly producing an MN protein, MN fusion protein or MN polypeptide comprising the steps of:

- 5 (a) transforming a unicellular host with the isolated nucleic acid operatively linked to an expression control sequence in a vector according to Claim 6;
- (b) culturing said unicellular host so that said MN protein or polypeptide is expressed; and
- 10 (c) extracting and isolating said MN protein or polypeptide.

9. A recombinant nucleic acid encoding a fusion protein, that consists essentially of an MN protein or polypeptide and a non-MN protein or polypeptide, wherein the nucleotide sequence for the portion of the nucleic acid
15 encoding the MN protein or polypeptide is selected from the group consisting of:

- (a) SEQ. ID. NO.: 1;
 - (b) nucleotide sequences that hybridize under stringent conditions to SEQ. ID. NO.: 1 or to its complement;
20 and
 - (c) degenerate variants of SEQ. ID. NO.: 1 and of the nucleotide sequences of (b);
- wherein the nucleic acid encoding said MN protein or polypeptide contains at least twenty-nine nucleotides.

25 10. A method of detecting mutations in an isolated MN gene and/or fragment(s) thereof comprising the steps of:
amplifying one or more fragment(s) of said gene by the polymerase chain reaction (PCR); and
determining whether said one or more fragments
30 contain any mutations.

11. An MN protein, MN fusion protein or MN polypeptide, wherein said MN protein, MN polypeptide or the MN protein portion of said MN fusion protein is encoded by a nucleic acid of at least twenty-nine nucleotides which is
35 selected from the group consisting of:

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(a) SEQ. ID. NO. NO.: 1;

(b) nucleotide sequences that hybridize under stringent hybridization conditions to SEQ. ID. NO.: 1 or to its complement; and

5 (c) nucleotide sequences that differ from SEQ. ID. NO.: 1 or from the sequences of (b) in codon sequence due to the degeneracy of the genetic code.

12. An MN protein, MN fusion protein, or MN polypeptide wherein said MN protein or polypeptide has, and
10 wherein said MN fusion protein contains, an amino acid sequence selected from the group consisting of SEQ. ID. NOS.: 2, 6, 10-16, and 50-53.

13. A vaccine comprising an immunogenic amount of one or more MN proteins, MN fusion proteins, and/or MN
15 polypeptides according to Claims 11 or 12 dispersed in a physiologically acceptable, nontoxic vehicle, which amount is effective to immunize a vertebrate against a neoplastic disease associated with expression of MN antigen.

14. An antibody which specifically binds to an MN
20 protein, an MN fusion protein and/or an MN polypeptide according to Claims 11 or 12.

15. A monoclonal antibody according to Claim 14 which is designated MN12 and is secreted by the hybridoma MN
12.2.2 which was deposited at the American Type Culture
25 Collection (ATCC) in Rockville, Maryland in the United States of America under ATCC No. HB 11647.

16. An antibody according to Claim 14 which specifically binds to an MN antigen epitope selected from the group of epitopes represented by the following amino acid
30 sequences: SEQ. ID. NOS. 10-16.

17. An antibody according to Claim 14 which is linked to an imaging agent, to a chemotherapeutic agent or to a toxic agent.

18. A method of imaging pre-neoplastic or
5 neoplastic disease in a patient comprising:

(a) injecting said patient with antibody linked to an imaging agent according to Claim 17; and

(b) detecting the binding of said antibody.

19. A hybridoma designated MN 12.2.2 which produces
10 the monoclonal antibody MN12, and which was deposited at the American Type Culture Collection (ATCC) in Rockville Maryland in the United States of America under ATCC Accession No. HB 11647.

20. A method of delivering a chemotherapeutic agent
15 or toxic agent to a cancer cell which comprises contacting said cell with an antibody linked to a chemotherapeutic agent or to a toxic agent according to Claim 17.

21. A method of treating neoplastic disease in a patient comprising administering to said patient a
20 therapeutically effective amount of a composition comprising antibodies which specifically bind to an MN protein, an MN fusion protein and/or an MN polypeptide according to Claims 11 or 12.

22. A method of detecting and/or quantitating in a
25 vertebrate sample MN antigen comprising the steps of:

(a) contacting said sample with one or more antibodies according to Claims 14, 15 or 16; and

(b) detecting and/or quantitating binding of said antibody in said sample.

23. A method according to Claim 22 wherein said
30 vertebrate sample is a human tissue specimen, such as, a cell

smear, a histological section from a biopsied tissue or organ, or an imprint preparation.

24. A method according to Claim 23 wherein said tissue specimen is ovarian, endometrial, or cervical.

5 25. A method of detecting and/or quantitating MN-specific antibodies in a vertebrate sample comprising the steps of:

10 (a) contacting and incubating the vertebrate sample with an MN protein, an MN fusion protein and/or an MN polypeptide according to Claims 11 or 12; and

(b) detecting and/or quantitating binding of said MN protein, MN fusion protein and/or MN polypeptide to antibody in said sample.

15 26. A method of treating neoplastic disease and/or pre-neoplastic disease comprising inhibiting the expression of MN genes by administering one or more antisense nucleic acid sequences that hybridize under stringent conditions to mRNA transcribed from MN genes.

20 27. Vectors containing an MN nucleic acid sequence wherein said MN nucleic acid sequence is selected from the group consisting of:

(a) SEQ. ID. NO.: 5 and its complement;

(b) nucleic acids that hybridize to SEQ. ID. NO.: 5 or to its complement; and

25 (c) nucleic acids that differ from the nucleic acids of (a) or (b) due to the degeneracy of the genetic code;

wherein said nucleic acid is at least twenty-nine nucleotides in length.

30 28. Vectors containing an MN nucleic acid sequence according to Claim 27 selected from the group consisting of the plasmids A4a, XE1 and XE3 which are deposited at the American Type Culture Collection (ATCC) in Rockville, Maryland

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in the United States of America under the respective ATCC accession numbers 97199, 97200 and 97198.

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1	1	ACA	GTC	AGC	CGC	ATG	GCT	CCC	CTG	TGC	CCC	AGC	CCC	TGG	CTC	CCT	CTG	12	48
13	49	L	I	P	A	P	A	P	G	L	T	V	Q	L	L	L	S	28	96
		TTG	ATC	CCG	GCC	CCT	GCT	CCA	GGC	CTC	ACT	GTG	CAA	CTG	CTG	TCA			
29	97	L	L	L	L	M	P	V	H	P	Q	R	L	P	R	M	44	144	
		CTG	CTG	CTT	CTG	ATG	CCT	GTC	CAT	CCC	CAG	AGG	TTG	CCC	CGG	ATG	CAG		
45	145	E	D	S	P	L	G	G	G	S	S	G	E	D	D	P	60	192	
		GAG	GAT	TCC	CCC	TTG	GGA	GGA	GGC	TCT	TCT	GGG	GAA	GAT	GAC	CCA	CTG		
61	193	G	E	E	D	L	P	S	E	E	D	S	P	R	E	E	76	240	
		GGC	GAG	GAG	GAT	CTG	CCC	AGT	GAA	GAG	GAT	TCA	CCC	AGA	GAG	GAG	GAT		
77	241	P	P	G	E	E	D	L	P	G	E	E	D	L	P	G	92	288	
		CCA	CCC	GGA	GAG	GAG	GAT	CTA	CCT	GGA	GAG	GAG	GAT	CTA	CCT	GGA	GAG		
93	289	E	D	L	P	E	V	K	P	K	S	E	E	E	G	S	108	336	
		GAG	GAT	CTA	CCT	GAA	GTT	AAG	CCT	AAA	TCA	GAA	GAA	GAG	GGC	TCC	CTG		
109	337	K	L	E	D	L	P	T	V	E	A	P	G	D	P	Q	124	384	
		AAG	TTA	GAG	GAT	CTA	CCT	ACT	GTT	GAG	GCT	CCT	GGA	GAT	CCT	CAA	GAA		
125	385	P	Q	N	N	A	H	R	D	K	E	G	D	D	Q	S	140	432	
		CCC	CAG	AAT	AAT	GCC	CAC	AGG	GAC	AAA	GAA	GGG	GAT	GAC	CAG	AGT	CAT		
141	433	W	R	Y	G	G	D	P	P	W	P	R	V	S	P	A	156	480	
		TGG	CGC	TAT	GGA	GGC	GAC	CCG	CCC	TGG	CCC	CGG	GTG	TCC	CCA	GCC	TGC		
157	481	A	G	R	F	Q	S	P	V	D	I	R	P	Q	L	A	172	528	
		GCG	GGC	CGC	TTC	CAG	TCC	CCG	GTG	GAT	ATC	CGC	CCC	CAG	CTC	GCC	GCC		

FIG.-1A

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173	F	C	P	A	L	R	P	L	E	L	L	G	F	Q	L	P	188
529	TTC	TGC	CCG	GCC	CTG	CGC	CCC	CTG	GAA	CTC	CTG	GGC	TTC	CAG	CTC	CCG	576
189	P	L	P	E	L	R	L	R	N	N	G	H	S	V	Q	L	204
577	CCG	CTC	CCA	GAA	CTG	CGC	CTG	CGC	AAC	AAT	GGC	CAC	AGT	GTG	CAA	CTG	624
205	T	L	P	P	G	L	E	M	A	L	G	P	G	R	E	Y	220
625	ACC	CTG	CCT	CCT	GGG	CTA	GAG	ATG	GCT	CTG	GGT	CCC	GGG	CGG	GAG	TAC	672
221	R	A	L	Q	L	H	L	H	W	G	A	A	G	R	P	G	236
673	CGG	GCT	CTG	CAG	CTG	CAT	CTG	CAC	TGG	GGG	GCT	GCA	GGT	CGT	CCG	GGC	720
237	S	E	H	T	V	E	G	H	R	F	P	A	E	I	H	V	252
721	TCG	GAG	CAC	ACT	GTG	GAA	GGC	CAC	CGT	TTC	CCT	GCC	GAG	ATC	CAC	GTG	768
253	V	H	L	S	T	A	F	A	R	V	D	E	A	L	G	R	268
769	GTT	CAC	CTC	AGC	ACC	GCC	TTT	GCC	AGA	GTT	GAC	GAG	GCC	TTG	GGG	CGC	816
269	P	G	G	L	A	V	L	A	A	F	L	E	E	G	P	E	284
817	CCG	GGA	GGC	CTG	GCC	GTG	TTG	GCC	GCC	TTT	CTG	GAG	GAG	GGC	CCG	GAA	864
285	E	N	S	A	Y	E	Q	L	L	S	R	L	E	E	I	A	300
865	GAA	AAC	AGT	GCC	TAT	GAG	CAG	TTG	CTG	TCT	CGC	TTG	GAA	GAA	ATC	GCT	912
301	E	E	G	S	E	T	Q	V	P	G	L	D	I	S	A	L	316
913	GAG	GAA	GGC	TCA	GAG	ACT	CAG	GTC	CCA	GGA	CTG	GAC	ATA	TCT	GCA	CTC	960
317	L	P	S	D	F	S	R	Y	F	Q	Y	E	G	S	L	T	332
961	CTG	CCC	TCT	GAC	TTC	AGC	CGC	TAC	TTC	CAA	TAT	GAG	GGG	TCT	CTG	ACT	1008
333	T	P	P	C	A	Q	G	V	I	W	T	V	F	N	Q	T	348
1009	ACA	CCG	CCC	TGT	GCC	CAG	GGT	GTC	ATC	TGG	ACT	GTG	TTT	AAC	CAG	ACA	1056

FIG.-1B

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349 V M L S A K Q L H T L S D T L W 364
 1057 GTG ATG CTG AGT GCT AAG CAG CAG CTC CAC ACC CTC TCT GAC ACC CTG TGG 1104

 365 G P G D S R L Q L N F R A T Q P 380
 1105 GGA CCT GGT GAC TCT CGG CTA CAG CTG AAC TTC CGA GCG ACG CAG CCT 1152

 381 L N G R V I E A S F P A G V D S 396
 1153 TTG AAT GGG CGA GTG ATT GAG GCC TCC TTC CCT GCT GGA GTG GAC AGC 1200

 297 S P R A A E P V Q L N S C L A A 412
 120: AGT CCT CGG GCT GCT GAG CCA CCA GTC CAG CTG AAT TCC TGC CTG GCT GCT 1248

 413 G D I L A L V F G L L F A V T S 428
 1249 GGT GAC ATC CTA GCC CTG GCT TTT GGC CTC CTT TTT GCT GTC ACC AGC 1296

 429 V A F L V Q M R R Q H R R G T K 444
 1297 GTC GCG TTC CTT GTG CAG ATG AGA AGG CAG CAC CAC AGA AGG GGA ACC AAA 1344

 445 G G V S Y R P A E V A E T G A * 460
 1345 GGG GGT GTG AGC TAC CGC CCA GCA GAG GTA GCC GAG ACT GGA GCC TAG 1392

 1393 AGG CTG GAT CTT GGA GAA TGT GAG AAG CCA GCC AGA GGC ATC TGA GGG 1440

 1441 GGA GCC GGT AAC TGT CCT GTC CTG CTC ATT ATG CCA CTT CCT TTT AAC 1488

 1489 TGC CAA GAA ATT TTT TAA AAT AAA TAT TTA TAA T 1522

FIG..1C

FIG..1A

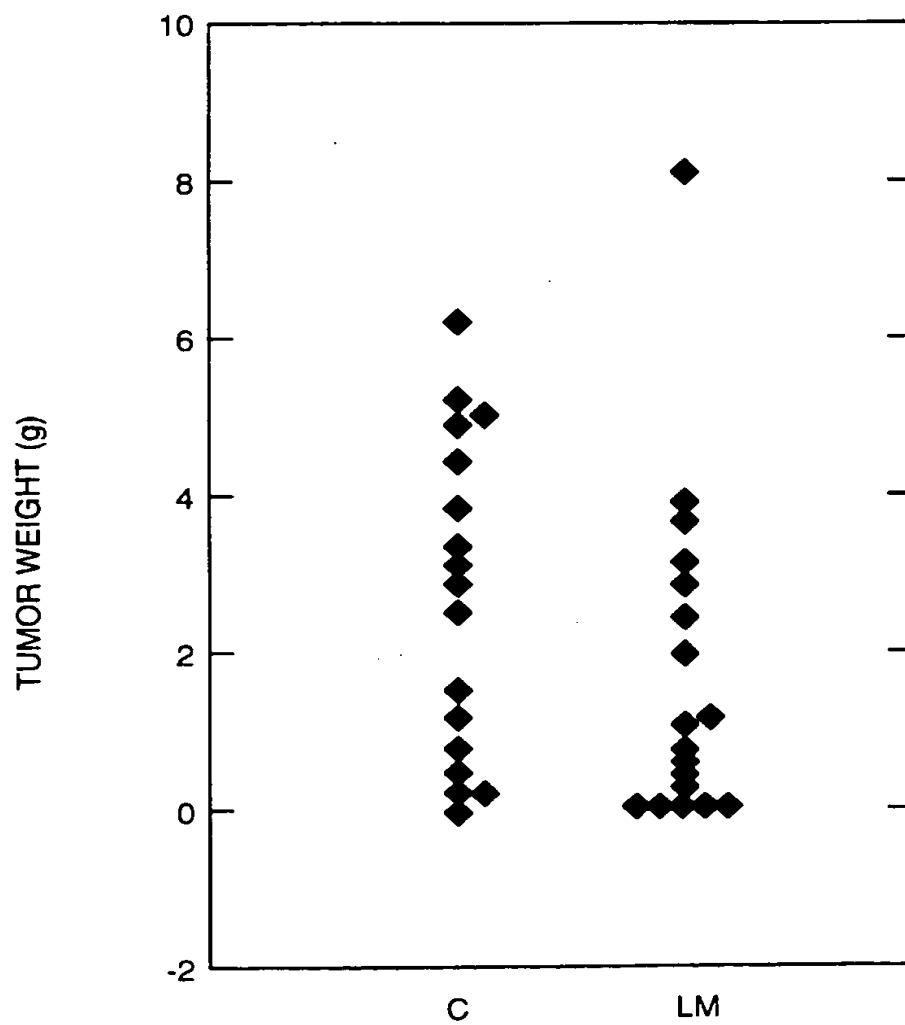
FIG..1B

FIG..1C

FIG..1

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**FIG. 2**

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1 ggatcctgtt gactcgtgac ctaccccca accctgtgct ctctgaaca tgagctgtgt
 61 ccactcaggg ttaaatggat taaggcggt gcaagatgtg cttgttaaa cagatgcttg
 121 aaggcagcat gctcgttaag agtcatacacc aatccctaata ctcaagtaat cagggacaca
 181 aacactgcgg aaggccgcag ggtcctctgc ctaggaaaaa tgaccctgcc aaatccccct ctgtgagaaa
 241 ttatatctgac ctccctcca ctattgtcca taaatgaata gctattggta aagccaaagta
 301 caccacaagaa ttatcaataa aaaaataaat taaaaaaa aatacaaaaa aaaaaaaa
 361 aaaaaaaa gacttacgaa tagttattga atcacagctc aagtcacct gattgatct
 421 aatgatcata ttcaaaacca gacggccatc gattagtcac catcctcaaa attctcccc
 481 ctttatcatt gtcattcttt gattcacta gttacatgaa gcttgaacct actaccttct
 541 aagttctaata tacgtttccaa acatttaggg ttacatgaa ccttacatgc tggggattaa
 601 ttgcttttga gccatgagtt gtaggaatga gcctttggct tatttttcta gctaattttg
 661 tttaaaacttt acctctaagt cagttgggta tgatagtttt cctccacact ttgccactag
 721 tagttaatgg atgcactgtg ttacagtaatt gcttacctaa gaccctaagc cctatttctc
 781 gggtaggtag gtactcagtt aatatgggca tatttaatac aataaattt ttggagtttt
 841 ttgtactggc ctttatctgt tttttttgag acggagtctt gcatctgtca tggccaggct
 901 ttgttttgtt tggtgcccac tcggctcact gcaagctcca cctcccgagt tcacgccatt
 961 ggagtagcag tggtgcccac gtagctggga ctacaggcg cccagaccat gccgggctaa
 1021 ttctgcctc agcctccga gacggggttt caccgtgtta gccagaaagg tctcgatctc
 1081 ttttttgtat ttttggtaga gacggggttt ccaagttctt ccaagttacag gtgtgagcca
 1141 ctgacttcgt gatccaccg ccaattttt gagtctttta aagtaaaaat atgtcttcta agctggtaac
 1201 ccgacactgg ccaattttt taatgtggtg ctgacgggtca tatagggtct tttgagttg
 1261 tatggtacat ttctttttat gtagtccttt cattacattt ttctctcttc atttgaagag
 1321 gcatgcataat gctacttttt cacttggtt aaaaagggtt ctcattagcc taacacagt
 1381 catgttatat taccacttgg atcataagt gaaaaacagt caagaaattg cacagtaata
 1441 tcattgttgg agaggatga ttcagggtgaa tctgacacta agaaactccc ctacctgagg
 1501 cttgttttga ctctgacatt gctgtatata ggcttttctt ttgacagcct gtgactgcgg
 1561 tctgagattc ttaagcaaga tatgctaaag ttttgtgagc ctttttccag agagaggtct
 1621 actatatttc tcaagtgaaga acatataatg gaaacttgtt cctcagtgc ccaaaagagg
 1681 catatctgca tatgctttta tatagacagg ccacgctttc tgaccttggg aacaataag
 1741 gcttgtgttt tatgttatat catcattggc tacaagaaat agctgctatg tttcttgaca
 1801 tgggaattgt tatggatat ctcaattctg taggaaataa gaatgtgaaa ctcttcagtt
 1861 ggttcataat ctcaattctg taggaaataa gaatgtgaaa ctcttcagtt
 1921 tccacttgg taggaaataa gaatgtgaaa ctcttcagtt

FIG.-3A

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1981 ttgcaatttc cttcttactg tgtaaaaaa aagtatgac ttgctctgag aggtgaggca
2041 ttcttaatca tgatctttaa agatcaataa tataatcctt tcaaggatta tgtctttatt
2101 ataataaaga taatttgtct ttaacagaat caataatata atcccttaaa ggattatatac
2161 ttgctgggc gcagtggctc acacctgtaa tcccagcact ttgggtggcc aaggtggaag
2221 gatcaaatct gcctacttct atattatctt ctaaagcaga attcatctct ctccctcaa
2281 tatgatgata ttgacagggt ttgacctcac tcaatagatt gtgagctcct gctcagggca
2341 ggtagcgttt ttgtgttttg tttttgtttt tcttttttga gacagggtct tgctctgtca
2401 ccaggccag agtgcaatgg tacagtctca gctcactgca gctcaaccg cctcggctca
2461 aaccatcatc ccatttcagc ctcttgagta gctgggacta caggcacatg ccattacacc
2521 tggctaattt ttttgtattt ctagttaga cagggttttg ccatgttgcc cgggctggtc
2581 tcgaactcct ggactcaagc aatccacca cctcagcctc ccaaatgag ggaccgtgtc
2641 ttattcatct ccatgtccct agtccatagc ccagtgtgga acctatggtg gtactaaata
2701 aatatttgtt gaatgcaata gtaaatagca tttcaggag caagaactag attaacaag
2761 gtggtaaaag gtttgagaa aaaaataata gtttaatttg gctagagtat gagggagagt
2821 agtaggagac aagatggaaa ggtctcttgg gcaagggttt gaaggagtt ggaagtcaga
2881 agtacacaat gtgcatactg tggcaggcag tggggagcca atgaaggctt ttgagcagga
2941 gagtaatgtg ttgaaaaata aatataggtt aaacctatac gagccccctc gacacataca
3001 cttgcttttc attcaagctc aagtttgtct ccacataacc cattacttaa ctacccctcg
3061 ggctcccta gcagcctgcc ctacctctt acctgcttcc tggaggagtc agggatgtat
3121 acatgagctg ctttccctct cagccagagg acatgggggg cccagctcc cctgaccttc
3181 ccttctgtg cctggagctg ggaagcaggc cagggttagc tgaggctggc tggcaagcag
3241 ctgggttggt ccagggagag cctgcatagt gccagggtgt gccttgggtt ccaagctagt
3301 ccatggcccc gataaccttc tgcctgtgca cacacctgcc cctcactcca ccccatcct
3361 agcttttggt tgggggagag ggcacagggc cagacaaacc tgtgagactt tggctccatc
3421 tctgcaaaag ggcgctctgt gagtacgctt gctccctcc aggtttgctc ctccccacc
3481 cagctctcgt ttccaatgca cgtacagccc gtacacaccc tgtgctggga caccacacag
3541 TCAGCCGCAT GGCTCCCCTG TGCCCCAGCC CCTGGCTCCC TCTGTTGATC CCGCCCCCTG
3601 CTCAGGCCCT CACTGTGCAA CTGCTGCTGT CACTGCTGCT TCTGGTGCCCT GTCCATCCCC
3661 AGAGGTTGCC CCGGATGCAG GAGGATCCC CCTTGGGAGG AGGCTCTTCT GGGGAAGATG
3721 ACCCACTGGG CGAGGAGGAT CTGCCCCAGTG AAGAGGATTC ACCCAGAGAG GAGGATCCAC
3781 CCGGAGAGGA GGATCTACCT GGAGAGGAGG ATCTACCTGG AGAGGAGGAT CTACCTGAAG
3841 TTAAGCCCTAA ATCAGAAGAA GAGGGCTCCC TGAAGTTAGA GGATCTACCT ACTGTTGAGG
3901 CTCCTGGAGA TCCTCAAGAA CCCAGAATA ATGCCCACAG GGACAAAGAA Ggtaagtgg

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3961 catcaatctc caaatccagg ttccaggagg ttcatgactc cctcccata cccagccta
4021 ggctctgttc actcaggga ggaggggaga ctgtactccc cacagaagcc cttccagagg
4081 tccatacca atatcccat cccactctc ggaggtagaa agggacagat gtggagagaa
4141 aataaaaaagg gtgcaaaaagg agagaggtga gctggatgag atgggagaga aggggaggc
4201 tggagaagag aaagggatga gaactgcaga tgagagaaa tgagggaag agcttgggag
4261 aaataggtag agaaggagag taccagagac aagcaagaag agctggtaga agtcatctca tcttaggcta
4321 gtgaagtggg taccagagac ttgagacctg ggaagaaggg acacagcagg tagagaaacg tggcttcttg
4381 caatgaggaa ttgagacctg caggaatttg ggaagaaggg ttggagacca tacaaggcag agggatgagt
4441 actcccaagc caggaatttg ggaagaaggg ggaagaaggg tgggtgactc actcatttg gactcaggac
4501 ggggagaaga aagaagggag aaaggaaaga aaaggaaaga tgggtgactc aaactttcac ttttgttgcc
4561 tgaagtggc actcactttt tttttttttt tttttttttt tttttgagac aaactttcac ttttgttgcc
4621 caggctggag tgcaatggcg tgcaatggcg cgaatctcggc tcaactgcaac ctccacctcc cgggttcaag
4681 tgattctcct gcctcagcct ctatcagggtg atccaaccac cctggcctcc caaagtgtcg ggattatagg
4741 ccggctaatt tttgtatttt ttgtattttt tagtagagac ggggttttcgc catgttggtc aggtgggtct
4801 cgaactcctg atctcagggtg atccaaccac cctggcctcc cactcacttt tacagacctt aagacaatga
4861 cgtgagccac agcgctggcg gttagattgc tgtttggccc accagctgc ggtgttagt ttgggtgcgg
4921 ttgcaagctg ttgacacctg ttgacacctg gcccgttaa ggcatttgtt acccgtaatg ctccgtgaag
4981 tctcctgtgc ttgtgacatc ttgttggtcg gttttgggtc ccaggaaggg attggggctc taagcttgag
5041 gcatctgcgt ttgtgacatc ttgtgacatc ttgttggtcg gttttgggtc ccaggaaggg attggggctc taagcttgag
5101 cggttcaccc ttgtgacatc ttgtgacatc ttgttggtcg gttttgggtc ccaggaaggg attggggctc taagcttgag
5161 acaccaccc gctgcacaga ccaatctgg gaaccagct ctgtggatct cccctacagc cccctacacg
5221 cgtccctgaa cactggtccc gggcgtccc gggcgtccc cccgtcccac accgtcccac cccctcacct
5281 tttctaccg ggttccctaa gttcctgacc gttcctgacc taggcgtcag acttccctcac tatactctcc
5341 caccacagc GACCCGCCCT GACCCGCCCT GACCCGCCCT GACCCGCCCT GACCCGCCCT GACCCGCCCT
5401 CCGGTGGAT ATCGCCCC ATCGCCCC ATCGCCCC ATCGCCCC ATCGCCCC ATCGCCCC
5461 CCTGGGCTTC CAGCTCCCGC CAGCTCCCGC CAGCTCCCGC CAGCTCCCGC CAGCTCCCGC CAGCTCCCGC
5521 tgagggggtc tcccgcgga gacttgggga tggggcgggg cgcagggaag ggaaccgtcg
5581 cgcagtgcct gcccgggggt tgggctggcc ctaccgggag ctaccgggag gggccgggctc acttgcctct
5641 ccctacgcag TGCAACTGAC CCTGCCCTCCT CCTGCCCTCCT CCTGCCCTCCT CCTGCCCTCCT CCTGCCCTCCT
5701 GAGTACCGGG CTCTGCAGCT GCATCTGCAC TGGGGGGGCTG TGGGGGGGCTG TGGGGGGGCTG TGGGGGGGCTG
5761 CACACTGTGG AAGCCACCG TTTTCCCTGCCC GAGgtgagcg cggactggcc gagaaggggc
5821 aaaggagcgg ggcggacggg ggcagagac gtggccctct cctaccctcg tgtccttttc
5881 agATCCACGT GGTTCACCTC AGCACCGCCT TTGCCAGAGT TGACGAGGCC TTGGGGCGCC

FIG.-3C

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5941 CGGAGGCCCT GGCCGTGTTG GCCGCCTTTC TGGAGgtacc agatcctgga caccacctac
6001 tccccgcttt cccatcccat gctcctccc gactctatcg tggagccaga gaccccatcc
6061 cagcaagctc actcaggccc ctggctgaca aactcattca cgcactgttt gttcatttaa
6121 caccactgt gaaccaggca ccagccccc ccagccccc tgaagctgta ggtccttgcc
6181 tctaaggagc ccacagccag tgggggaggg tgacatgaca gacacatagg aaggacatag
6241 taagatgggt ggtcacagag gaggtgacac ttaaagcctt cactggtaga aaagaaaagg
6301 aggtgttcat tgcagaggaa acagaatgtg caaagactca gaatatggcc tatttaggga
6361 atggctacat acaccatgat tagaggaggc ccagtaaaagg gaagggatgg tgagatgcct
6421 gctaggttca ctactcact tttatttatt tatttatttt ttgacagtc tctctgtcgc
6481 ccaggctgga gtgcagtgggt gtgactttgg gtcactgcaa cttccgcctc cggggttcaa
6541 gggattctcc tgcctcagct tcctgagtag ctggggttac aggtgtgtgc caccatgccc
6601 agctaatttt tttttgtatt tttagtagac aggttttcac catgttggtc aggtggtct
6661 caaactcctg gcctcaagtg atccgcctga ctcagcctac caaagtgtg attacaagt
6721 tgagccaccg tgcccagcca cactcactga ttctttaatg ccagccacac agcacaagt
6781 tcagagaaat gcctccatca tagcatgtca atatgttcat actcttaggt tcatgatgtt
6841 cttaacatta ggttcataag caaataaga aaaaagaata ataaataaaa gaagtggcat
6901 gtcaggacct cactgaaa gccaacacac gaatcatgaa ggtgaatgca gaggtgacac
6961 caacacaaag gtgtatatat ggtttcctgt ggggagtatg tacggaggca gcagtgagt
7021 agactgcaaa cgtcagaagg gcacgggtca ctgagagcct agtatcctag taaagtgggc
7081 tctctccctc tctctccagc ttgtcattga aaaccagtcc accaagctg ttggttcgca
7141 cagcaagagt acatagagtt tgaataata cataggattt taagaggagg acactgtctc
7201 taaaaaaa aacaacagca acaaaaaa gcaacaacca ttacaatttt atgttccctc
7261 agcatctca gagctgagga atgggagagg actatggga ccccttcat gttccggcct
7321 tcagccatgg ccctggatc atgcactcat ctgtcttaca atgtcattcc ccagGAGGG
7381 CCCGAAGAA AACAGTGCCT ATGAGCAGTT GCTGTCTCGC TTGGAAGAAA TCGCTGAGGA
7441 AGgtcagttt gttggtctgg ccactaatct ctgtggccta gttcataaag aatcacctt
7501 tggagcttca ggtctgaggc tggagatggg ctccctccag tgcaggaggg attgaagcat
7561 gagccagcgc tcatcttgat aataaccatg aagctgacag acacagtac ccgcaaacgg
7621 ctgcctacag attgaaaacc aagcaaaaac cgccgggcac ggtggtcac gcctgtaac
7681 ccagcacttt gggaggccaa ggcaggtgga tcacgaggtc aagagatcaa gaccatcctg
7741 gccaacatgg tgaaacccca tctctactaa aaatacgaaa aaatagccag gcgtggtggc
7801 ggggtgcctgt aatcccagct actcgggagg ctgaggcagg agaattggcat gaacccggga
7861 ggcagaagtt gcagtgagcc gagatcgtgc cactgcactc cagcctgggc aacagagcga

FIG._3D

SUBSTITUTE SHEET (RULE 26)

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7921 gactcttgct tcaaaaaaa aaaaaaaa gaaaccaa gaaaccaa aatgagacaa
7981 aaaaaacaag accaaaaaat ggtgtttgga aattgtcaag gtcaagtctg gagagctaaa
8041 ctttttctga gaactgttta tctttaataa gcatcaataa ttttaacttt gtaaatactt
8101 ttgttggaat tagttctctt cttagtcaat cttagtcaat ttttaacttc acttactcta
8161 ctgaccttt taggtttctg ctgacctag ttacaagt tttagtaga ccttgcat tcttgctct
8221 gttttgtata gttatcaata ttatatttta tttagtaga attcagatca ttttttctt
8281 tctttttttt tttttttttt tttttttttt cttagtaga gacagggttt caccatattg
8341 gccaggctgc tctcaaac tctgacctgt gatccaccag cctcggcctc ccaaagtgc
8401 gggattcatt ttttctttt aatttgctct gggcttaaac ttgtggccca gcactttatg
8461 atggtacaca gagttaagag tgtagactca gacggtcttt ctctttctct tctcttctt
8521 cctcccttcc ctccacctt ccttctctc caagccctg tactttttt ttgagttaacg tcttatggga
8581 caggcctctt ccagttgctc ctagtgctc aagtggtctc agagttgagt taccttggct tctgggaggt
8641 agggcctgca cttagtgaag aagtggtctc aagtggtgca atgtagatga gacccaaca
8701 gaaactgtat ccctataccc tgaagcttta aggggtgca GTGGACATATC TGGACTCCTG
8761 tagatcctct tcacagGCTC AGAGACTCAG GTCCACAGGAC TGACTACACC GCCCTGTGCC
8821 CCTCTGACT TCAGCCGCTA CTTCCAATAT GAGGGTCTC TGAGTGCTAA GCAGtgggc
8881 CAGGGTGCTA TCTGGACTGT GTTTAACCAG ACAGTGATGC TGAGTGCTAA GCAGtgggc
8941 ctggggtgtg tgtggacaca gttgggtgctg gggaaagagg atgtaagatg agatgagaaa
9001 caggagaaga aagaaatcaa ggtgggctc tgtggcttac gcctataatc ccaccagtt
9061 gggaggctga ggtgggagaa tgggttgagc ccaggagttc aagacaaggc ggggcaacat
9121 agtgtgacct catctctacc aaaaaaac tactcaagg ggtgaggtg ggaagatcgc ttgattccag
9181 gtatgcggcc tagtcccagc ttagtgatccc aagaggctgg atggggaata caggagctgg agggtgaggc
9241 gagtttgaga ctgcagtgag ctatgatccc ctggtggtg acccttctta ggatacatct
9301 atttatttat aaagaaatc aagaggctgg atggggaata caggagctgg agggtgaggc
9361 cctgagggtgc tgggttgtag ctggcctggg acccttctta cctgtcatgc catgaacca
9421 cccacactgt ccactgacct ccctagCTCC ACACCTCTC TGACACCTG TGGGACCTG
9481 GTGACTCTCG GCTACAGCTG AACTTCCGAG CGACGACGCC TTTGAATGGG CGAGTGATTG
9541 AGGCTCCTT CCTGTGGA GTGGACAGCA GTCCTCGGC TGCTGAGCCA Ggtacagctt
9601 tgtctggttt ccccccagcc agtagtccct tatcctccca ttgtgtgtgc agtgtctgtc
9661 attggtgggc acagcccgc tctcacatct ccttttctc ttggtcctga tggcaggaga ctcctcagca
9721 GCCTGGCTGC TGgtgagctt gcccctctc ttggtcctga tggcaggaga ctcctcagca
9781 ccattcagcc ccagggtgc tcaggaccgc ctctgctccc tctcctttc tgcagaacag
9841 accccaacc caatataga gaggcagatc atggtgggga tccccatt gtccccagag

FIG._3E

SUBSTITUTE SHEET (RULE 26)

10 / 14

9901 gctaattgat tagaatgaag cttgagaaat ctcccagcat ccctctcgca aaagaatccc
9961 cccccctttt tttaagata gggctctcact ctgtttgccc caggctgggg tgttgtggca
10021 cgatcatagc tcactgcagc ctcgaactcc taggctcagg caatccttc accttagctt
10081 ctcaagcac tgggactgta ggcatagacc actgtgacct gcccacaacg gcccttttac
10141 ttggctttta ggaagcaaaa acggtgctta tcttaccctt tctcgtgtat ccaccctcat
10201 cccttggctg gcctcttctg gagactgagg cactatgggg ctgcctgaga actcggggca
10261 ggggtggtgg agtgcactga ggcaggtgtt gaggaactct gcagaccctt ctctcttccc
10321 aaagcagccc tctctgctct ccatcgcagg TGACATCCTA GCCCTGGTTT TTGGCCTCCT
10381 TTTTGCTGTC ACCAGCGTCG CGTTCCTTGT GCAGATGAGA AGGCAGCACA Ggtattacac
10441 tgacccttc ttcaggcaca agcttcccc acccttgtgg agtcaactca tgcaaacgc
10501 atgcaaatga gctgctcctg ggccagtttt ctgattagcc ttctcctgtt tgtaacacaca
10561 GAAGGGGAAC CAAAGGGGT GTGAGCTACC GCCCAGCAGA GGTAGCCGAG ACTGGAGCCT
10621 AGAGGCTGA TCTTGGAGAA TGTGAGAAGC CAGCCAGAGG CATCTGAGGG GGAGCCGGTA
10681 ACTGTCCTGT CCTGCTCATT ATGCCACTTC CTTTAACTG CCAAGAAATT TTTTAAATA
10741 AATATTATA Aataaatatg tgtagtcac ctttgttccc caaatcagaa ggaggtatct
10801 gaatttccta ttactgttat tagcaccaat ttagtggtaa tgcatttatt ctattacagt
10861 tcggcctcct tccacacatc actccaatgt gttgctcc

FIG._3F

FIG._3A

FIG._3B

FIG._3C

FIG._3D

FIG._3E

FIG._3F

FIG._3



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-506 CTTGCTTTTC ATTCAAGCTC AAGTTTGTCT CCCACATACC CATTACTAA CTCACCCTCG
-446 GGCTCCCCTA GCAGCCTGCC CTACCTCTTT ACCTGCTTCC TGGTGGAGTC AGGGATGTAT
AP2
-386 ACATGAGCTG CTTTCCCTCT CAGCCAGAGG ACATGGGGG CCCAGCTCC CCTGCCCTTC
-326 CCGTTCTGTG CCTGGAGCTG GGAAGCAGGC CAGGGTTAGC TGAGGCTGGC TGGCAAGCAG
-266 CTGGGTGGTG CCAGGGAGAG CCTGCATAGT GCCAGGTGGT GCCTTGGGTT CCAAGCTAGT
p53
-206 CCATGGCCCC GATAACCTTC TGCCTGTGCA CACACCTGCC CCTCACTCCA CCCCCATCCT
Inr
-146 AGCTTTGGTA TGGGGGAGAG GGCACAGGGC CAGACAAACC TGTGAGACTT TGGCTCCATC
Inr
-86 TCTGCAAAAG GCGGCTCTGT GAGTCAGCCT GCTCCCCCTCC AGGTTGCTC CTCCCCCACC
AP1 p53 AP2
-26 CAGCTCTCGT TTCCAATGCA CGTACAGCCC GTACACACCG TGTGCTGGA CACCCACAG

...

FIG._6

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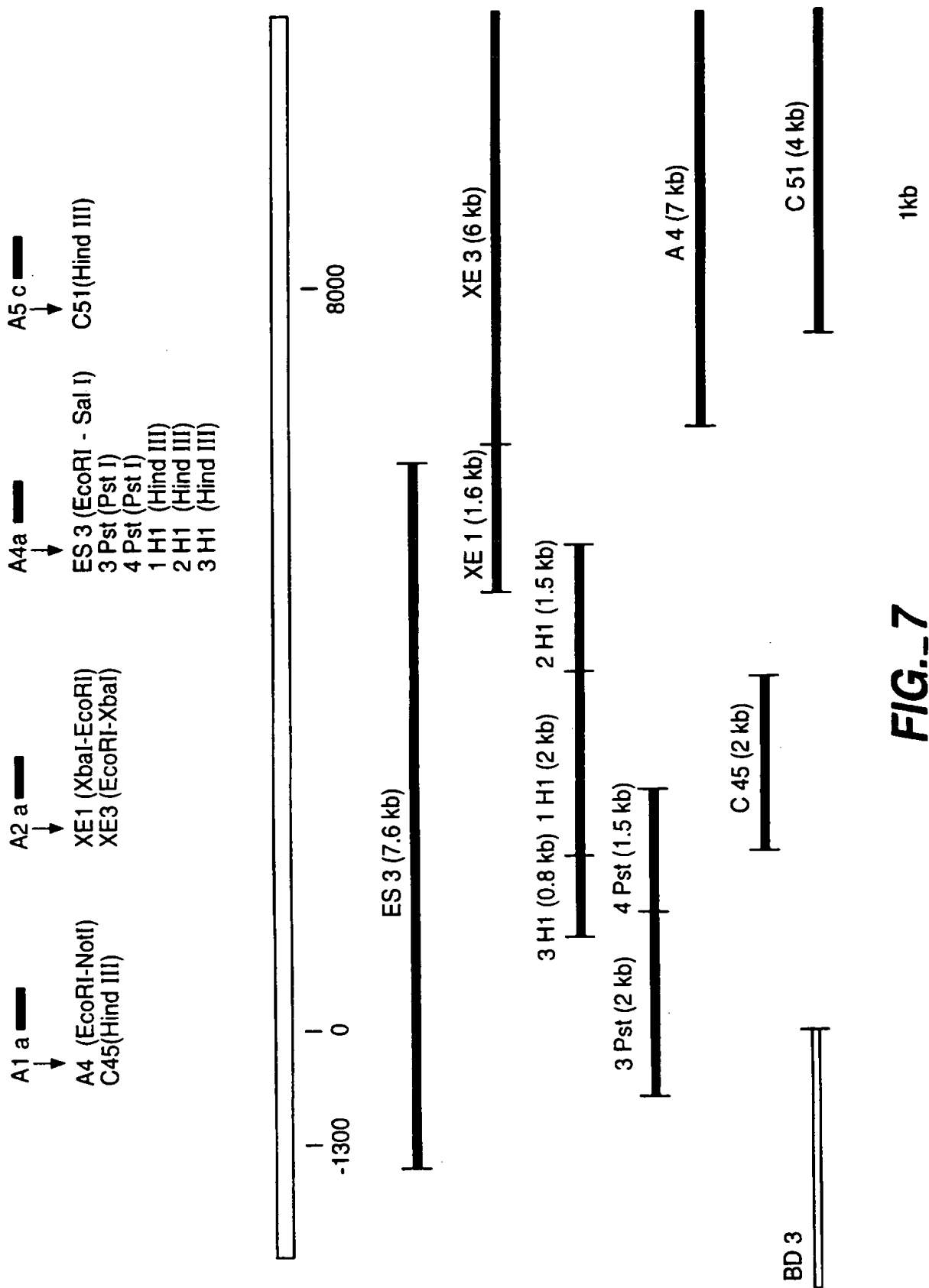
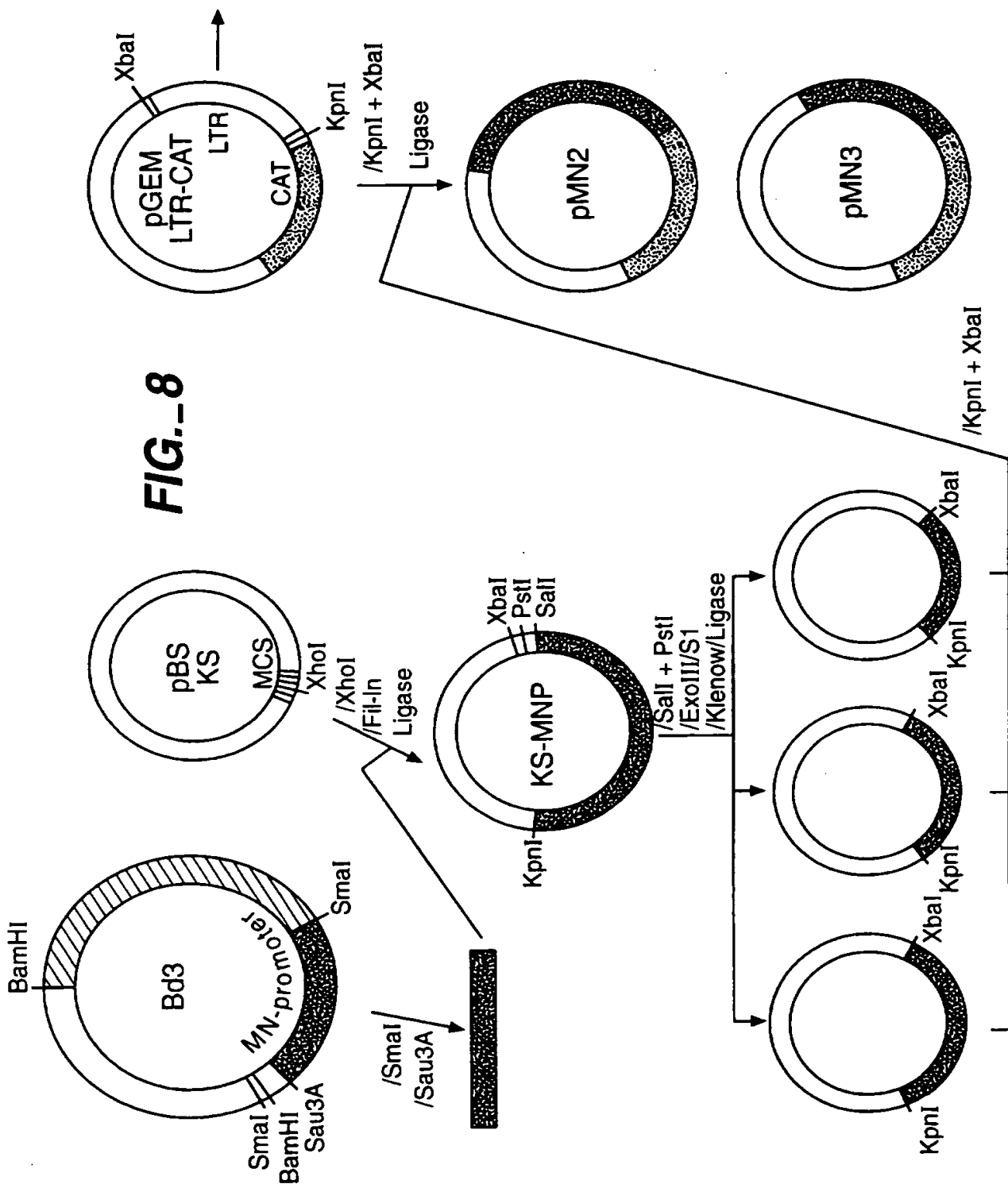


FIG. 7

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SUBSTITUTE SHEET (RULE 26)

INTERNATIONAL SEARCH REPORT

International Application No.
PCT/US 007628

A. CLASSIFICATION OF SUBJECT MATTER

IPC 6 C12N15/12 C12N1/21 C12N15/62 C12Q1/68 A61K39/00
C07K14/82 C12N5/16 C12N15/87 C07K16/30 G01N33/50
A61K48/00

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 C12N A61K C07K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO,A,93 18152 (CIBA CORNING DIAGNOSTICS CORP. AND INSTITUTE OF VIROLOGY) 16 September 1993 cited in the application see the whole document ---	1-14, 16-18, 21-27
X	PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF USA, vol. 85, December 1988 WASHINGTON US, pages 8998-9002, M.A.FROHMAN ET AL. 'Rapid production of full-length cDNAs from rare transcripts: Amplification using a single gene-specific oligonucleotide primer' see page 8998 --- -/--	10,28

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

* Special categories of cited documents:

- *A* document defining the general state of the art which is not considered to be of particular relevance
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T later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

X document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

Y document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

A document member of the same patent family

Date of the actual completion of the international search

4 January 1996

Date of mailing of the international search report

16. 01 96

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Fax (+31-70) 340-3016

Authorized officer

Cupido, M

INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 95/07628

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	<p>VIROLOGY, vol. 187, no. 2, April 1992 ORLANDO US, pages 620-626, S.PASTOREKOVA ET AL. 'A novel quasi-viral agent, MaTu, is a two-component system' see the whole document ---</p>	1-28
P,X	<p>ONCOGENE, vol. 9, no. 10, October 1994 pages 2877-2888, J.PASTOREK ET AL. 'Cloning and characterization of MN, a human tumor-associated protein with a domain homologous to carbonic anhydrase and a putative helix-loop-helix DNA binding segment' see the whole document -----</p>	1-14, 16-18, 21-27

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US/07628

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This international search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☒ Claims Nos.: 18,20,21,26
because they relate to subject matter not required to be searched by this Authority, namely:
Remark: Although claims 18,20,21 and 26 are directed to methods of treatment of, and diagnostic methods practised on the human body the search has been carried out and based on the alleged effects of the composition.
2. ☐ Claims Nos.:
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
- ☐ No protest accompanied the payment of additional search fees.

ormation on patent family members

PCT/US 95/07628

Form PCT/ISA/210 (patent family annex) (July 1992)



INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁶ :

C12N 15/12, 1/21, 15/62, C12Q 1/68,
A61K 39/00, C07K 14/82,
C12N 5/16, 15/87, C07K 16/30,
G01N 33/50, A61K 48/00

A3

(11) International Publication Number:

WO 95/34650

(43) International Publication Date: 21 December 1995 (21.12.95)

(21) International Application Number: PCT/US95/07628

(22) International Filing Date: 15 June 1995 (15.06.95)

(30) Priority Data:

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08/485,049	7 June 1995 (07.06.95)	US
08/486,756	7 June 1995 (07.06.95)	US
08/447,504	7 June 1995 (07.06.95)	US
08/481,658	7 June 1995 (07.06.95)	US
08/485,862	7 June 1995 (07.06.95)	US
08/485,863	7 June 1995 (07.06.95)	US
08/487,077	7 June 1995 (07.06.95)	US

(71) Applicant (for AM BF BJ CM MG MR TD TG only): CIBA CORNING DIAGNOSTICS CORP. [US/US]; 63 North Street, Medfield, MA 02052 (US).

(71) Applicant (for all designated States except US): INSTITUTE OF VIROLOGY [SK/SK]; Slovak Academy of Sciences, Dubravska Cesta 9, 84 246 Bratislava (SK).

(72) Inventors; and

(75) Inventors/Applicants (for US only): ZAVADA, Jan [CZ/CZ]; Na pekne vyhlidce 1, 160 00 Prague 6 (CZ). PASTOREKOVA, Silvia [SK/SK]; I. Bukovcna 18, 841 07 Bratislava (SK). PASTOREK, Jaromir [SK/SK]; I. Bukovcna 18, 841 07 Bratislava (SK).

(74) Agent: LAUDER, Leona, L.; 6 Mariposa Court, Tiburon, CA 94920-2017 (US).

(81) Designated States: AM, AT, AU, BB, BG, BR, BY, CA, CH, CN, CZ, DE, DK, EE, ES, FI, GB, GE, HU, IS, JP, KE, KG, KP, KR, KZ, LK, LR, LT, LU, LV, MD, MG, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, TJ, TM, TT, UA, UG, US, UZ, VN, European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG), ARIPO patent (KE, MW, SD, SZ, UG).

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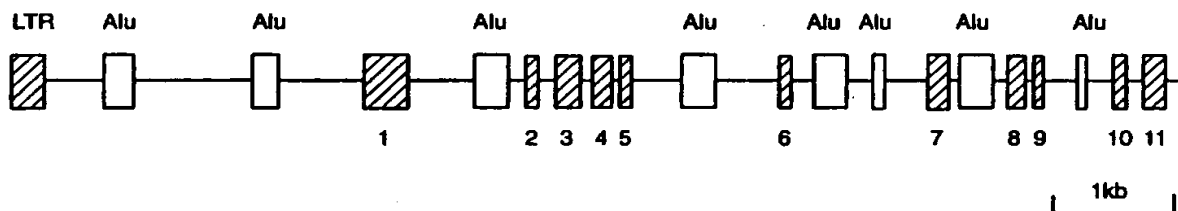
With international search report.

Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.

(88) Date of publication of the international search report:

15 February 1996 (15.02.96)

(54) Title: MN GENE AND PROTEIN



(57) Abstract

A complete genomic sequence including a full-length cDNA sequence for the MN gene, a putative oncogene, is disclosed, as well as proteins/polypeptides encoded thereby. Recombinant nucleic acid molecules for expressing MN proteins/polypeptides and recombinant proteins are also provided. Expression of the MN gene is disclosed as being associated with tumorigenicity, and the invention concerns methods and compositions for detecting and/or quantitating MN antigen and/or MN-specific antibodies in vertebrate samples that are diagnostic/prognostic for neoplastic and preneoplastic disease. Test kits embodying the immunoassays of this invention are provided. MN-specific antibodies are disclosed that can be used diagnostically/prognostically, therapeutically, for imaging, and/or for affinity purification of MN proteins/polypeptides. Also provided are nucleic acid probes for the MN gene as well as test kits comprising said probes. The invention also concerns vaccines comprising MN proteins/polypeptides which are effective to immunize a vertebrate against neoplastic diseases associated with the expression of MN proteins. The invention still further concerns antisense nucleic acid sequences that can be used to inhibit MN gene expression, and polymerase chain reaction (PCR) assays to detect genetic rearrangements in MN genes.

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GA	Gabon				

INTERNATIONAL SEARCH REPORT

International Application No.
PCT/US 93/07628

A. CLASSIFICATION OF SUBJECT MATTER

IPC 6	C12N15/12	C12N1/21	C12N15/62	C12Q1/68	A61K39/00
	C07K14/82	C12N5/16	C12N15/87	C07K16/30	G01N33/50
	A61K48/00				

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 C12N A61K C07K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

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X	PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF USA, vol. 85, December 1988 WASHINGTON US, pages 8998-9002, M.A.FROHMAN ET AL. 'Rapid production of full-length cDNAs from rare transcripts: Amplification using a single gene-specific oligonucleotide primer' see page 8998 --- -/--	10,28

☒ Further documents are listed in the continuation of box C.☒ Patent family members are listed in annex.

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- *A* document member of the same patent family

Date of the actual completion of the international search

4 January 1996

Date of mailing of the international search report

16. 01 96

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Fax (+31-70) 340-3016

Authorized officer

Cupido, M

INTERNATIONAL SEARCH REPORT

International Application No
PCT/US 95/07628

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	<p>VIROLOGY, vol. 187, no. 2, April 1992 ORLANDO US, pages 620-626, S.PASTOREKOVA ET AL. 'A novel quasi-viral agent, MaTu, is a two-component system' see the whole document ---</p>	1-28
P,X	<p>ONCOGENE, vol. 9, no. 10, October 1994 pages 2877-2888, J.PASTOREK ET AL. 'Cloning and characterization of MN, a human tumor-associated protein with a domain homologous to carbonic anhydrase and a putative helix-loop-helix DNA binding segment' see the whole document -----</p>	1-14, 16-18, 21-27

INTERNATIONAL SEARCH REPORT

International Application No.

PCT/US95/07628

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This international search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☒ Claims Nos.: 18,20,21,26
because they relate to subject matter not required to be searched by this Authority, namely:
Remark: Although claims 18,20,21 and 26 are directed to methods of treatment of, and diagnostic methods practised on the human body the search has been carried out and based on the alleged effects of the composition.
2. ☐ Claims Nos.:
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3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
- ☐ No protest accompanied the payment of additional search fees.

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/US 95/07628

Patent document cited in search report	Publication date	Patent family member(s)		Publication date
WO-A-9318152	16-09-93	CA-A-	2131826	16-09-93
		EP-A-	0637336	08-02-95
		JP-T-	7508160	14-09-95
		NO-A-	943344	09-09-94
		US-A-	5387676	07-02-95



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(51) International Patent Classification⁶:

C12N 15/12, 1/21, 15/62, C12Q 1/68,
A61K 39/00, C07K 14/82,
C12N 5/16, 15/87, C07K 16/30,
G01N 33/50, A61K 48/00

A3

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WO 95/34650

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21 December 1995 (21.12.95)

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15 June 1995 (15.06.95)

(30) Priority Data:

08/260,190	15 June 1994 (15.06.94)	US
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08/486,756	7 June 1995 (07.06.95)	US
08/447,504	7 June 1995 (07.06.95)	US
08/481,658	7 June 1995 (07.06.95)	US
08/485,862	7 June 1995 (07.06.95)	US
08/485,863	7 June 1995 (07.06.95)	US
08/487,077	7 June 1995 (07.06.95)	US

(71) Applicant (for AM BF BJ CM MG MR TD TG only): CIBA
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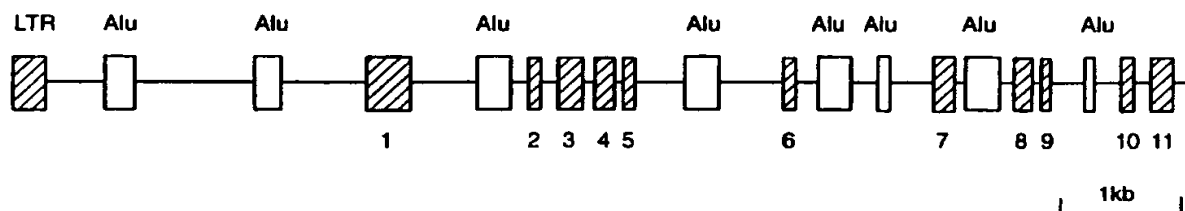
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57) Abstract

A complete genomic sequence including a full-length cDNA sequence for the MN gene, a putative oncogene, is disclosed, as well as proteins/polypeptides encoded thereby. Recombinant nucleic acid molecules for expressing MN proteins/polypeptides and recombinant proteins are also provided. Expression of the MN gene is disclosed as being associated with tumorigenicity, and the invention concerns methods and compositions for detecting and/or quantitating MN antigen and/or MN-specific antibodies in vertebrate samples that are diagnostic/prognostic for neoplastic and preneoplastic disease. Test kits embodying the immunoassays of this invention are provided. MN-specific antibodies are disclosed that can be used diagnostically/prognostically, therapeutically, for imaging, and/or for affinity purification of MN proteins/polypeptides. Also provided are nucleic acid probes for the MN gene as well as test kits comprising said probes. The invention also concerns vaccines comprising MN proteins/polypeptides which are effective to immunize a vertebrate against neoplastic diseases associated with the expression of MN proteins. The invention still further concerns antisense nucleic acid sequences that can be used to inhibit MN gene expression, and polymerase chain reaction (PCR) assays to detect genetic rearrangements in MN genes.

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MN GENE AND PROTEIN
FIELD OF THE INVENTION

The present invention is in the general area of medical genetics and in the fields of biochemical engineering and immunochemistry. More specifically, it relates to the identification of a new gene--the MN gene--a cellular gene
5 coding for the MN protein. The inventors hereof found MN proteins to be associated with tumorigenicity. Evidence indicates that the MN protein appears to represent a potentially novel type of oncoprotein. Identification of MN antigen as well as antibodies specific therefor in patient
10 samples provides the basis for diagnostic/prognostic assays for cancer.

BACKGROUND OF THE INVENTION

A novel quasi-viral agent having rather unusual properties was detected by its capacity to complement mutants
15 of vesicular stomatitis virus (VSV) with heat-labile surface G protein in HeLa cells (cell line derived from human cervical adenocarcinoma), which had been cocultivated with human breast carcinoma cells. [Zavada et al., Nature New Biol., 240: 124 (1972); Zavada et al., J. Gen. Virol., 24: 327 (1974);
20 Zavada, J., Arch. Virol., 50: 1 (1976); Zavada, J., J. Gen. Virol., 63: 15-24 (1982); Zavada and Zavadova, Arch. Virol., 118: 189 (1991).] The quasi viral agent was called MaTu as it was presumably derived from a human mammary tumor.

There was significant medical interest in studying
25 and characterizing MaTu as it appeared to be an entirely new type of molecular parasite of living cells, and possibly originated from a human tumor. Zavada et al., International Publication Number WO 93/18152 (published 1 September 1993), describes the elucidation of the biological and molecular
30 nature of MaTu which resulted in the discovery of the MN gene and protein. MaTu was found by the inventors to be a two-

component system, having an exogenous transmissible component, MX, and an endogenous cellular component, MN. The MN component was found to be a cellular gene, showing only very little homology with known DNA sequences. The MN gene was
5 found to be present in the chromosomal DNA of all vertebrates tested, and its expression was found to be strongly correlated with tumorigenicity.

The exogenous MaTu-MX transmissible agent was identified as lymphocytic choriomeningitis virus (LCMV) which
10 persistently infects HeLa cells. The inventors discovered that the MN expression in HeLa cells is positively regulated by cell density, and also its expression level is increased by persistent infection with LCMV.

Research results provided herein show that cells
15 transfected with MN cDNA undergo changes indicative of malignant transformation. Further research findings indicate that the disruption of cell cycle control is one of the mechanisms by which MN may contribute to the complex process of tumor development.

20 Described herein is the cloning and sequencing of the MN gene and the recombinant production of MN proteins. The full-length MN cDNA sequence [SEQ. ID. NO.: 1], the amino acid sequence deduced therefrom [SEQ. ID. NO.: 2], a full-length genomic sequence for MN [SEQ. ID. NO.: 5] including a
25 proposed promoter sequence [SEQ. ID. NO.: 27] are provided. Eleven exons [SEQ. ID. NOS. 28-38] and ten introns [SEQ. ID. NOS.: 39-48] are comprised by the MN gene. Also a 1.4 kilobase region [SEQ. ID. NO. 49] within the middle of the MN genomic sequence is described herein, which has the character
30 of a typical CpG-rich island, and which contains multiple putative binding sites for transcription factors AP2 and Sp1.

Also described are antibodies prepared against proteins/polypeptides. MN proteins/ polypeptides can be used in serological assays according to this invention to detect
35 MN-specific antibodies. Further, MN proteins/polypeptides and/or antibodies reactive with MN antigen can be used in immunoassays according to this invention to detect and/or

quantitate MN antigen. Such assays may be diagnostic and/or prognostic for neoplastic/pre-neoplastic disease.

SUMMARY OF THE INVENTION

This invention is directed to the MN gene, fragments thereof and the related cDNA which are useful, for example, as follows: 1) to produce MN proteins/ polypeptides by biochemical engineering; 2) to prepare nucleic acid probes to test for the presence of the MN gene in cells of a subject; 3) to prepare appropriate polymerase chain reaction (PCR) primers for use, for example, in PCR-based assays or to produce nucleic acid probes; 4) to identify MN proteins and polypeptides as well as homologs or near homologs thereto; 5) to identify various mRNAs transcribed from MN genes in various tissues and cell lines, preferably human; and 6) to identify mutations in MN genes. The invention further concerns purified and isolated DNA molecules comprising the MN gene or fragments thereof, or the related cDNA or fragments thereof.

Thus, this invention in one aspect concerns isolated nucleic acid sequences that encode MN proteins or polypeptides wherein the nucleotide sequences for said nucleic acids are selected from the group consisting of:

(a) SEQ. ID. NO.: 1;

(b) nucleotide sequences that hybridize under stringent conditions to SEQ. ID. NO.: 1 or to its complement;

(c) nucleotide sequences that differ from SEQ. ID. NO.: 1 or from the nucleotide sequences of (b) in codon sequence because of the degeneracy of the genetic code. Further, such nucleic acid sequences are selected from nucleotide sequences that but for the degeneracy of the genetic code would hybridize to SEQ. ID. NO.: 1 or to its complement under stringent hybridization conditions.

Further, such isolated nucleic acids that encode MN proteins or polypeptides can also include the MN nucleic acids of the genomic sequence shown in Figure 3a-d, that is, SEQ. ID. NO.: 5, as well as sequences that hybridize to it or its complement under stringent conditions, or would hybridize to SEQ. ID. NO.: 5 or to its complement under such conditions,

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but for the degeneracy of the genetic code. Degenerate variants of SEQ. ID. NOS.: 1 and 5 are within the scope of the invention.

Further, this invention concerns nucleic acid probes which are fragments of the isolated nucleic acids that encode MN proteins or polypeptides as described above. Preferably said nucleic acid probes are comprised of at least 29 nucleotides, more preferably of at least 50 nucleotides, still more preferably at least 100 nucleotides, and even more preferably at least 150 nucleotides.

Still further, this invention is directed to isolated nucleic acids containing at least twenty-seven nucleotides selected from the group consisting of:

(a) SEQ. ID. NOS.: 1, 5 and 27-49 and that are complementary to SEQ. ID. NOS.: 1, 5 and 27-49;

(b) nucleotide sequences that hybridize under standard stringent hybridization conditions to one or more of the following nucleotide sequences: SEQ. ID. NOS.: 1, 5, and 27-49 and the respective complements of SEQ. ID. NOS.: 1, 5 and 27-49; and

(c) nucleotide sequences that differ from the nucleotide sequences of (a) and (b) in codon sequence because of the degeneracy of the genetic code. The invention also concerns nucleic acids that but for the degeneracy of the genetic code would hybridize to the nucleic acids of (a) and (b) under standard stringent hybridization conditions. Further this invention concerns nucleic acids of (b) and (c) that hybridize partially or wholly to the non-coding regions of SEQ. ID. NO.: 5 or its complement as, for example, sequences that function as nucleic acid probes to identify MN nucleic acid sequences. Conventional technology can be used to determine whether the nucleic acids of (b) and (c) or of fragments of SEQ. ID. NO.: 5 are useful to identify MN nucleic acid sequences, for example, as outlined in Benton and Davis, Science, 196: 180 (1977) and Fuscoe et al. Genomics, 5: 100 (1989). In general, such nucleic acids are preferably at least 29 nucleotides, most preferably at least 50 nucleotides and still more preferably at least 100

nucleotides. An exemplary and preferred nucleic acid probe is SEQ. ID. NO.: 55 (a 470 bp probe useful in RNase portection assays).

Test kits of this invention can comprise the nucleic acid probes of the invention which are useful diagnostically/prognostically for neoplastic and/or pre-neoplastic disease. Preferred test kits comprise means for detecting or measuring the hybridization of said probes to the MN gene or to the mRNA product of the MN gene, such as a visualizing means.

Fragments of the isolated nucleic acids of the invention, can also be used as PCR primers to amplify segments of MN genes, and may be useful in identifying mutations in MN genes. Typically, said PCR primers are oligonucleotides, preferably at least 16 nucleotides, but they may be considerably longer. Exemplary primers may be from about 16 nucleotides to about 50 nucleotides, preferably from about 19 nucleotides to about 45 nucleotides.

Further, the invention concerns the use of such PCR primers in methods to detect mutations in an isolated MN gene and/or fragment(s) thereof. For example, such methods can comprise amplifying one or more fragment(s) of an MN gene by PCR, and determining whether any of said one or more fragments contain mutations, by, for example, comparing the size of the amplified fragments to those of similarly amplified corresponding fragments of MN genes known to be normal, by using a PCR-single-strand conformation polymorphism assay or a denaturing gradient gel electrophoretic assay.

This invention also concerns nucleic acids which encode MN proteins or polypeptides that are specifically bound by monoclonal antibodies designated M75 that are produced by the hybridoma VU-M75 deposited at the American Type Culture Collection (ATCC) at 12301 Parklawn Drive in Rockville, Maryland 20852 (USA) under ATCC No. HB 11128, and/or by monoclonal antibodies designated MN12 produced by the hybridoma MN 12.2.2 deposited at the ATCC under ATCC No. HB 11647.

This invention further concerns isolated nucleic acids containing at least sixteen nucleotides, preferably at least twenty-nine nucleotides, more preferably at least fifty nucleotides, wherein said nucleic acid is selected from the group consisting of:

- (a) the MN nucleic acids contained in plasmids A4a, XE1 and XE3 which were deposited at the American Type Culture Collection (ATCC) in Rockville, Maryland in the United States of America under the respective ATCC Nos. 97199, 97200, and 97198;
- (b) nucleic acids that hybridize under stringent conditions to the MN nucleic acids of (a); and
- (c) nucleic acids that differ from the nucleic acids of (a) or (b) in codon sequence due to the degeneracy of the genetic code. Such isolated nucleic acids, for example, can be polymerase chain reaction (PCR) primers.

The invention further concerns isolated nucleic acids that code for an MN protein, MN fusion protein or MN polypeptide that is operatively linked to an expression control sequence within a vector; unicellular hosts, prokaryotic or eukaryotic, that are transformed or transfected therewith; and methods of recombinantly producing MN proteins, MN fusion proteins and MN polypeptides comprising transforming or transfecting unicellular hosts with said nucleic acid operatively linked to an expression control sequence, culturing said transformed or transfected unicellular hosts so that said MN proteins, fusion proteins or polypeptides are expressed, and extracting and isolating said MN protein fusion protein or polypeptide.

Recombinant nucleic acids that encode MN fusion proteins are claimed as consisting essentially of an MN protein or MN polypeptide and a non-MN protein or polypeptide wherein the nucleotide sequence for the portion of the nucleic acid encoding the MN protein or polypeptide is selected from the group consisting of:

- (a) SEQ. ID. NO.: 1;

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(b) nucleotide sequences that hybridize under stringent conditions to SEQ. ID. NO.: 1 or to its complement; and

5 (c) degenerate variants of SEQ. ID. NO.: 1, and of the nucleotide sequences of (b);

wherein the nucleic acid encoding said MN protein or polypeptide contains at least twenty-nine nucleotides.

Said non-MN protein or polypeptide may preferably be nonimmunogenic to humans and not typically reactive to
10 antibodies in human body fluids. Examples of such a DNA sequence is the alpha-peptide coding region of beta-galactosidase and a sequence coding for glutathione S-transferase or a fragment thereof. However, in some instances, a non-MN protein or polypeptide that is
15 serologically active, immunogenic and/or antigenic may be preferred as a fusion partner to a MN antigen. Further, claimed herein are such recombinant fusion proteins/polypeptides which are substantially pure and non-naturally occurring. Exemplary fusion proteins of this invention are
20 GEX-3X-MN, MN-Fc and MN-PA, described infra.

In HeLa and in tumorigenic HeLa x fibroblast hybrid (H/F-T) cells, MN protein is manifested as a "twin" protein p54/58N; it is glycosylated and forms disulfide-linked oligomers. As determined by electrophoresis upon reducing
25 gels, MN proteins have molecular weights in the range of from about 40 kd to about 70 kd, preferably from about 45 kd to about 65 kd, more preferably from about 48 kd to about 58 kd. Upon non-reducing gels, MN proteins in the form of oligomers have molecular weights in the range of from about 145 kd to
30 about 160 kd, preferably from about 150 to about 155 kd, still more preferably from about 152 to about 154 kd. A predicted amino acid sequence for a preferred MN protein of this invention is shown in Figure 1 [SEQ. ID. NO. 2].

Other particular MN proteins or polypeptides are
35 exemplified by the putative MN signal peptide shown as the first thirty-seven amino acids in Figure 1 [SEQ. ID. NO.: 6], preferred MN antigen epitopes [SEQ. ID. NOS.: 10-16], and domains of the MN protein represented in Figure 1 as amino

acids 38-135 [SEQ. IS. NO.: 50], 136-391 [SEQ. ID. NO.: 51],
414-433 [SEQ. ID. NO.: 52], and 434-459 [SEQ. ID. NO.: 53].

The discovery of the MN gene and protein and thus,
of substantially complementary MN genes and proteins encoded
5 thereby, led to the finding that the expression of MN proteins
was associated with tumorigenicity. That finding resulted in
the creation of methods that are diagnostic/ prognostic for
cancer and precancerous conditions. Methods and compositions
are provided for identifying the onset and presence of
10 neoplastic disease by detecting and/or quantitating MN antigen
in patient samples, including tissue sections and smears, cell
and tissue extracts from vertebrates, preferably mammals and
more preferably humans. Such MN antigen may also be found in
body fluids.

15 MN proteins and genes are of use in research
concerning the molecular mechanisms of oncogenesis, in cancer
diagnostics/prognostics, and may be of use in cancer
immunotherapy. The present invention is useful for detecting
a wide variety of neoplastic and/or pre-neoplastic diseases.
20 Exemplary neoplastic diseases include carcinomas, such as
mammary, bladder, ovarian, uterine, cervical, endometrial,
squamous cell and adenosquamous carcinomas; and head and neck
cancers; mesodermal tumors, such as neuroblastomas and
retinoblastomas; sarcomas, such as osteosarcomas and Ewing's
25 sarcoma; and melanomas. Of particular interest are head and
neck cancers, gynecologic cancers including ovarian, cervical,
vaginal, endometrial and vulval cancers; gastrointestinal
cancer, such as, stomach, colon and esophageal cancers;
urinary tract cancer, such as, bladder and kidney cancers;
30 skin cancer; liver cancer; prostate cancer; lung cancer; and
breast cancer. Of still further particular interest are
gynecologic cancers; breast cancer; urinary tract cancers,
especially bladder cancer; lung cancer; and liver cancer.
Even further of particular interest are gynecologic cancers
35 and breast cancer. Gynecologic cancers of particular interest
are carcinomas of the uterine cervix, endometrium and ovaries;
more particularly such gynecologic cancers include cervical
squamous cell carcinomas, adenosquamous carcinomas,

adenocarcinomas as well as gynecologic precancerous conditions, such as metaplastic cervical tissues and condylomas.

The invention further relates to the biochemical engineering of the MN gene, fragments thereof or related cDNA. For example, said gene or a fragment thereof or related cDNA can be inserted into a suitable expression vector, wherein it is operatively linked to an expression control sequence; host cells, preferably unicellular, can be transformed or transfected with such an expression vector; and an MN protein/polypeptide, preferably an MN protein, is expressed therein. Such a recombinant protein or polypeptide can be glycosylated or nonglycosylated, preferably glycosylated, and can be purified to substantial purity. The invention further concerns MN proteins/polypeptides which are synthetically or otherwise biologically prepared.

Said MN proteins/polypeptides can be used in assays to detect MN antigen in patient samples and in serological assays to test for MN-specific antibodies. MN proteins/polypeptides of this invention are serologically active, immunogenic and/or antigenic. They can further be used as immunogens to produce MN-specific antibodies, polyclonal and/or monoclonal, as well as an immune T-cell response.

The invention further is directed to MN-specific antibodies, which can be used diagnostically/prognostically and may be used therapeutically. Preferred according to this invention are MN-specific antibodies reactive with the epitopes represented respectively by the amino acid sequences of the MN protein shown in Figure 1 as follows: from AA 62 to AA 67 [SEQ. ID. NO.: 10]; from AA 55 to AA 60 [SEQ. ID. NO.: 11]; from AA 127 to AA 147 [SEQ. ID. NO.: 12]; from AA 36 to AA 51 [SEQ. ID. NO.: 13]; from AA 68 to AA 91 [SEQ. ID. NO.: 14]; from AA 279 to AA 291 [SEQ. ID. NO.: 15]; and from AA 435 to AA 450 [SEQ. ID. NO.: 16]. More preferred are antibodies reactive with epitopes represented by SEQ. ID. NOS.: 10, 11 and 12. Still more preferred are antibodies reactive with the epitopes represented by SEQ. ID NOS: 10 and

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11, as for example, respectively Mabs M75 and MN12. Most preferred are monoclonal antibodies reactive with the epitope represented by SEQ. ID. NO.: 10.

Also preferred according to this invention are
5 antibodies prepared against recombinantly produced MN proteins as, for example, GEX-3X-MN, MN 20-19, MN-Fc and MN-PA. Also preferred are MN-specific antibodies prepared against glycosylated MN proteins, such as, MN 20-19 expressed in baculovirus infected Sf9 cells.

10 A hybridoma that produces a representative MN-specific antibody, the monoclonal antibody M75 (Mab M75), was deposited at the under ATCC Number HB 11128 as indicated above. The M75 antibody was used to discover and identify the MN protein and can be used to identify readily MN antigen in
15 Western blots, in radioimmunoassays and immunohistochemically, for example, in tissue samples that are fresh, frozen, or formalin-, alcohol-, acetone- or otherwise fixed and/or paraffin-embedded and deparaffinized. Another representative MN-specific antibody, Mab MN12, is secreted by the hybridoma
20 MN 12.2.2, which was deposited at the ATCC under the designation HB 11647.

MN-specific antibodies can be used, for example, in laboratory diagnostics, using immunofluorescence microscopy or immunohistochemical staining; as a component in immunoassays
25 for detecting and/or quantitating MN antigen in, for example, clinical samples; as probes for immunoblotting to detect MN antigen; in immunoelectron microscopy with colloid gold beads for localization of MN proteins and/or polypeptides in cells; and in genetic engineering for cloning the MN gene or
30 fragments thereof, or related cDNA. Such MN-specific antibodies can be used as components of diagnostic/prognostic kits, for example, for in vitro use on histological sections; such antibodies can also and used for in vivo diagnostics/prognostics, for example, such antibodies can be labeled
35 appropriately, as with a suitable radioactive isotope, and used in vivo to locate metastases by scintigraphy. Further such antibodies may be used in vivo therapeutically to treat cancer patients with or without toxic and/or cytostatic agents

attached thereto. Further, such antibodies can be used in vivo to detect the presence of neoplastic and/or pre-neoplastic disease. Still further, such antibodies can be used to affinity purify MN proteins and polypeptides.

5 This invention also concerns methods of treating neoplastic disease and/or pre-neoplastic disease comprising inhibiting the expression of MN genes by administering antisense nucleic acid sequences that are substantially complementary to mRNA transcribed from MN genes. Said
10 antisense nucleic acid sequences are those that hybridize to such mRNA under stringent hybridization conditions. Preferred are antisense nucleic acid sequences that are substantially complementary to sequences at the 5' end of the MN cDNA sequence shown in Figure 1. Preferably said antisense nucleic
15 acid sequences are oligonucleotides.

This invention also concerns vaccines comprising an immunogenic amount of one or more substantially pure MN proteins and/or polypeptides dispersed in a physiologically acceptable, nontoxic vehicle, which amount is effective to
20 immunize a vertebrate, preferably a mammal, more preferably a human, against a neoplastic disease associated with the expression of MN proteins. Said proteins can be recombinantly, synthetically or otherwise biologically produced. A particular use of said vaccine would be to
25 prevent recidivism and/or metastasis. For example, it could be administered to a patient who has had an MN-carrying tumor surgically removed, to prevent recurrence of the tumor.

The immunoassays of this invention can be embodied in test kits which comprise MN proteins/polypeptides and/or
30 MN-specific antibodies. Such test kits can be in solid phase formats, but are not limited thereto, and can also be in liquid phase format, and can be based on immunohistochemical assays, ELISAS, particle assays, radiometric or fluorometric assays either unamplified or amplified, using, for example,
35 avidin/biotin technology.

Abbreviations

The following abbreviations are used herein:

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	AA	-	amino acid
	ATCC	-	American Type Culture Collection
	bp	-	base pairs
	BLV	-	bovine leukemia virus
5	BSA	-	bovine serum albumin
	BRL	-	Bethesda Research Laboratories
	CA	-	carbonic anhydrase
	CAT	-	chloramphenicol acetyltransferase
	Ci	-	curie
10	cm	-	centimeter
	CMV	-	cytomegalovirus
	cpm	-	counts per minute
	C-terminus	-	carboxyl-terminus
	°C	-	degrees centigrade
15	DEAE	-	diethylaminoethyl
	DMEM	-	Dulbecco modified Eagle medium
	EDTA	-	ethylenediaminetetraacetate
	EIA	-	enzyme immunoassay
	ELISA	-	enzyme-linked immunosorbent assay
20	F	-	fibroblasts
	FCS	-	fetal calf serum
	FITC	-	fluorescein isothiocyanate
	GEX-3X-MN	-	fusion protein MN glutathione S-transferase
	H	-	HeLa cells
25	HEF	-	human embryo fibroblasts
	HeLa K	-	standard type of HeLa cells
	HeLa S	-	Stanbridge's mutant HeLa D98/AH.2
	H/F-T	-	hybrid HeLa fibroblast cells that are tumorigenic; derived from HeLa D98/AH.2
30	H/F-N	-	hybrid HeLa fibroblast cells that are nontumorigenic; derived from HeLa D98/AH.2
	HRP	-	horseradish peroxidase
	Inr	-	initiator
	IPTG	-	isopropyl-Beta-D-thiogalacto-pyranoside
35	kb	-	kilobase
	kbp	-	kilobase pairs
	kd	-	kilodaltons
	LCMV	-	lymphocytic choriomeningitis virus

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	LTR	-	long terminal repeat
	M	-	molar
	mA	-	milliampere
	MAB	-	monoclonal antibody
5	ME	-	mercaptoethanol
	MEM	-	minimal essential medium
	min.	-	minute(s)
	mg	-	milligram
	ml	-	milliliter
10	mM	-	millimolar
	MMC	-	mitomycin C
	MLV	-	murine leukemia virus
	N	-	normal concentration
	NEG	-	negative
15	ng	-	nanogram
	nt	-	nucleotide
	N-terminus	-	amino-terminus
	ODN	-	oligodeoxynucleotide
	ORF	-	open reading frame
20	PA	-	Protein A
	PBS	-	phosphate buffered saline
	PCR	-	polymerase chain reaction
	PEST	-	combination of one-letter abbreviations for proline, glutamic acid, serine, threonine
25	pI	-	isoelectric point
	PMA	-	phorbol 12-myristate 13-acetate
	POS	-	positive
	Py	-	pyrimidine
	RIA	-	radioimmunoassay
30	RIP	-	radioimmunoprecipitation
	RIPA	-	radioimmunoprecipitation assay
	RNP	-	RNase protection assay
	SDRE	-	serum dose response element
	SDS	-	sodium dodecyl sulfate
35	SDS-PAGE	-	sodium dodecyl sulfate-polyacrylamide gel electrophoresis
	SINE	-	short interspersed repeated sequence
	SSDS	-	synthetic splice donor site

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	SP-RIA	-	solid-phase radioimmunoassay
	SSDS	-	synthetic splice donor sit
	SSPE	-	NaCl (0.18 M), sodium phosphate (0.01 M), EDTA (0.001 M)
5	TBE	-	Tris-borate/EDTA electrophoresis buffer
	TCA	-	trichloroacetic acid
	TC media	-	tissue culture media
	TMB	-	tetramethylbenzidine
	Tris	-	tris (hydroxymethyl) aminomethane
10	μCi	-	microcurie
	μg	-	microgram
	μl	-	microliter
	μM	-	micromolar
	VSV	-	vesicular stomatitis virus
15	X-MLV	-	xenotropic murine leukemia virus

Cell Lines

	HeLa K	--	standard type of HeLa cells; aneuploid, epithelial-like cell line isolated from a human cervical adenocarcinoma [Gey et al., <u>Cancer Res.</u> , 12: 264 (1952); Jones et al., <u>Obstet. Gynecol.</u> , 38: 945-949 (1971)] obtained from Professor B. Korych, [Institute of Medical Microbiology and Immunology, Charles University; Prague, Czech Republic]
20			
25	HeLa D98/AH.2 (also HeLa S)	--	Mutant HeLa clone that is hypoxanthine guanine phosphoribosyl transferase-deficient (HGPRT ⁻) kindly provided by Eric J. Stanbridge [Department of Microbiology, College of Medicine, University of California, Irvine, CA (USA)] and reported in Stanbridge et al., <u>Science</u> , 215: 252-259 (15 Jan. 1982); parent of hybrid cells H/F-N and H/F-T, also obtained from E.J. Stanbridge.
30			
	NIH-3T3	--	murine fibroblast cell line reported in Aaronson, <u>Science</u> , 237: 178 (1987).
35			

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- XC -- cells derived from a rat rhabdomyosarcoma induced with Rous sarcoma virus-induced rat sarcoma [Svoboda, J., Natl. Cancer Center Institute Monograph No. 17, IN:
- 5 "International Conference on Avian Tumor Viruses" (J.W. Beard ed.), pp. 277-298 (1964)], kindly provided by Jan Svoboda [Institute of Molecular Genetics, Czechoslovak Academy of Sciences; Prague, Czech Republic]; and
- 10
- CGL1 -- H/F-N hybrid cells (HeLa D98/AH.2 derivative)
- CGL2 -- H/F-N hybrid cells (HeLa D98/AH.2 derivative)
- CGL3 -- H/F-T hybrid cells (HeLa D98/AH.2 derivative)
- CGL4 -- H/F-T hybrid cells (HeLa D98/Ah.2 derivative)

15 Nucleotide and Amino Acid Sequence Symbols

The following symbols are used to represent nucleotides herein:

	<u>Base Symbol</u>	<u>Meaning</u>
20	A	adenine
	C	cytosine
	G	guanine
	T	thymine
	U	uracil
25	I	inosine
	M	A or C
	R	A or G
	W	A or T/U
	S	C or G
30	Y	C or T/U
	K	G or T/U
	V	A or C or G
	H	A or C or T/U

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D	A or G or T/U
B	C or G or T/U
N/X	A or C or G or T/U

There are twenty main amino acids, each of which is
 5 specified by a different arrangement of three adjacent
 nucleotides (triplet code or codon), and which are linked
 together in a specific order to form a characteristic protein.
 A three-letter or one-letter convention is used herein to
 identify said amino acids, as, for example, in Figure 1 as
 10 follows:

	<u>Amino acid name</u>	<u>3 Ltr. Abbrev.</u>	<u>1 Ltr. Abbrev.</u>
	Alanine	Ala	A
	Arginine	Arg	R
15	Asparagine	Asn	N
	Aspartic Acid	Asp	D
	Cysteine	Cys	C
	Glutamic Acid	Glu	E
	Glutamine	Gln	Q
20	Glycine	Gly	G
	Histidine	His	H
	Isoleucine	Ile	I
	Leucine	Leu	L
	Lysine	Lys	K
25	Methionine	Met	M
	Phenylalanine	Phe	F
	Proline	Pro	P
	Serine	Ser	S
	Threonine	Thr	T
30	Tryptophan	Trp	W
	Tyrosine	Tyr	Y
	Valine	Val	V
	Unknown or other		X

BRIEF DESCRIPTION OF THE FIGURES

35 Figure 1 provides the nucleotide sequence for a
 full-length MN cDNA [SEQ. ID. NO.: 1] clone isolated as

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described herein. Figur 1 also sets forth the predicted amino acid sequence [SEQ. ID. NO.: 2] encoded by the cDNA.

Figure 2 compares the results of immunizing baby rats to XC tumor cells with rat serum prepared against the fusion protein MN glutathione S-transferase (GEX-3X-MN) (the IM group) with the results of immunizing baby rats with control rat sera (the C group). Each point on the graph represents the tumor weight of a tumor from one rat. Example 2 details those experiments.

Figure 3a-d provides a 10,898 bp complete genomic sequence of MN [SEQ. ID. NO.: 5]. The base count is as follows: 2654 A; 2739 C; 2645 G; and 2859 T. The 11 exons are shown in capital letters.

Figure 4 is a restriction map of the full-length MN cDNA. The open reading frame is shown as an open box. The thick lines below the restriction map illustrate the sizes and positions of two overlapping cDNA clones. The horizontal arrows indicate the positions of primers R1 [SEQ. ID. NO.: 7] and R2 [SEQ. ID. NO.: 8] used for the 5' end RACE. Relevant restriction sites are BamHI (B), EcoRV (V), EcoRI (E), PstI (Ps), PvuII (Pv).

Figure 5 is a map of the human MN gene. The numbered cross-hatched boxes represent exons. The box designated LTR denotes a region of homology to HERV-K LTR. The empty boxes are Alu-related sequences.

Figure 6 is a nucleotide sequence for the proposed promoter of the human MN gene [SEQ. ID. No.: 27]. The nucleotides are numbered from the transcription initiation site according to RNase protection assay. Potential regulatory elements are overlined. Transcription start sites are indicated by asterisks (RNase protection) and dots (RACE). The sequence of the 1st exon begins under the asterisks.

Figure 7 provides a schematic of the alignment of MN genomic clones according to their position related to the transcription initiation site. All the genomic fragments except Bd3 were isolated from a lambda FIX III genomic library derived from HeLa cells. Clone Bd3 was derived from a human fetal brain library.

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Figure 8 shows the construction and cloning of a series of 5' deletion mutants of MN's putative promoter region linked to the bacterial CAT gene.

DETAILED DESCRIPTION

5 The MN gene is shown herein to be organized into 11 exons and 10 introns. Described herein is the cloning and sequencing of the MN cDNA and genomic sequences, and the genetic engineering of MN proteins -- such as the GEX-3X-MN, MN-PA, MN-Fc and MN 20-19 proteins. The recombinant MN
10 proteins can be conveniently purified by affinity chromatography.

 MN is manifested in HeLa cells by a twin protein, p54/58N. Immunoblots using a monoclonal antibody reactive with p54/58N (Mab M75) revealed two bands at 54 kd and 58 kd.
15 Those two bands may correspond to one type of protein that differs by glycosylation pattern or by how it is processed. Herein, the phrase "twin protein" indicates p54/58N.

 The expression of MN proteins appears to be diagnostic/prognostic for neoplastic disease. The MN twin
20 protein, p54/58N, was found to be expressed in HeLa cells and in Stanbridge's tumorigenic (H/F-T) hybrid cells [Stanbridge et al., Somatic Cell Genet. 7: 699-712 (1981); and Stanbridge et al., Science, 215: 252-259 (1982)] but not in fibroblasts or in non-tumorigenic (H/F-N) hybrid cells [Stanbridge et al.,
25 id.]. In early studies reported in Zavada et al. WO 93/18152, supra, MN proteins were found in immunoblots prepared from human ovarian, endometrial and uterine cervical carcinomas, and in some benign neoplasias (as mammary papilloma) but not from normal ovarian, endometrial, uterine or placental
30 tissues. Example 1 herein details further research on MN gene expression wherein MN antigen, as detected by immunohistochemical staining, was found to be prevalent in tumor cells of a number of cancers, including cervical, bladder, head and neck, and renal cell carcinomas among
35 others. Further, the immunohistochemical staining experiments of Example 1 show that among normal tissues tested, only normal stomach tissues showed routinely and extensively the

presence of MN antigen. MN antigen is further shown herein to be present sometimes in morphologically normal-appearing areas of tissue specimens exhibiting dysplasia and/or malignancy.

MN Gene--Cloning and Sequencing

5 Figure 1 provides the nucleotide sequence for a full-length MN cDNA clone isolated as described below [SEQ. ID. NO.: 1]. Figure 3a-d provides a complete MN genomic sequence [SEQ. ID. NO.: 5]. Figure 6 shows the nucleotide sequence for a proposed MN promoter [SEQ. ID. NO.: 27].

10 It is understood that because of the degeneracy of the genetic code, that is, that more than one codon will code for one amino acid [for example, the codons TTA, TTG, CTT, CTC, CTA and CTG each code for the amino acid leucine (leu)], that variations of the nucleotide sequences in, for example,
15 SEQ. ID. NOS.: 1 and 5 wherein one codon is substituted for another, would produce a substantially equivalent protein or polypeptide according to this invention. All such variations in the nucleotide sequences of the MN cDNA and complementary nucleic acid sequences are included within the scope of this
20 invention.

It is further understood that the nucleotide sequences herein described and shown in Figures 1, 3a-d and 6, represent only the precise structures of the cDNA, genomic and promoter nucleotide sequences isolated and described herein.

25 It is expected that slightly modified nucleotide sequences will be found or can be modified by techniques known in the art to code for substantially similar or homologous MN proteins and polypeptides, for example, those having similar epitopes, and such nucleotide sequences and proteins/
30 polypeptides are considered to be equivalents for the purpose of this invention. DNA or RNA having equivalent codons is considered within the scope of the invention, as are synthetic nucleic acid sequences that encode proteins/polypeptides homologous or substantially homologous to MN
35 proteins/polypeptides, as well as those nucleic acid sequences that would hybridize to said exemplary sequences [SEQ. ID. NOS. 1, 5 and 27] under stringent conditions, or that, but for

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the degeneracy of the genetic code would hybridize to said cDNA nucleotide sequences under stringent hybridization conditions. Modifications and variations of nucleic acid sequences as indicated herein are considered to result in
5 sequences that are substantially the same as the exemplary MN sequences and fragments thereof.

Partial cDNA clone

In Zavada et al., *id.*, the isolation of a partial MN cDNA clone of 1397 bp in length was described. A lambda gt11
10 cDNA library of LMCV-infected HeLa cells was prepared and subjected to immunoscreening with Mab M75 in combination with goat anti-mouse antibodies conjugated with alkaline phosphatase. One positive clone was picked and subcloned into the NotI site of pBluescript KS [Stratagen; La Jolla, CA
15 (USA)] thereby creating pBluscript-MN.

Two oppositely oriented nested deletions were made using Erase-a-Base™ kit [Promega; Madison, WI (USA)] and sequenced by dideoxy method with a T7 sequencing kit [Pharmacia; Piscataway, NJ (USA)]. The sequencing showed a
20 partial cDNA clone, the insert being 1397 bp long. The sequence comprises a large 1290 bp open reading frame and 107 bp 3' untranslated region containing a polyadenylation signal (AATAAA). However, the sequence surrounding the first ATG codon in the open reading frame (ORF) did not fit the
25 definition of a translational start site. In addition, as followed from a comparison of the size of the MN clone with that of the corresponding mRNA in a Northern blot, the cDNA was shown to be missing about 100 bp from the 5' end of its sequence.

Full-Length cDNA Clone

Attempts to isolate a full-length clone from the original cDNA library failed. Therefore, the inventors performed a rapid amplification of cDNA ends (RACE) using MN-specific primers, R1 and R2 [SEQ. ID. NOS.: 7 and 8], derived
35 from the 5' region of the original cDNA clone. The RACE product was inserted into pBluescript, and the entire

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population of recombinant plasmids was sequenced with an MN-specific primer ODN1 [SEQ. ID. NO.: 3]. In that way, a reliable sequence at the very 5' end of the MN cDNA as shown in Figure 1 [SEQ. ID. NO.: 1] was obtained.

5 Specifically, RACE was performed using 5' RACE System [GIBCO BRL; Gaithersburg, MD (USA)] as follows. 1 µg of mRNA (the same as above) was used as a template for the first strand cDNA synthesis which was primed by the MN-specific antisense oligonucleotide, R1 (5'-
10 TGGGGTCTTGAGGATCTCCAGGAG-3') [SEQ. ID. NO.: 7]. The first strand product was precipitated twice in the presence of ammonium acetate and a homopolymeric C tail was attached to its 3' end by TdT. Tailed cDNA was then amplified by PCR using a nested primer, R2 (5'-CTCTAACTTCAGGGAGCCCTCTTCTT-3')
15 [SEQ. ID. NO.: 8] and an anchor primer that anneals to the homopolymeric tail (5'-CUACUACUACUAGGCCACGCGTCGACTAGTACGGGI
IGGGIIGGGIIG-3') [SEQ. ID. NO.: 9]. The amplified product was digested with BamHI and SalI restriction enzymes and cloned into pBluescript II KS plasmid. After transformation,
20 plasmid DNA was purified from the whole population of transformed cells and used as a template for sequencing with the MN-specific primer ODN1 [SEQ. ID. NO.: 3; a 29-mer 5' CGCCAGTGGGTCATCTTCCCCAGAAGAG 3'].

Based upon results of the RACE analysis, the full-
25 length MN cDNA sequence was seen to contain a single ORF starting at position 12, with an ATG codon that is in a good context (GCGCATGG) with the rule proposed for translation initiation [Kozak, J. Cell. Biol., 108: 229-241 (1989)].
[See below under Mapping of MN Gene Transcription Initiation
30 Site for fine mapping of the 5' end of the MN gene.] The AT rich 3' untranslated region contains a polyadenylation signal (AATAAA) preceding the end of the cDNA by 10 bp. Surprisingly, the sequence from the original clone as well as from four additional clones obtained from the same cDNA
35 library did not reveal any poly(A) tail. Moreover, just downstream of the poly(A) signal, an ATTTA motif that is thought to contribute to mRNA instability [Shaw and Kamen, Cell, 46: 659-667 (1986)] was found. That fact raised the

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possibility that the poly (A) tail is missing due to the specific degradation of the MN mRNA.

Genomic clones

To study MN regulation, MN genomic clones were isolated. One MN genomic clone (Bd3) was isolated from a human cosmid library prepared from fetal brain using both MN cDNA as a probe and the MN-specific primers derived from the 5' end of the cDNA ODN1 [SEQ. ID. NO.: 3, supra] and ODN2 [SEQ. ID NO.: 4; 19-mer (5' GGAATCCTCCTGCATCCGG 3')]. Sequence analysis revealed that that genomic clone covered a region upstream from a MN transcription start site and ending with the BamHI restriction site localized inside the MN cDNA. Other MN genomic clones can be similarly isolated.

In order to identify the complete genomic region of MN, the human genomic library in Lambda FIX II vector (Stratagene) was prepared from HeLa chromosomal DNA and screened by plaque hybridization using MN cDNA as described below. Several independent MN recombinant phages were identified, isolated and characterized by restriction mapping and hybridization analyses. Four overlapping recombinants covering the whole genomic region of MN were selected, digested and subcloned into pBluescript. The subclones were then subjected to bidirectional nested deletions and sequencing. DNA sequences were compiled and analyzed by computer using the DNASIS software package.

The details of isolating genomic clones covering the complete genomic region for MN are provided below. Figure 7 provides a schematic of the alignment of MN genomic clones according to the transcription initiation site. Plasmids containing the A4a clone and the XE1 and XE3 subclones were deposited at the American Type Culture Collection (ATCC) at 12301 Parklawn Drive, Rockville, MD 20852 (USA) on June 6, 1995, respectively under ATCC Deposit Nos. 97199, 97200, and 97198.

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Isolation of Genomic DNA Clones

The Sau3AI human HeLa genomic library was prepared in Lambda FIX II vector [Stratagene; La Jolla, CA (USA)] according to manufacturer's protocol. Human fetal brain cosmid library in SuperCos cosmid was from Stratagene. Recombinant phages or bacteria were plated at 1×10^5 plaque forming units on 22x22 cm Nunc plates or 5×10^4 cells on 150 mm Petri dishes, and plaques or colonies were transferred to Hybond N membranes (Amersham). Hybridization was carried out with the full-length MN cDNA labeled with [P^{32}]PdCTP by the Multiprime DNA labeling method (Amersham) at 65°C in 6 x SSC, 0.5% SDS, 10 x Denhardt's and 0.2 mg/1 ml salmon sperm DNA. Filters were washed twice in 2 x SSC, 0.1% SDS at 65°C for 20 min. The dried filters were exposed to X-ray films, and positive clones were picked up. Phages and bacteria were isolated by 3-4 sequential rounds of screening.

Subcloning and DNA Sequencing

Genomic DNA fragments were subcloned into a pBluescript KS and templates for sequencing were generated by serial nested deletions using the Erase-a-Base system. Sequencing was performed by the dideoxynucleotide chain termination method using T7 sequencing kit (Pharmacia). Nucleotide sequence alignments and analyses were carried out using the DNASIS software package (Hitachi Software Engineering).

Exon-Intron Structure of Complete MN Genomic Region

The complete sequence of the overlapping clones contains 10,898 bp (SEQ. ID. NO.: 5). Figure 5 depicts the organization of the human MN gene, showing the location of all 11 exons as well as the 2 upstream and 6 intronic Alu repeat elements. All the exons are small, ranging from 27 to 191 bp, with the exception of the first exon which is 415 bp. The intron sizes range from 89 to 1400 bp.

Table 1 below lists the splice donor and acceptor sequences that conform to consensus splice sequences including

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the AG-GT motif [Mount, "A catalogue of splice junction sequences," Nucleic Acids Res. 10: 459-472 (1982)].

TABLE 1
Exon-Intron Structure of the Human MN Gene

5	Exon	Size	Genomic Position**	SEQ ID NO	5'splice donor	SEQ ID No
	1	415	*3537-3951	28	AGAAG gtaagt	67
	2	30	5126-5155	29	TGGAG gtgaga	68
	3	171	5349-5519	30	CAGTC gtgagg	69
	4	143	5651-5793	31	CCGAG gtgagc	70
	5	93	5883-5975	32	TGGAG gtacca	71
10	6	67	7376-7442	33	GGAAG gtcagt	72
	7	158	8777-8934	34	AGCAG gtgggc	73
	8	145	9447-9591	35	GCCAG gtacag	74
	9	27	9706-9732	36	TGCTG gtgagt	75
	10	82	10350-10431	37	CACAG gtatta	76
	11	191	10562-10752	38	ATAAT end	

	Intron	Size	Genomic Position**	SEQ ID NO	3'splice acceptor	SEQ ID NO
	1	1174	3952-5125	39	atacag GGGAT	77
	2	193	5156-5348	40	ccccag GCGAC	78
20	3	131	5520-5650	41	acgcag TGCAA	79
	4	89	5794-5882	42	tttcag ATCCA	80
	5	1400	5976-7375	43	ccccag GAGGG	81
	6	1334	7443-8776	44	tcacag GCTCA	82
	7	512	8935-9446	45	ccctag CTCCA	83
	8	114	9592-9705	46	ctccag TCCAG	84
25	9	617	9733-10349	47	tcgcag GTGACA	85
	10	130	10432-10561	48	acacag AAGGG	86

** positions are related to nt numbering in whole genomic sequence including the 5' flanking region [Figure 3a-d]

* number corresponds to transcription initiation site determined below by RNase protection assay

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A search for sequences related to MN gene in the EMBL Data Library did not reveal any specific homology except for 6 complete and 2 partial Alu-type repeats with homology to Alu sequences ranging from 69.8% to 91% [Jurka and

- 5 Milosavljevic, "Reconstruction and analysis of human Alu genes," J. Mol. Evol. 32: 105-121 (1991)]. Below under the Characterization of the 5' Flanking Region, also a 222 bp sequence proximal to the 5' end of the genomic region is shown to be closely homologous to a region of the HERV-K LTR.

10 Mapping of MN Gene Transcription Initiation Site

- In the earlier attempt to localize the site of transcription initiation of the MN gene by RACE (above), the obtained a major PCR fragment whose sequence placed the start site 12 bp upstream from the first codon of the ORF. That
15 result was obtained probably due to a preferential amplification of the shortest form of mRNA. Therefore, the inventors used an RNase protection assay (RNP) for fine mapping of the 5' end of the MN gene. The probe was a uniformly labeled 470 nucleotide copy RNA (nt -205 to +265)
20 [SEQ. ID. NO.: 55], which was hybridized to total RNA from MN-expressing HeLa and CGL3 cells and analyzed on a sequencing gel. That analysis has shown that the MN gene transcription initiates at multiple sites, the 5' end of the longest MN transcript being 30 nt longer than that previously
25 characterized by RACE.

RNase Protection Assay

- ³²P-labeled RNA probes were prepared with an RNA Transcription kit (Stratagene). In vitro transcription reactions were carried out using 1 µg of the linearized
30 plasmid as a template, 50 µCi of [³²P]rUTP (800 Ci/mmol), 10 U of either T3 or T7 RNA polymerase and other components of the Transcription Kit following instructions of the supplier. For mapping of the 5' end of MN mRNA, the 470 bp NcoI-BamHI fragment (NcoI filled in by Klenow enzyme) of Bd3 clone (nt -
35 205 to +265 related to transcription start) was subcloned to EcoRV-BamHI sites of pBluescript SK+, linearized with HindIII

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and labeled with T3 RNA polymerase. For the 3' end mRNA analysis, probe, that was prepared using T7 RNA polymerase on KS-dXE3-16 template (one of the nested deletion clones of MN genomic region XE3 subclone) digested with Sau3AI (which cuts
5 exon 11 at position 10,629), was used. Approximately 3×10^5 cpm of RNA probe were used per one RNase protection assay reaction.

RNase protection assays (RNP) were performed using Lysate RNase Protection Kit (USB/Amersham) according to
10 protocols of the supplier. Briefly, cells were lysed using Lysis Solution at concentration of approximately 10^7 cells/ml, and 45 μ l of the cell homogenate were used in RNA/RNA hybridization reactions with 32 P-labeled RNA probes prepared as described above. Following overnight hybridizations at 42°C,
15 homogenates were treated for 30 min at 37°C with RNase cocktail mix. Protected RNA duplexes were run on polyacrylamide/urea denaturing sequencing gels. Fixed and dried gels were exposed to X-ray film for 24 - 72 hours.

Mapping of MN Gene Transcription Termination Site

20 An RNase protection assay, as described above, was also used to verify also the 3' end of the MN cDNA. That was important with respect to our previous finding that the cDNA contains a poly(A) signal but lacks a poly(A) tail, which could be lost during the proposed degradation of MN mRNA due
25 to the presence of an instability motif in its 3' untranslated region. RNP analysis of MN mRNA with the fragment of the genomic clone XE3 covering the region of interest corroborated our data from MN cDNA sequencing, since the 3' end of the protected fragment corresponded to the last base of MN cDNA
30 (position 10,752 of the genomic sequence). That site also meets the requirement for the presence of a second signal in the genomic sequence that is needed for transcription termination and polyadenylation [McLauchlan et al., Nucleic Acids Res., 13: 1347 (1985)]. Motif TGTGTTAGT (nt 10,759-
35 10,767) corresponds well to both the consensus sequence and the position of that signal within 22 bp downstream from the polyA signal (nt 10,737-10,742).

Characterization of the 5' Flanking Region

The Bd3 genomic clone isolated from human fetal brain cosmid library was found to cover a region of 3.5 kb upstream from the transcription start site of the MN gene. It contains no significant coding region. Two Alu repeats are situated at positions -2587 to -2296 [SEQ. ID. NO.: 56] and -1138 to -877 [SEQ. ID. NO.: 57] (with respect to the transcription start determined by RNP). The sequence proximal to the 5' end is strongly homologous (91.4% identity) to the U3 region of long terminal repeats of human endogenous retroviruses HERV-K [Ono, M., "Molecular cloning and long terminal repeat sequences of human endogenous retrovirus genes related to types A and B retrovirus genes," J. Virol. 58: 937-944 (1986)]. The LTR-like fragment is 222 bp long with an A-rich tail at its 3' end. Most probably, it represents part of SINE (short interspersed repeated sequence) type nonviral retroposon derived from HERV-K [Ono et al., "A novel human nonviral retroposon derived from an endogenous retrovirus," Nucleic Acids Res., 15: 8725-8373 (1987)]. There are no sequences corresponding to regulatory elements in this fragment, since the 3' part of U3, and the entire R and U5 regions of LTR are absent from the Bd3 genomic clone, and the glucocorticoid responsive element as well as the enhancer core sequences are beyond its 5' border.

However, two keratinocyte-dependent enhancers were identified in the sequence downstream from the LTR-like fragment at positions -3010 and -2814. Those elements are involved in transcriptional regulation of the E6-E7 oncogenes of human papillomaviruses and are thought to account for their tissue specificity [Cripe et al., "Transcriptional regulation of the human papilloma-virus-16 E6-E7 promoter by a keratinocyte-dependent enhancer, and by viral E2 trans-activator and repressor gene products: implications for cervical carcinogenesis," EMBO J., 6: 3745-3753 (1987)].

Nucleotide sequence analysis of the DNA 5' to the transcription start (from nt -507) revealed no recognizable TATA box within the expected distance from the beginning of the first exon (Figure 6). However, the presence of potential

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binding sites for transcription factors suggests that this region might contain a promoter for the MN gene. There are several consensus sequences for transcription factors AP1 and AP2 as well as for other regulatory elements, including a p53 binding site [Locker and Buzard, "A dictionary of transcription control sequences," J. DNA Sequencing and Mapping, 1: 3-11 (1990); Imagawa et al., "Transcription factor AP-2 mediates induction by two different signal-transduction pathways: protein kinase C and cAMP," Cell, 51: 251-260 (1987); El Deiry et al., "Human genomic DNA sequences define a consensus binding site for p53," Nat. Genet., 1: 44-49 (1992)]. Although the putative promoter region contains 59.3% C+G, it does not have additional attributes of CpG-rich islands that are typical for TATA-less promoters of housekeeping genes [Bird, "CpG-rich islands and the function of DNA methylation," Nature, 321: 209-213 (1986)]. Another class of genes lacking TATA box utilizes the initiator (Inr) element as a promoter. Many of these genes are not constitutively active, but they are rather regulated during differentiation or development. The Inr has a consensus sequence of PyPyPyCAPyPyPyPyPy [SEQ. ID. NO.: 23] and encompasses the transcription start site [Smale and Baltimore, "The 'initiator' as a transcription control element," Cell, 57: 103-113 (1989)]. There are two such consensus sequences in the MN putative promoter; however, they do not overlap the transcription start (Figure 6).

In the initial experiments, the inventors were unable to show promoter activity in human carcinoma cells HeLa and CGL3 that express MN, using the 3.5 kb Bd3 fragment and series of its deletion mutants (from nt -933 to -30) [SEQ. ID. NO.: 58] fused to chloramphenicol acetyl transferase (CAT) gene in a transient system. This might indicate that either the promoter activity of the region 5' to the MN transcription start is below the sensitivity of the CAT assay, or additional regulatory elements not present in our constructs are required for driving the expression of MN gene.

With respect to this fact, an interesting region was found in the middle of the MN gene. The region is about 1.4

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kb in length [nt 4,600-6,000 of the genomic sequence; SEQ. ID. NO.: 49] and spans from the 3' part of the 1st intron to the end of the 5th exon. The region has the character of a typical CpG-rich island, with 62.8% C+G content and 82 CpG: 5 131 GpC dinucleotides. Moreover, there are multiple putative binding sites for transcription factors AP2 and Sp1 [Locker and Buzard, supra; Briggs et al., "Purification and biochemical characterization of the promoter-specific transcription factor Sp-1," Science, 234: 47-52 (1986)] 10 concentrated in the center of this area. Particularly the 3rd intron of 131 bp in length contains three Sp1 and three AP2 consensus sequences. That data indicates the possible involvement of that region in the regulation of MN gene expression. However, functionality of that region, as well as 15 other regulatory elements found in the proposed 5' MN promoter, remains to be determined.

MN Promoter Analysis

To define sequences necessary for MN gene expression, a series of 5' deletion mutants of the putative 20 promoter region were fused to the bacterial chloramphenicol acetyltransferase (CAT) gene. [See Figure 8.] The pMN-CAT deletion constructs were transfected using a DEAE dextran method for transient expression into HeLa and CGL3 cells. Those cells were used since they naturally express MN protein, 25 and thus, should contain all the required transcription factors.

After 48 hours, crude cell lysates were prepared and the activity of the expressed CAT was evaluated according to acetylation of [¹⁴C]chloramphenicol by thin layer 30 chromatography. However, no MN promoter CAT activity was detected in either the HeLa or the CGL3 cells in a transient system. On the other hand, reporter CAT plasmids with viral promoters (e.g. pBLV-LTR + tax transactivator, pRSV CAT and pSV2 CAT), that served as positive controls, gave strong 35 signals on the chromatogram. [pSV2 CAT carries the SV40 origin and expresses CAT from the SV40 early promoter (P_E).

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pRSV CAT expresses CAT from the Rous sarcoma virus (RSV) LTR promoter (P_{LTR}).]

No detectable CAT activity was observed in additional experiments using increasing amounts of transfected plasmids (from 2 to 20 g DNA per dish) and prolonged periods of cell incubation after transcription. Increased cell density also did not improve the results (in contrast to the expectations based on density-dependent expression of native MN protein in HeLa cells). Since the inventors had found consensus sequences for transcription factors AP2 and AP1 in the putative MN promoter, they studied the effect of their inducers dexamethasone (1 μ M) and phorbol ester phorbol 12-myristate 13-acetate (PMA 50 ng/ml) on CAT activity. However, the MN promoter was unresponsive to those compounds.

The following provides explanations for the results: --the putative MN promoter immediately preceding the transcription initiation site is very weak, and its activity is below the sensitivity of a standard CAT assay; --additional sequences (e.g. enhancers) are necessary for MN transcription.

To further shed light on the regulation of MN expression at the level of transcription, constructs, analogously prepared to the MN-CAT constructs, are prepared, wherein the MN promoter region is upstream from the neomycin phosphotransferase gene engineered for mammalian expression. Such constructs are then transfected to cells which are subjected to selection with G418. Activity of the promoter is then evaluated on the basis of the number of G418 resistant colonies that result. That method has the capacity to detect activity of a promoter that is 50 to 100 times weaker in comparison to promoters detectable by a CAT assay.

Deduced Amino Acid Sequence

The ORF of the MN cDNA shown in Figure 1 has the coding capacity for a 459 amino acid protein with a calculated molecular weight of 49.7 kd. MN protein has an estimated pI of about 4. As assessed by amino acid sequence analysis, the deduced primary structure of the MN protein can be divided into four distinct regions. The initial hydrophobic region of

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37 amino acids (AA) corresponds to a signal peptide. The mature protein has an N-terminal part of 377 AA, a hydrophobic transmembrane segment of 20 AA and a C-terminal region of 25 AA. Alternatively, the MN protein can be viewed as having

5 five domains as follows: (1) a signal peptide [amino acids (AA) 1-37; SEQ. ID. NO.: 6]; (2) a region of homology to collagen alpha1 chain (AA 38-135; SEQ. ID. NO.: 50); (3) a carbonic anhydrase domain (AA 136-391; SEQ. ID. NO.: 51); (4)

10 a transmembrane region (AA 414-433; SEQ. ID. NO.: 52); and (5) an intracellular C terminus (AA 434-459; SEQ. ID. NO.: 53). (The AA numbers are keyed to Figure 1.)

More detailed insight into MN protein primary structure disclosed the presence of several consensus sequences. One potential N-glycosylation site was found at

15 position 346 of Figure 1. That feature, together with a predicted membrane-spanning region are consistent with the results, in which MN was shown to be an N-glycosylated protein localized in the plasma membrane. MN protein sequence deduced from cDNA was also found to contain seven S/TPXX sequence

20 elements [SEQ. ID. NOS.: 25 AND 26] (one of them is in the signal peptide) defined by Suzuki, J. Mol. Biol., 207: 61-84 (1989) as motifs frequently found in gene regulatory proteins. However, only two of them are composed of the suggested consensus amino acids.

25 Experiments have shown that the MN protein is able to bind zinc cations, as shown by affinity chromatography using Zn-charged chelating sepharose. MN protein immunoprecipitated from HeLa cells by Mab M75 was found to have weak catalytic activity of CA. The CA-like domain of MN

30 has a structural predisposition to serve as a binding site for small soluble domains. Thus, MN protein could mediate some kind of signal transduction.

MN protein from LCMV-infected HeLa cells was shown by using DNA cellulose affinity chromatography to bind to

35 immobilized double-stranded salmon sperm DNA. The binding activity required both the presence of zinc cations and the absence of a reducing agent in the binding buffer.

Sequence Similarities

Computer analysis of the MN cDNA sequence was carried out using DNASIS and PROSID (Pharmacia Software packages). GenBank, EMBL, Protein Identification Resource and
5 SWISS-PROT databases were searched for all possible sequence similarities. In addition, a search for proteins sharing sequence similarities with MN was performed in the MIPS databank with the FastA program [Pearson and Lipman, PNAS (USA), 85: 2444 (1988)].

10 The MN gene was found to clearly be a novel sequence derived from the human genome. Searches for amino acid sequence similarities in protein databases revealed as the closest homology a level of sequence identity (38.9% in 256 AA or 44% in an 170 AA overlap) between the central part of the
15 MN protein [AAs 136-391 (SEQ. ID. NO: 51)] or 221-390 [SEQ. ID. NO.: 54] of Figure 1 and carbonic anhydrases (CA). However, the overall sequence homology between the cDNA MN sequence and cDNA sequences encoding different CA isoenzymes is in a homology range of 48-50% which is considered by ones
20 in the art to be low. Therefore, the MN cDNA sequence is not closely related to any CA cDNA sequences.

Only very closely related nt sequences having a homology of at least 80-90% would hybridize to each other under stringent conditions. A sequence comparison of the MN
25 cDNA sequence shown in Figure 1 and a corresponding cDNA of the human carbonic anhydrase II (CA II) showed that there are no stretches of identity between the two sequences that would be long enough to allow for a segment of the CA II cDNA sequence having 50 or more nucleotides to hybridize under
30 stringent hybridization conditions to the MN cDNA or vice versa.

Although MN deduced amino acid sequences show some homology to known carbonic anhydrases, they differ from them in several respects. Seven carbonic anhydrases are known
35 [Dodgson et al. (eds.), The Carbonic Anhydrases, (Plenum Press; New York/London (1991))]. All of the known carbonic anhydrases are proteins of about 30 kd, smaller than the p54/58N-related products of the MN gene. Further, the

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carbonic anhydrases do not form oligomers as do the MN-related proteins.

The N-terminal part of the MN protein (AA 38-135; SEQ. ID. NO.: 50) shows a 27-30% identity with human collagen 5 alpha1 chain, which is an important component of the extracellular matrix.

MN Proteins and/or Polypeptides

The phrase "MN proteins and/or polypeptides" (MN proteins/polypeptides) is herein defined to mean proteins and/or polypeptides encoded by an MN gene or fragments thereof. An exemplary and preferred MN protein according to this invention has the deduced amino acid sequence shown in Figure 1. Preferred MN proteins/polypeptides are those proteins and/or polypeptides that have substantial homology 15 with the MN protein shown in Figure 1. For example, such substantially homologous MN proteins/ polypeptides are those that are reactive with the MN-specific antibodies of this invention, preferably the Mabs M75, MN12, MN9 and MN7 or their equivalents.

20 A "polypeptide" is a chain of amino acids covalently bound by peptide linkages and is herein considered to be composed of 50 or less amino acids. A "protein" is herein defined to be a polypeptide composed of more than 50 amino acids.

25 MN proteins exhibit several interesting features: cell membrane localization, cell density dependent expression in HeLa cells, correlation with the tumorigenic phenotype of HeLa x fibroblast somatic cell hybrids, and expression in several human carcinomas among other tissues. As demonstrated 30 herein, for example, in Example 1, MN protein can be found directly in tumor tissue sections but not in general in counterpart normal tissues (exceptions noted infra in Example 1 as in normal stomach tissues). MN is also expressed sometimes in morphologically normal appearing areas of tissue 35 specimens exhibiting dysplasia and/or malignancy. Taken together, these features suggest a possible involvement of MN

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in the regulation of cell proliferation, differentiation and/or transformation.

It can be appreciated that a protein or polypeptide produced by a neoplastic cell in vivo could be altered in sequence from that produced by a tumor cell in cell culture or by a transformed cell. Thus, MN proteins and/or polypeptides which have varying amino acid sequences including without limitation, amino acid substitutions, extensions, deletions, truncations and combinations thereof, fall within the scope of this invention. It can also be appreciated that a protein extant within body fluids is subject to degradative processes, such as, proteolytic processes; thus, MN proteins that are significantly truncated and MN polypeptides may be found in body fluids, such as, sera. The phrase "MN antigen" is used herein to encompass MN proteins and/or polypeptides.

It will further be appreciated that the amino acid sequence of MN proteins and polypeptides can be modified by genetic techniques. One or more amino acids can be deleted or substituted. Such amino acid changes may not cause any measurable change in the biological activity of the protein or polypeptide and result in proteins or polypeptides which are within the scope of this invention, as well as, MN muteins.

The MN proteins and polypeptides of this invention can be prepared in a variety of ways according to this invention, for example, recombinantly, synthetically or otherwise biologically, that is, by cleaving longer proteins and polypeptides enzymatically and/or chemically. A preferred method to prepare MN proteins is by a recombinant means. Particularly preferred methods of recombinantly producing MN proteins are described below for the GEX-3X-MN, MN 20-19, MN-Fc and MN-PA proteins.

Recombinant Production of MN Proteins and Polypeptides

A representative method to prepare the MN proteins shown in Figure 1 or fragments thereof would be to insert the full-length or an appropriate fragment of MN cDNA into an appropriate expression vector as exemplified below. In Zavada et al., WO 93/18152, supra, production of a fusion protein

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GEX-3X-MN using the partial cDNA clone (described above) in the vector pGEX-3X (Pharmacia) is described. Nonglycosylated GEX-3X-MN (the Mn fusion protein MN glutathione S-transferase) from XL1-Blue cells. Herein described is the recombinant
5 production of both a glycosylated MN protein expressed from insect cells and a nonglycosylated MN protein expressed from E. coli using the expression plasmid pEt-22b [Novagen Inc.; Madison, WI (USA)].

Baculovirus Expression Systems. Recombinant
10 baculovirus express vectors have been developed for infection into several types of insect cells. For example, recombinant baculoviruses have been developed for among others: Aedes aegypti, Autographa californica, Bombyx mor, Drosophila melanogaster, Heliothis zea, Spodoptera frugiperda, and
15 Trichoplusia ni [PCT Pub. No. WO 89/046699; Wright, Nature, 321: 718 (1986); Fraser et al., In Vitro Cell Dev. Biol., 25: 225 (1989). Methods of introducing exogenous DNA into insect hosts are well-known in the art. DNA transfection and viral infection procedures usually vary with the insect genus to be
20 transformed. See, for example, Autographa [Carstens et al., Virology, 101: 311 (1980)]; Spodoptera [Kang, "Baculovirus Vectors for Expression of Foreign Genes," in: Advances in Virus Research, 35 (1988)]; and Heliothis (virescens) [PCT Pub. No. WO 88/02030].

25 A wide variety of other host-cloning vector combinations may be usefully employed in cloning the MN DNA isolated as described herein. For example, useful cloning vehicles may include chromosomal, nonchromosomal and synthetic DNA sequences such as various known bacterial plasmids such as pBR322, other E. coli plasmids and their derivatives and wider host range plasmids such as RP4, phage DNA, such as, the numerous derivatives of phage lambda, e.g., NB989 and vectors derived from combinations of plasmids and phage DNAs such as plasmids which have been modified to employ phage DNA
30 expression control sequences.

Useful hosts may be eukaryotic or prokaryotic and include bacterial hosts such as E. coli and other bacterial strains, yeasts and other fungi, animal or plant hosts such as

animal or plant cells in culture, insect cells and other hosts. Of course, not all hosts may be equally efficient. The particular selection of host-cloning vehicle combination may be made by those of skill in the art after due
5 consideration of the principles set forth herein without departing from the scope of this invention.

The particular site chosen for insertion of the selected DNA fragment into the cloning vehicle to form a recombinant DNA molecule is determined by a variety of
10 factors. These include size and structure of the protein or polypeptide to be expressed, susceptibility of the desired protein or polypeptide to endoenzymatic degradation by the host cell components and contamination by its proteins, expression characteristics such as the location of start and
15 stop codons, and other factors recognized by those of skill in the art.

The recombinant nucleic acid molecule containing the MN gene, fragment thereof, or cDNA therefrom, may be employed to transform a host so as to permit that host (transformant)
20 to express the structural gene or fragment thereof and to produce the protein or polypeptide for which the hybrid DNA encodes. The recombinant nucleic acid molecule may also be employed to transform a host so as to permit that host on replication to produce additional recombinant nucleic acid
25 molecules as a source of MN nucleic acid and fragments thereof. The selection of an appropriate host for either of those uses is controlled by a number of factors recognized in the art. These include, for example, compatibility with the chosen vector, toxicity of the co-products, ease of recovery
30 of the desired protein or polypeptide, expression characteristics, biosafety and costs.

Where the host cell is a procaryote such as E. coli, competent cells which are capable of DNA uptake are prepared from cells harvested after exponential growth phase and
35 subsequently treated by the CaCl_2 method by well known procedures. Transformation can also be performed after forming a protoplast of the host cell.

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Where the host used is an eucaryote, transfection methods such as the use of a calcium phosphate-precipitate, electroporation, conventional mechanical procedures such as microinjection, insertion of a plasmid encapsulated in red blood cell ghosts or in liposomes, treatment of cells with agents such as lysophosphatidyl-choline or use of virus vectors, or the like may be used.

The level of production of a protein or polypeptide is governed by three major factors: (1) the number of copies of the gene or DNA sequence encoding for it within the cell; (2) the efficiency with which those gene and sequence copies are transcribed and translated; and (3) the stability of the mRNA. Efficiencies of transcription and translation (which together comprise expression) are in turn dependent upon nucleotide sequences, normally situated ahead of the desired coding sequence. Those nucleotide sequences or expression control sequences define, inter alia, the location at which an RNA polymerase interacts to initiate transcription (the promoter sequence) and at which ribosomes bind and interact with the mRNA (the product of transcription) to initiate translation. Not all such expression control sequences function with equal efficiency. It is thus of advantage to separate the specific coding sequences for the desired protein from their adjacent nucleotide sequences and fuse them instead to known expression control sequences so as to favor higher levels of expression. This having been achieved, the newly engineered DNA fragment may be inserted into a multicopy plasmid or a bacteriophage derivative in order to increase the number of gene or sequence copies within the cell and thereby further improve the yield of expressed protein.

Several expression control sequences may be employed. These include the operator, promoter and ribosome binding and interaction sequences (including sequences such as the Shine-Dalgarno sequences) of the lactose operon of E. coli ("the lac system"), the corresponding sequences of the tryptophan synthetase system of E. coli ("the trp system"), a fusion of the trp and lac promoter ("the tac system"), the major operator and promoter regions of phage lambda ($O_L P_L$ and

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O_RP_R), and the control region of the phage fd coat protein. DNA fragments containing these sequences are excised by cleavage with restriction enzymes from the DNA isolated from transducing phages that carry the lac or trp operons, or from the DNA of phage lambda or fd. Those fragments are then manipulated in order to obtain a limited population of molecules such that the essential controlling sequences can be joined very close to, or in juxtaposition with, the initiation codon of the coding sequence.

The fusion product is then inserted into a cloning vehicle for transformation or transfection of the appropriate hosts and the level of antigen production is measured. Cells giving the most efficient expression may be thus selected. Alternatively, cloning vehicles carrying the lac, trp or lambda P_L control system attached to an initiation codon may be employed and fused to a fragment containing a sequence coding for a MN protein or polypeptide such that the gene or sequence is correctly translated from the initiation codon of the cloning vehicle.

The phrase "recombinant nucleic acid molecule" is herein defined to mean a hybrid nucleotide sequence comprising at least two nucleotide sequences, the first sequence not normally being found together in nature with the second.

The phrase "expression control sequence" is herein defined to mean a sequence of nucleotides that controls and regulates expression of structural genes when operatively linked to those genes.

The following are representative examples of genetically engineering MN proteins of this invention. The descriptions are exemplary and not meant to limit the invention in any way.

Expression of MN 20-19 Protein

A representative, recombinantly produced MN protein of this invention is the MN 20-19 protein which, when produced in baculovirus-infected Sf9 cells [*Spodoptera frugiperda* cells; Clontech; Palo Alto, CA (USA)], is glycosylated. The MN 20-19 protein misses the putative signal peptide (AAs 1-37)

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of SEQ. ID. NO.: 6 (Figure 1), has a methionine (Met) at the N-terminus for expression, and a Leu-Glu-His-His-His-His-His [SEQ. ID NO.: 22] added to the C-terminus for purification.

5 In order to insert the portion of the MN coding sequence for the GEX-3X-MN fusion protein into alternate expression systems, a set of primers for PCR was designed. The primers were constructed to provide restriction sites at each end of the coding sequence, as well as in-frame start and
10 stop codons. The sequences of the primers, indicating restriction enzyme cleavage sites and expression landmarks, are shown below.

Primer #20:N-terminus

15 5'GTCGCTAGCTCCATGGGTCATATGCAGAGGTTGCCCCGGATGCAG 3'
NheI NcoI NdeI Translation start MN cDNA #1 [SEQ. ID. NO. 17]

Primer #19:C-terminus

20 5'GAAGATCTCTTACTCGAGCATTCTCCAAGATCCAGCCTCTAGG 3'
BglII XhoI Translation stop MN cDNA [SEQ. ID. NO. 18]

The SEQ. ID. NOS.: 17 and 18 primers were used to amplify the MN coding sequence present in the GEX-3X-MN vector using standard PCR techniques. The resulting PCR product (termed MN 20-19) was electrophoresed on a 0.5% agarose/1X TBE gel; the
25 1.3 kb band was excised; and the DNA recovered using the Gene Clean II kit according to the manufacturer's instructions [Bio101; LaJolla, CA (USA)].

30 MN 20-19 and plasmid pET-22b were cleaved with the restriction enzymes NdeI and XhoI, phenol-chloroform extracted, and the appropriate bands recovered by agarose gel electrophoresis as above. The isolated fragments were ethanol co-precipitated at a vector:insert ratio of 1:4. After

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resuspension, the fragments were ligated using T4 DNA ligase. The resulting product was used to transform competent Novablu E. coli cells [Novagen, Inc.]. Plasmid mini-preps [Magic Minipreps; Promega] from the resultant ampicillin resistant colonies were screened for the presence of the correct insert by restriction mapping. Insertion of the gene fragment into the pET-22b plasmid using the NdeI and XhoI sites added a 6-histidine tail to the protein that could be used for affinity isolation.

- 10 To prepare MN 20-19 for insertion into the baculovirus expression system, the MN 20-19 gene fragment was excised from pET-22b using the restriction endonucleases XbaI and PvuI. The baculovirus shuttle vector pBacPAK8 [Clontech] was cleaved with XbaI and PacI. The desired fragments (1.3 kb for MN 20-19 and 5.5 kb for pBacPAK8) were isolated by agarose gel electrophoresis, recovered using Gene Clean II, and co-precipitated at an insert:vector ratio of 2.4:1.

- After ligation with T4 DNA ligase, the DNA was used to transform competent NM522 E. coli cells (Stratagene).
20 Plasmid mini-preps from resultant ampicillin resistant colonies were screened for the presence of the correct insert by restriction mapping. Plasmid DNA from an appropriate colony and linearized BacPAK6 baculovirus DNA [Clontech] were used to transform Sf9 cells by standard techniques.
25 Recombination produced BacPAK viruses carrying the MN 20-19 sequence. Those viruses were plated onto Sf9 cells and overlaid with agar.

- Plaques were picked and plated onto Sf9 cells. The conditioned media and cells were collected. A small aliquot of the conditioned media was set aside for testing. The cells were extracted with PBS with 1% Triton X100.

- The conditioned media and the cell extracts were dot blotted onto nitrocellulose paper. The blot was blocked with 5% non-fat dried milk in PBS. Mab M75 were used to detect the MN 20-19 protein in the dot blots. A rabbit anti-mouse Ig-HRP was used to detect bound Mab M75. The blots were developed with TMB/H₂O₂ with a membrane enhancer [KPL; Gaithersburg, MD (USA)]. Two clones producing the strongest reaction on the

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dot blots were selected for expansion. One was used to produce MN 20-19 protein in High Five cells [Invitrogen Corp., San Diego, CA (USA); BTI-TN-5BI-4; derived from Trichoplusia ni egg cell homogenate]. MN 20-19 protein was purified from the conditioned media from the virus infected High Five cells.

The MN 20-19 protein was purified from the conditioned media by immunoaffinity chromatography. 6.5 mg of Mab M75 was coupled to 1 g of Tresyl activated Toyopearl™ [Tosoh, Japan (#14471)]. Approximately 150 ml of the conditioned media was run through the M75-Toyopearl column. The column was washed with PBS, and the MN 20-19 protein was eluted with 1.5 M MgCl. The eluted protein was then dialyzed against PBS.

15 Fusion Proteins with C-Terminal Part Including Transmembrane Region Replaced by Fc or PA

MN fusion proteins in which the C terminal part including the transmembrane region is replaced by the Fc fragment of human IgG or by Protein A were constructed. Such fusion proteins are useful to identify MN binding protein(s). In such MN chimaeras, the whole N-terminal part of MN is accessible to interaction with heterologous proteins, and the C terminal tag serves for simple detection and purification of protein complexes.

Fusion Protein MN-PA (Protein A)

In a first step, the 3' end of the MN cDNA encoding the transmembrane region of the MN protein was deleted. The plasmid pFLMN (e.g. pBluescript with full length MN cDNA) was cleaved by EcoRI and blunt ended by S1 nuclease. Subsequent cleavage by SacI resulted in the removal of the EcoRI-SacI fragment. The deleted fragment was then replaced by a Protein A coding sequence that was derived from plasmid pEZZ (purchased from Pharmacia), which had been cleaved with RsaI and SacI. The obtained MN-PA construct was subcloned into a eukaryotic expression vector pSG5C (described in Example 3), and was then ready for transfection experiments.

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Fusion Protein MN-Fc

The cloning of the fusion protein MN-Fc was rather complicated due to the use of a genomic clone containing the Fc fragment of human IgG which had a complex structure in that it contained an enhancer, a promoter, exons and introns. Moreover, the complete sequence of the clone was not available. Thus, it was necessary to ensure the correct in-phase splicing and fusion of MN to the Fc fragment by the addition of a synthetic splice donor site (SSDS) designed according to the splicing sequences of the MN gene.

The construction procedure was as follows:

1. Plasmid pMH4 (e.g. pSV2gpt containing a genomic clone of the human IgG Fc region) was cleaved by *Bam*HI in order to get a 13 kb fragment encoding Fc. [In pSV2gpt, the *E. coli* xanthine-guanine phosphoribosyl transferase gene (*gpt*) is expressed using the SV40 early promoter (P_E) located in the SV40 origin, the SV40 small T intron, and the SV40 polyadenylation site.]

2. At the same time, plasmid pFLMN (with full length MN cDNA) was cleaved by *Sal*I-*Eco*RI. The released fragment was purified and ligated with a synthetic adapter *Eco*RI-*Bgl*II containing a synthetic splice donor site (SSDS).

3. Simultaneously, the plasmid pBKCMV was cleaved by *Sal*I-*Bam*HI. Then advantage was taken of the fact that the *Bam*HI cohesive ends (of the Fc coding fragment) are compatible with the *Bgl*II ends of the SSDS, and Fc was ligated to MN. The MN-Fc ligation product was then inserted into pBKCMV by directional cloning through the *Sal*I and *Bam*HI sites.

Verification of the correct orientation and in-phase fusion of the obtained MN-Fc chimaeric clones was problematic in that the sequence of Fc was not known. Thus, functional constructs are selected on the basis of results of transient eukaryotic expression analyses.

Synthetic and Biologic Production of MN Proteins and Polypeptides

MN proteins and polypeptides of this invention may be prepared not only by recombinant means but also by

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synthetic and by other biologic means. Synthetic formation of the polypeptide or protein requires chemically synthesizing the desired chain of amino acids by methods well known in the art. Exemplary of other biologic means to prepare the desired polypeptide or protein is to subject to selective proteolysis a longer MN polypeptide or protein containing the desired amino acid sequence; for example, the longer polypeptide or protein can be split with chemical reagents or with enzymes.

Chemical synthesis of a peptide is conventional in the art and can be accomplished, for example, by the Merrifield solid phase synthesis technique [Merrifield, J., Am. Chem. Soc., 85: 2149-2154 (1963); Kent et al., Synthetic Peptides in Biology and Medicine, 29 f.f. eds. Alitalo et al., (Elsevier Science Publishers 1985); and Haug, J.D., "Peptide Synthesis and Protecting Group Strategy", American Biotechnology Laboratory, 5(1): 40-47 (Jan/Feb. 1987)].

Techniques of chemical peptide synthesis include using automatic peptide synthesizers employing commercially available protected amino acids, for example, Biosearch [San Rafael, CA (USA)] Models 9500 and 9600; Applied Biosystems, Inc. [Foster City, CA (USA)] Model 430; Milligen [a division of Millipore Corp.; Bedford, MA (USA)] Model 9050; and Du Pont's RAMP (Rapid Automated Multiple Peptide Synthesis) [Du Pont Compass, Wilmington, DE (USA)].

25 Regulation of MN Expression and MN Promoter

MN appears to be a novel regulatory protein that is directly involved in the control of cell proliferation and in cellular transformation. In HeLa cells, the expression of MN is positively regulated by cell density. Its level is increased by persistent infection with LCMV. In hybrid cells between HeLa and normal fibroblasts, MN expression correlates with tumorigenicity. The fact that MN is not present in nontumorigenic hybrid cells (CGL1), but is expressed in a tumorigenic segregant lacking chromosome 11, indicates that MN is negatively regulated by a putative suppressor in chromosome 11.

Evidence supporting the regulatory role of MN protein was found in the generation of stable transfectants of NIH 3T3 cells that constitutively express MN protein as described in Example 3. As a consequence of MN expression, the NIH 3T3 cells acquired features associated with a transformed phenotype: altered morphology, increased saturation density, proliferative advantage in serum-reduced media, enhanced DNA synthesis and capacity for anchorage-independent growth. Further, as shown in Example 4, flow cytometric analyses of asynchronous cell populations indicated that the expression of MN protein leads to accelerated progression of cells through G1 phase, reduction of cell size and the loss of capacity for growth arrest under inappropriate conditions. Also, Example 4 shows that MN expressing cells display a decreased sensitivity to the DNA damaging drug mitomycin C.

Nontumorigenic human cells, CGL1 cells, were also transfected with the full-length MN cDNA. The same pSG5C-MN construct in combination with pSV2neo plasmid as used to transfect the NIH 3T3 cells (Example 3) was used. Also the protocol was the same except that the G418 concentration was increased to 1000 μ g/ml.

Out of 15 MN-positive clones (tested by SP-RIA and Western blotting), 3 were chosen for further analysis. Two MN-negative clones isolated from CGL1 cells transfected with empty plasmid were added as controls. Initial analysis indicates that the morphology and growth habits of MN-transfected CGL1 cells are not changed dramatically, but their proliferation rate and plating efficiency is increased.

MN cDNA and promoter. When the promoter region from the MN genomic clone, isolated as described above, was linked to MN cDNA and transfected into CGL1 hybrid cells, expression of MN protein was detectable immediately after selection. However, then it gradually ceased, indicating thus an action of a feedback regulator. The putative regulatory element appeared to be acting via the MN promoter, because when the full-length cDNA (not containing the promoter) was used for transfection, no similar effect was observed.

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An "antisense" MN cDNA/MN promoter construct was used to transfect CGL3 cells. The effect was the opposite of that of the CGL1 cells transfected with the "sense" construct. Whereas the transfected CGL1 cells formed colonies several
5 times larger than the control CGL1, the transfected CGL3 cells formed colonies much smaller than the control CGL3 cells.

For those experiments, the part of the promoter region that was linked to the MN cDNA through a BamHI site was derived from a NcoI - BamHI fragment of the MN genomic clone
10 [Bd3] and represents a region a few hundred bp upstream from the transcription initiation site. After the ligation, the joint DNA was inserted into a pBK-CMV expression vector [Stratagene]. The required orientation of the inserted
15 sequence was ensured by directional cloning and subsequently verified by restriction analysis. The tranfection procedure was the same as used in transfecting the NIH 3T3 cells (Example 3), but co-transfection with the pSV2neo plasmid was not necessary since the neo selection marker was already
included in the pBK-CMV vector.

20 After two weeks of selection in a medium containing G418, remarkable differences between the numbers and sizes of the colonies grown were evident as noted above. Immediately following the selection and cloning, the MN-transfected CGL1 and CGL3 cells were tested by SP-RIA for expression and
25 repression of MN, respectively. The isolated transfected CGL1 clones were MN positive (although the level was lower than obtained with the full-length cDNA), whereas MN protein was almost absent from the transfected CGL3 clones. However, in subsequent passages, the expression of MN in transfected CGL1
30 cells started to cease, and was then blocked perhaps evidencing a control feedback mechanism.

As a result of the very much lowered proliferation of the transfected CGL3 cells, it was difficult to expand the majority of cloned cells (according to SP-RIA, those with the
35 lowest levels of MN), and they were lost during passaging. However, some clones overcame that problem and again expressed MN. It is possible that once those cells reached a higher quantity, that the level of endogenously produced MN mRNA

increased over the amount of ectopically expressed antisense mRNA.

Transformation and Reversion

As illustrated in Examples 3 and 4, vertebrate cells
5 transfected with MN cDNA in suitable vectors show striking
morphologic transformation. Transformed cells may be very
small, densely packed, slowly growing, with basophilic
cytoplasm and enlarged Golgi apparatus. However, it has been
found that transformed clones revert over time, for example,
10 within 3-4 weeks, to nearly normal morphology, even though the
cells may be producing MN protein at high levels. MN protein
is biologically active even in yeast cells; depending upon the
level of its expression, it stimulates or retards their growth
and induces morphologic alterations.

15 Full-length MN cDNA was inserted into pGD, a MLV-
derived vector, which together with standard competent MLV
(murine leukemia virus), forms an infectious, transmissible
complex [pGD-MN + MLV]. That complex also transforms
vertebrate cells, such as, NIH 3T3 cells and mouse embryo
20 fibroblasts BALB/c, which also revert to nearly normal
morphology. Such revertants again contain MN protein and
produce the [pGD-MN + MLV] artificial virus complex, which
retains its transforming capacity. Thus, reversion of MN-
transformed cells is apparently not due to a loss, silencing
25 or mutation of MN cDNA, but may be the result of the
activation of suppressor gene(s).

Nucleic Acid Probes and Test Kits

Nucleic acid probes of this invention are those
comprising sequences that are complementary or substantially
30 complementary to the MN cDNA sequence shown in Figure 1 or to
other MN gene sequences, such as, the complete genomic
sequence of Figure 3a-d [SEQ. ID. NO.: 5] and the putative
promoter sequence [SEQ. ID. NO.: 27 of Figure 6]. The phrase
"substantially complementary" is defined herein to have the
35 meaning as it is well understood in the art and, thus, used in
the context of standard hybridization conditions. The

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stringency of hybridization conditions can be adjusted to control the precision of complementarity. Two nucleic acids are, for example, substantially complementary to each other, if they hybridize to each other under stringent hybridization conditions.

Stringent hybridization conditions are considered herein to conform to standard hybridization conditions understood in the art to be stringent. For example, it is generally understood that stringent conditions encompass relatively low salt and/or high temperature conditions, such as provided by 0.02 M to 0.15 M NaCl at temperatures of 50°C to 70°C. Less stringent conditions, such as, 0.15 M to 0.9 M salt at temperatures ranging from 20°C to 55°C can be made more stringent by adding increasing amounts of formamide, which serves to destabilize hybrid duplexes as does increased temperature.

Exemplary stringent hybridization conditions are described in Sambrook et al., Molecular Cloning: A Laboratory Manual, pages 1.91 and 9.47-9.51 (Second Edition, Cold Spring Harbor Laboratory Press; Cold Spring Harbor, NY; 1989); Maniatis et al., Molecular Cloning: A Laboratory Manual, pages 387-389 (Cold Spring Harbor Laboratory; Cold Spring Harbor, NY; 1982); Tsuchiya et al., Oral Surgery, Oral Medicine, Oral Pathology, 71(6): 721-725 (June 1991).

Preferred nucleic acid probes of this invention are fragments of the isolated nucleic acid sequences that encode MN proteins or polypeptides according to this invention. Preferably those probes are composed of at least twenty-nine nucleotides, more preferably, fifty nucleotides.

Nucleic acid probes of this invention need not hybridize to a coding region of MN. For example, nucleic acid probes of this invention may hybridize partially or wholly to a non-coding region of the genomic sequence shown in Figure 3a-d [SEQ. ID. NO.: 5]. Conventional technology can be used to determine whether fragments of SEQ. ID. NO.: 5 or related nucleic acids are useful to identify MN nucleic acid sequences. [See, for example, Benton and Davis, supra and Fuscoe et al., supra.]

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Areas of homology of the MN nt sequence to other non-MN nt sequences are indicated above. In general, nucleotide sequences that are not in the Alu or LTR-like regions, of preferably 29 bases or more, or still more preferably of 50 bases or more, can be routinely tested and screened and found to hybridize under stringent conditions to only MN nucleotide sequences. Further, not all homologies within the Alu-like MN genomic sequences are so close to Alu repeats as to give a hybridization signal under stringent hybridization conditions. The percent of homology between MN Alu-like regions and a standard Alu-J sequence are indicated as follows:

15	<u>Region of Homology within</u>	<u>SEQ.</u>	<u>% Homology to</u>
	<u>MN Genomic Sequence</u>		
	<u>[SEQ. ID. NO.: 5;</u>	<u>ID.</u>	<u>Entire Alu-J</u>
	<u>Figure 3a-d]</u>	<u>NOS.</u>	<u>Sequence</u>
	921-1212	59	89.1%
	2370-2631	60	78.6%
	4587-4880	61	90.1%
20	6463-6738	62	85.4%
	7651-7939	63	91.0%
	9020-9317	64	69.8%
			<u>% Homology to</u>
			<u>One Half of</u>
			<u>Alu-J Sequence</u>
	8301-8405	65	88.8%
	10040-10122	66	73.2%.

25 Nucleic acid probes of this invention can be used to detect MN DNA and/or RNA, and thus can be used to test for the presence or absence of MN genes, and amplification(s), mutation(s) or genetic rearrangements of MN genes in the cells of a patient. For example, overexpression of an MN gene may
30 be detected by Northern blotting and RNase protection analysis using probes of this invention. Gene alterations, as amplifications, translocations, inversions, and deletions among others, can be detected by using probes of this

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invention for in situ hybridization to chromosomes from a patient's cells, whether in metaphase spreads or interphase nuclei. Southern blotting could also be used with the probes of this invention to detect amplifications or deletions of MN genes. Restriction Fragment Length Polymorphism (RFLP) analysis using said probes is a preferred method of detecting gene alterations, mutations and deletions. Said probes can also be used to identify MN proteins and/or polypeptides as well as homologs or near homologs thereto by their hybridization to various mRNAs transcribed from MN genes in different tissues.

Probes of this invention thus can be useful diagnostically/prognostically. Said probes can be embodied in test kits, preferably with appropriate means to enable said probes when hybridized to an appropriate MN gene or MN mRNA target to be visualized. Such samples include tissue specimens including smears, body fluids and tissue and cell extracts.

PCR Assays

To detect relatively large genetic rearrangements, hybridization tests can be used. To detect relatively small genetic rearrangements, as, for example, small deletions or amplifications, or point mutations, PCR would preferably be used. [U.S. Patent Nos. 4,800,159; 4,683,195; 4,683,202; and Chapter 14 of Sambrook et al., Molecular Cloning: A Laboratory Manual, supra]

An exemplary assay would use cellular DNA from normal and cancerous cells, which DNA would be isolated and amplified employing appropriate PCR primers. The PCR products would be compared, preferably initially, on a sizing gel to detect size changes indicative of certain genetic rearrangements. If no differences in sizes are noted, further comparisons can be made, preferably using, for example, PCR-single-strand conformation polymorphism (PCR-SSCP) assay or a denaturing gradient gel electrophoretic assay. [See, for example, Hayashi, K., "PCR-SSCP: A Simple and Sensitive Method for Detection of Mutations in the Genomic DNA," in PCR

Methods and Applications, 1: 34-38 (1991); and Meyers et al., "Detection and Localization of Single Base Changes by Denaturing Gradient Gel Electrophoresis," Methods in Enzymology, 155: 501 (1987).]

5

Assays

Assays according to this invention are provided to detect and/or quantitate MN antigen or MN-specific antibodies in vertebrate samples, preferably mammalian samples, more preferably human samples. Such samples include tissue specimens, body fluids, tissue extracts and cell extracts. MN antigen may be detected by immunoassay, immunohistochemical staining, immunoelectron and scanning microscopy using immunogold among other techniques.

Preferred tissue specimens to assay by immunohistochemical staining include cell smears, histological sections from biopsied tissues or organs, and imprint preparations among other tissue samples. Such tissue specimens can be variously maintained, for example, they can be fresh, frozen, or formalin-, alcohol- or acetone- or otherwise fixed and/or paraffin-embedded and deparaffinized. Biopsied tissue samples can be, for example, those samples removed by aspiration, bite, brush, cone, chorionic villus, endoscopic, excisional, incisional, needle, percutaneous punch, and surface biopsies, among other biopsy techniques.

Preferred cervical tissue specimens include cervical smears, conization specimens, histologic sections from hysterectomy specimens or other biopsied cervical tissue samples. Preferred means of obtaining cervical smears include routine swab, scraping or cytobrush techniques, among other means. More preferred are cytobrush or swab techniques. Preferably, cell smears are made on microscope slides, fixed, for example, with 55% EtOH or an alcohol based spray fixative and air-dried.

Papanicolaou-stained cervical smears (Pap smears) can be screened by the methods of this invention, for example, for retrospective studies. Preferably, Pap smears would be decolorized and re-stained with labeled antibodies against MN

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antigen. Also archival specimens, for example, matched smears and biopsy and/or tumor specimens, can be used for retrospective studies. Prospective studies can also be done with matched specimens from patients that have a higher than
5 normal risk of exhibiting abnormal cervical cytopathology.

Preferred samples in which to assay MN antigen by, for example, Western blotting or radioimmunoassay, are tissue and/or cell extracts. However, MN antigen may be detected in body fluids, which can include among other fluids: blood,
10 serum, plasma, semen, breast exudate, saliva, tears, sputum, mucous, urine, lymph, cytosols, ascites, pleural effusions, amniotic fluid, bladder washes, bronchioalveolar lavages and cerebrospinal fluid. It is preferred that the MN antigen be concentrated from a larger volume of body fluid before
15 testing. Preferred body fluids to assay would depend on the type of cancer for which one was testing, but in general preferred body fluids would be breast exudate, pleural effusions and ascites.

MN-specific antibodies can be bound by serologically
20 active MN proteins/polypeptides in samples of such body fluids as blood, plasma, serum, lymph, mucous, tears, urine, spinal fluid and saliva; however, such antibodies are found most usually in blood, plasma and serum, preferably in serum. Correlation of the results from the assays to detect and/or
25 quantitate MN antigen and MN-specific antibodies reactive therewith, provides a preferred profile of the disease condition of a patient.

The assays of this invention are both diagnostic and/or prognostic, i.e., diagnostic/prognostic. The term
30 "diagnostic/ prognostic" is herein defined to encompass the following processes either individually or cumulatively depending upon the clinical context: determining the presence of disease, determining the nature of a disease, distinguishing one disease from another, forecasting as to the
35 probable outcome of a disease state, determining the prospect as to recovery from a disease as indicated by the nature and symptoms of a case, monitoring the disease status of a patient, monitoring a patient for recurrence of disease,

and/or determining the preferred therapeutic regimen for a patient. The diagnostic/prognostic methods of this invention are useful, for example, for screening populations for the presence of neoplastic or pre-neoplastic disease, determining
5 the risk of developing neoplastic disease, diagnosing the presence of neoplastic and/or pre-neoplastic disease, monitoring the disease status of patients with neoplastic disease, and/or determining the prognosis for the course of neoplastic disease. For example, it appears that the
10 intensity of the immunostaining with MN-specific antibodies may correlate with the severity of dysplasia present in samples tested.

The present invention is useful for screening for the presence of a wide variety of neoplastic diseases as
15 indicated above. The invention provides methods and compositions for evaluating the probability of the presence of malignant or pre-malignant cells, for example, in a group of cells freshly removed from a host. Such an assay can be used to detect tumors, quantitate their growth, and help in the
20 diagnosis and prognosis of disease. The assays can also be used to detect the presence of cancer metastasis, as well as confirm the absence or removal of all tumor tissue following surgery, cancer chemotherapy and/or radiation therapy. It can further be used to monitor cancer chemotherapy and tumor
25 reappearance.

The presence of MN antigen or antibodies can be detected and/or quantitated using a number of well-defined diagnostic assays. Those in the art can adapt any of the conventional immunoassay formats to detect and/or quantitate
30 MN antigen and/or antibodies.

Many formats for detection of MN antigen and MN-specific antibodies are, of course available. Those can be Western blots, ELISAs, RIAs, competitive EIA or dual antibody sandwich assays, immunohistochemical staining, among other
35 assays all commonly used in the diagnostic industry. In such immunoassays, the interpretation of the results is based on the assumption that the antibody or antibody combination will

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not cross-react with other proteins and protein fragments present in the sample that are unrelated to MN.

Representative of one type of ELISA test for MN antigen is a format wherein a microtiter plate is coated with antibodies made to MN proteins/polypeptides or antibodies made to whole cells expressing MN proteins, and to this is added a patient sample, for example, a tissue or cell extract. After a period of incubation permitting any antigen to bind to the antibodies, the plate is washed and another set of anti-MN antibodies which are linked to an enzyme is added, incubated to allow reaction to take place, and the plate is then rewashed. Thereafter, enzyme substrate is added to the microtiter plate and incubated for a period of time to allow the enzyme to work on the substrate, and the adsorbance of the final preparation is measured. A large change in absorbance indicates a positive result.

It is also apparent to one skilled in the art of immunoassays that MN proteins and/or polypeptides can be used to detect and/or quantitate the presence of MN antigen in the body fluids, tissues and/or cells of patients. In one such embodiment, a competition immunoassay is used, wherein the MN protein/polypeptide is labeled and a body fluid is added to compete the binding of the labeled MN protein/polypeptide to antibodies specific to MN protein/polypeptide.

In another embodiment, an immunometric assay may be used wherein a labeled antibody made to a MN protein or polypeptide is used. In such an assay, the amount of labeled antibody which complexes with the antigen-bound antibody is directly proportional to the amount of MN antigen in the sample.

A representative assay to detect MN-specific antibodies is a competition assay in which labeled MN protein/polypeptide is precipitated by antibodies in a sample, for example, in combination with monoclonal antibodies recognizing MN proteins/polypeptides. One skilled in the art could adapt any of the conventional immunoassay formats to detect and/or quantitate MN-specific antibodies. Detection of the binding of said antibodies to said MN protein/polypeptide

could be by many ways known to those in the art, e.g., in humans with the use of anti-human labeled IgG.

An exemplary immunoassay method of this invention to detect and/or quantitate MN antigen in a vertebrate sample
5 comprises the steps of:

- a) incubating said vertebrate sample with one or more sets of antibodies (an antibody or antibodies) that bind to MN antigen wherein one set is labeled or otherwise detectable;
- 10 b) examining the incubated sample for the presence of immune complexes comprising MN antigen and said antibodies.

Another exemplary immunoassay method according to this invention is that wherein a competition immunoassay is used to detect and/or quantitate MN antigen in a vertebrate
15 sample and wherein said method comprises the steps of:

- a) incubating a vertebrate sample with one or more sets of MN-specific antibodies and a certain amount of a labeled or otherwise detectable MN protein/polypeptide wherein said MN protein/ polypeptide competes for binding to said
20 antibodies with MN antigen present in the sample;
- b) examining the incubated sample to determine the amount of labeled/detectable MN protein/polypeptide bound to said antibodies; and
- c) determining from the results of the examination
25 in step b) whether MN antigen is present in said sample and/or the amount of MN antigen present in said sample.

Once antibodies (including biologically active antibody fragments) having suitable specificity have been prepared, a wide variety of immunological assay methods are
30 available for determining the formation of specific antibody-antigen complexes. Numerous competitive and non-competitive protein binding assays have been described in the scientific and patent literature, and a large number of such assays are commercially available. Exemplary
35 immunoassays which are suitable for detecting a serum antigen include those described in U.S. Patent Nos. 3,984,533; 3,996,345; 4,034,074; and 4,098,876.

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Antibodies employed in assays may be labeled or unlabeled. Unlabeled antibodies may be employed in agglutination; labeled antibodies may be employed in a wide variety of assays, employing a wide variety of labels.

5 Suitable detection means include the use of labels such as radionuclides, enzymes, coenzymes, fluorescers, chemiluminescers, chromogens, enzyme substrates or co-factors, enzyme inhibitors, free radicals, particles, dyes and the like. Such labeled reagents may be used in a variety of well
10 known assays, such as radioimmunoassays, enzyme immunoassays, e.g., ELISA, fluorescent immunoassays, and the like. See for example, U.S. Patent Nos. 3,766,162; 3,791,932; 3,817,837; and 4,233,402.

Immunoassay Test Kits

15 The above outlined assays can be embodied in test kits to detect and/or quantitate MN antigen and/or MN-specific antibodies (including biologically active antibody fragments). Kits to detect and/or quantitate MN antigen can comprise MN
20 protein(s)/polypeptides(s) and/or MN-specific antibodies, polyclonal and/or monoclonal. Such diagnostic/prognostic test kits can comprise one or more sets of antibodies, polyclonal and/or monoclonal, for a sandwich format wherein antibodies recognize epitopes on the MN antigen, and one set is
appropriately labeled or is otherwise detectable.

25 Test kits for an assay format wherein there is competition between a labeled (or otherwise detectable) MN protein/polypeptide and MN antigen in the sample, for binding to an antibody, can comprise the combination of the labeled protein/polypeptide and the antibody in amounts which provide
30 for optimum sensitivity and accuracy.

 Test kits for MN-specific antibodies preferably comprise labeled/detectable MN proteins(s) and/or polypeptides(s), and may comprise other components as necessary, such as, controls, buffers, diluents and
35 detergents. Such test kits can have other appropriate formats for conventional assays.

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A kit for use in an enzyme-immunoassay typically includes an enzyme-labelled reagent and a substrate for the enzyme. The enzyme can, for example, bind either an MN-specific antibody of this invention or to an antibody to such an MN-specific antibody.

Preparation of MN-Specific Antibodies

The term "antibodies" is defined herein to include not only whole antibodies but also biologically active fragments of antibodies, preferably fragments containing the antigen binding regions. Such antibodies may be prepared by conventional methodology and/or by genetic engineering. Antibody fragments may be genetically engineered, preferably from the variable regions of the light and/or heavy chains (V_H and V_L), including the hypervariable regions, and still more preferably from both the V_H and V_L regions. For example, the term "antibodies" as used herein comprehends polyclonal and monoclonal antibodies and biologically active fragments thereof including among other possibilities "univalent" antibodies [Glennie et al., Nature, 295: 712 (1982)]; Fab proteins including Fab' and $F(ab')_2$ fragments whether covalently or non-covalently aggregated; light or heavy chains alone, preferably variable heavy and light chain regions (V_H and V_L regions), and more preferably including the hypervariable regions [otherwise known as the complementarity determining regions (CDRs) of said V_H and V_L regions]; F_c proteins; "hybrid" antibodies capable of binding more than one antigen; constant-variable region chimeras; "composite" immunoglobulins with heavy and light chains of different origins; "altered" antibodies with improved specificity and other characteristics as prepared by standard recombinant techniques and also by oligonucleotide-directed mutagenesis techniques [Dalbadie-McFarland et al., PNAS (USA), 79: 6409 (1982)].

It may be preferred for therapeutic and/or imaging uses that the antibodies be biologically active antibody fragments, preferably genetically engineered fragments, more preferably genetically engineered fragments from the V_H and/or

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V_L regions, and still more preferably comprising the hypervariable regions thereof.

There are conventional techniques for making polyclonal and monoclonal antibodies well-known in the immunoassay art. Immunogens to prepare MN-specific antibodies include MN proteins and/or polypeptides, preferably purified, and MX-infected tumor line cells, for example, MX-infected HeLa cells, among other immunogens.

Anti-peptide antibodies are also made by conventional methods in the art as described in European Patent Publication No. 44,710 (published Jan. 27, 1982). Briefly, such anti-peptide antibodies are prepared by selecting a peptide from an MN amino acid sequence as from Figure 1, chemically synthesizing it, conjugating it to an appropriate immunogenic protein and injecting it into an appropriate animal, usually a rabbit or a mouse; then, either polyclonal or monoclonal antibodies are made, the latter by a Kohler-Milstein procedure, for example.

Besides conventional hybridoma technology, newer technologies can be used to produce antibodies according to this invention. For example, the use of the PCR to clone and express antibody V-genes and phage display technology to select antibody genes encoding fragments with binding activities has resulted in the isolation of antibody fragments from repertoires of PCR amplified V-genes using immunized mice or humans. [Marks et al., BioTechnology, 10: 779 (July 1992) for references; Chiang et al., BioTechniques, 7(4): 360 (1989); Ward et al., Nature, 341: 544 (Oct. 12, 1989); Marks et al., J. Mol. Biol., 222: 581 (1991); Clackson et al., Nature, 352: (15 August 1991); and Mullinax et al., PNAS (USA), 87: 8095 (Oct. 1990).]

Descriptions of preparing antibodies, which term is herein defined to include biologically active antibody fragments, by recombinant techniques can be found in U.S. Patent No. 4,816,567 (issued March 28, 1989); European Patent Application Publication Number (EP) 338,745 (published Oct. 25, 1989); EP 368,684 (published June 16, 1990); EP 239,400 (published September 30, 1987); WO 90/14424 (published Nov.

29, 1990); WO 90/14430 (published May 16, 1990); Huse et al., Science, 246: 1275 (Dec. 8, 1989); Marks et al., BioTechnology, 10: 779 (July 1992); La Sastry et al., PNAS (USA), 86: 5728 (August 1989); Chiang et al., BioTechniques, 7(40): 360 (1989); Orlandi et al., PNAS (USA), 86: 3833 (May 1989); Ward et al. Nature, 341: 544 (October 12, 1989); Marks et al., J. Mol. Biol., 222: 581 (1991); and Hoogenboom et al., Nucleic Acids Res., 19(15): 4133 (1991).

Representative Mabs

10 Monoclonal antibodies for use in the assays of this invention may be obtained by methods well known in the art for example, Galfre and Milstein, "Preparation of Monoclonal Antibodies: Strategies and Procedures," in Methods in Enzymology: Immunochemical Techniques, 73: 1-46 [Langone and 15 Vanatis (eds); Academic Press (1981)]; and in the classic reference, Milstein and Kohler, Nature, 256: 495-497 (1975).]

Although representative hybridomas of this invention are formed by the fusion of murine cell lines, human/human hybridomas [Olsson et al., PNAS (USA), 77: 5429 (1980)] and 20 human/murine hybridomas [Schlom et al., PNAS (USA), 77: 6841 (1980); Shearman et al. J. Immunol., 146: 928-935 (1991); and Gorman et al., PNAS (USA), 88: 4181-4185 (1991)] can also be prepared among other possibilities. Such humanized monoclonal antibodies would be preferred monoclonal antibodies for 25 therapeutic and imaging uses.

Monoclonal antibodies specific for this invention can be prepared by immunizing appropriate mammals, preferably rodents, more preferably rabbits or mice, with an appropriate immunogen, for example, MaTu-infected HeLa cells, MN fusion 30 proteins, or MN proteins/polypeptides attached to a carrier protein if necessary. Exemplary methods of producing antibodies of this invention are described below.

The monoclonal antibodies useful according to this invention to identify MN proteins/polypeptides can be labeled 35 in any conventional manner, for example, with enzymes such as horseradish peroxidase (HRP), fluorescent compounds, or with radioactive isotopes such as, ¹²⁵I, among other labels. A

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preferred label, according to this invention is ^{125}I , and a preferred method of labeling the antibodies is by using chloramine-T [Hunter, W.M., "Radioimmunoassay," In: Handbook of Experimental Immunology, pp. 14.1-14.40 (D.W. Weir ed.; Blackwell, Oxford/London/Edinburgh/Melbourne; 1978)].

Representative mabs of this invention include Mabs M75, MN9, MN12 and MN7 described below. Monoclonal antibodies of this invention serve to identify MN proteins/polypeptides in various laboratory diagnostic tests, for example, in tumor cell cultures or in clinical samples.

Mabs Prepared Against HeLa Cells

Mab M75. Monoclonal antibody M75 (Mab M75) is produced by mouse lymphocytic hybridoma VU-M75, which was initially deposited in the Collection of Hybridomas at the Institute of Virology, Slovak Academy of Sciences (Bratislava, Czechoslovakia) and was deposited under ATCC Designation HB 11128 on September 17, 1992 at the American Type Culture Collection (ATCC) in Rockville, MD (USA). The production of hybridoma VU-M75 is described in Zavada et al., WO 93/18152.

Mab M75 recognizes both the nonglycosylated GEX-3X-MN fusion protein and native MN protein as expressed in CGL3 cells equally well. Mab M75 was shown by epitope mapping to be reactive with the epitope represented by the amino acid sequence from AA 62 to AA 67 [SEQ. ID. NO.: 10] of the MN protein shown in Figure 1.

Mabs Prepared Against Fusion Protein GEX-3X-MN

Monoclonal antibodies of this invention were also prepared against the MN glutathione S-transferase fusion protein (GEX-3X-MN). BALB/C mice were immunized intraperitoneally according to standard procedures with the GEX-3X-MN fusion protein in Freund's adjuvant. Spleen cells of the mice were fused with SP/20 myeloma cells [Milstein and Kohler, supra].

Tissue culture media from the hybridomas were screened against CGL3 and CGL1 membrane extracts in an ELISA employing HRP labelled-rabbit anti-mouse. The membrane

extracts were coated onto microtiter plates. Selected were antibodies reacted with the CGL3 membrane extract. Selected hybridomas were cloned twice by limiting dilution.

The mabs prepared by the just described method were characterized by Western blots of the GEX-3X-MN fusion protein, and with membrane extracts from the CGL1 and CGL3 cells. Representative of the mabs prepared are Mabs MN9, MN12 and MN7.

Mab MN9. Monoclonal antibody MN9 (Mab MN9) reacts to the same epitope as Mab M75, represented by the sequence from AA 62 to AA 67 [SEQ. ID. NO.: 10] of the Figure 1 MN protein. As Mab M75, Mab MN9 recognizes both the GEX-3X-MN fusion protein and native MN protein equally well.

Mabs corresponding to Mab MN9 can be prepared reproducibly by screening a series of mabs prepared against an MN protein/polypeptide, such as, the GEX-3X-MN fusion protein, against the peptide representing the epitope for Mabs M75 and MN9, that is, SEQ. ID. NO.: 10. Alternatively, the Novatope system [Novagen] or competition with the deposited Mab M75 could be used to select mabs comparable to Mabs M75 and MN9.

Mab MN12. Monoclonal antibody MN12 (Mab MN12) is produced by the mouse lymphocytic hybridoma MN 12.2.2 which was deposited under ATCC Designation HB 11647 on June 9, 1994 at the American Type Culture Collection (ATCC) at 12301 Parklawn Drive, Rockville, MD 20852 (USA). Antibodies corresponding to Mab MN12 can also be made, analogously to the method outlined above for Mab MN9, by screening a series of antibodies prepared against an MN protein/polypeptide, against the peptide representing the epitope for Mab MN12. That peptide is AA 55 - AA 60 of Figure 1 [SEQ. ID. NO.: 11]. The Novatope system could also be used to find antibodies specific for said epitope.

Mab MN7. Monoclonal antibody MN7 (Mab MN7) was selected from mabs prepared against nonglycosylated GEX-3X-MN as described above. It recognizes the epitope on MN represented by the amino acid sequence from AA 127 to AA 147 [SEQ. ID. NO.: 12] of the Figure 1 MN protein. Analogously to methods described above for Mabs MN9 and MN12, mabs

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corresponding to Mab MN7 can be prepared by selecting mabs prepared against an MN protein/polypeptide that are reactive with the peptide having SEQ. ID. NO.: 12, or by the stated alternative means.

5 Epitope Mapping

Epitope mapping was performed by the Novatope system, a kit for which is commercially available from Novagen, Inc. [See, for analogous example, Li et al., Nature, 363: 85-88 (6 May 1993).] In brief, the MN cDNA was cut into
10 overlapping short fragments of approximately 60 base pairs. The fragments were expressed in E. coli, and the E. coli colonies were transferred onto nitrocellulose paper, lysed and probed with the mab of interest. The MN cDNA of clones
15 reactive with the mab of interest was sequenced, and the epitopes of the mabs were deduced from the overlapping polypeptides found to be reactive with each mab.

Therapeutic Use of MN-Specific Antibodies

The MN-specific antibodies of this invention, monoclonal and/or polyclonal, preferably monoclonal, and as
20 outlined above, may be used therapeutically in the treatment of neoplastic and/or pre-neoplastic disease, either alone or in combination with chemotherapeutic drugs or toxic agents, such as ricin A. Further preferred for therapeutic use would be biologically active antibody fragments as described herein.
25 Also preferred MN-specific antibodies for such therapeutic uses would be humanized monoclonal antibodies.

The MN-specific antibodies can be administered in a therapeutically effective amount, preferably dispersed in a physiologically acceptable, nontoxic liquid vehicle.

30 Imaging Use of Antibodies

Further, the MN-specific antibodies of this invention when linked to an imaging agent, such as a radionuclide, can be used for imaging. Biologically active antibody fragments or humanized monoclonal antibodies, may be
35 preferred for imaging use.

A patient's neoplastic tissue can be identified as, for example, sites of transfused stem cells, of tumors and locations of any metastases. Antibodies, appropriately labeled or linked to an imaging agent, can be injected in a physiologically acceptable carrier into a patient, and the binding of the antibodies can be detected by a method appropriate to the label or imaging agent, for example, by scintigraphy.

Antisense MN Nucleic Acid Sequences

MN genes are herein considered putative oncogenes and the encoded proteins thereby are considered to be putative oncoproteins. Antisense nucleic acid sequences substantially complementary to mRNA transcribed from MN genes, as represented by the antisense oligodeoxynucleotides ODN1 and ODN2 [SEQ. ID. NOS.: 3 and 4] can be used to reduce or prevent expression of the MN gene. [Zamecnick, P.C., "Introduction: Oligonucleotide Base Hybridization as a Modulator of Genetic Message Readout," pp. 1-6, Prospects for Antisense Nucleic Acid Therapy of Cancer and AIDS, (Wiley-Liss, Inc., New York, NY, USA; 1991); Wickstrom, E., "Antisense DNA Treatment of HL-60 Promyelocytic Leukemia Cells: Terminal Differentiation and Dependence on Target Sequence," pp. 7-24, id.; Leserman et al., "Targeting and Intracellular Delivery of Antisense Oligonucleotides Interfering with Oncogene Expression," pp. 25-34, id.; Yokoyama, K., "Transcriptional Regulation of c-myc Proto-oncogene by Antisense RNA," pp. 35-52, id.; van den Berg et al., "Antisense fos Oligodeoxyribonucleotides Suppress the Generation of Chromosomal Aberrations," pp. 63-70, id.; Mercola, D., "Antisense fos and fun RNA," pp. 83-114, id.; Inouye, Gene, 72: 25-34 (1988); Miller and Ts'o, Ann. Reports Med. Chem., 23: 295-304 (1988); Stein and Cohen, Cancer Res., 48: 2659-2668 (1988); Stevenson and Inversen, J. Gen. Virol., 70: 2673-2682 (1989); Goodchild, "Inhibition of Gene Expression by Oligonucleotides," pp. 53-77, Oligodeoxynucleotides: Antisense Inhibitors of Gene Expression (Cohen, J.S., ed; CRC Press, Boca Raton, Florida,

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USA; 1989); Dervan et al., "Oligonucleotide Recognition of Double-helical DNA by Triple-helix Formation," pp. 197-210, id.; Neckers, L.M., "Antisense Oligodeoxynucleotides as a Tool for Studying Cell Regulation: Mechanisms of Uptake and Application to the Study of Oncogene Function," pp. 211-232, id.; Leitner et al., PNAS (USA), 87: 3430-3434 (1990); Bevilacqua et al., PNAS (USA), 85: 831-835 (1988); Loke et al. Curr. Top. Microbiol. Immunol., 141: 282-288 (1988); Sarin et al., PNAS (USA), 85: 7448-7451 (1988); Agrawal et al., "Antisense Oligonucleotides: A Possible Approach for Chemotherapy and AIDS," International Union of Biochemistry Conference on Nucleic Acid Therapeutics (Jan. 13-17, 1991; Clearwater Beach, Florida, USA); Armstrong, L., Ber. Week, pp. 88-89 (March 5, 1990); and Weintraub et al., Trends, 1: 22-25 (1985).] Such antisense nucleic acid sequences, preferably oligonucleotides, by hybridizing to the MN mRNA, particularly in the vicinity of the ribosome binding site and translation initiation point, inhibits translation of the mRNA. Thus, the use of such antisense nucleic acid sequences may be considered to be a form of cancer therapy.

Preferred antisense oligonucleotides according to this invention are gene-specific ODNs or oligonucleotides complementary to the 5' end of MN mRNA. Particularly preferred are the 29-mer ODN1 and 19-mer ODN2 [SEQ. ID. NOS.: 3 and 4]. Those antisense ODNs are representative of the many antisense nucleic acid sequences that can function to inhibit MN gene expression. Ones of ordinary skill in the art could determine appropriate antisense nucleic acid sequences, preferably antisense oligonucleotides, from the nucleic acid sequences of Figures 1 and 3a-d.

Also, as described above, CGL3 cells transfected with an "antisense" MN cDNA/promoter construct formed colonies much smaller than control CGL3 cells.

Vaccines

It will be readily appreciated that MN proteins and polypeptides of this invention can be incorporated into vaccines capable of inducing protective immunity against

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neoplastic disease and a dampening effect upon tumorigenic activity. Efficacy of a representative MN fusion protein GEX-3X-MN as a vaccine in a rat model is shown in Example 2.

5 MN proteins and/or polypeptides may be synthesized or prepared recombinantly or otherwise biologically, to comprise one or more amino acid sequences corresponding to one or more epitopes of the MN proteins either in monomeric or multimeric form. Those proteins and/or polypeptides may then be incorporated into vaccines capable of inducing protective
10 immunity. Techniques for enhancing the antigenicity of such polypeptides include incorporation into a multimeric structure, binding to a highly immunogenic protein carrier, for example, keyhole limpet hemocyanin (KLH), or diphtheria toxoid, and administration in combination with adjuvants or
15 any other enhancers of immune response.

Preferred MN proteins/polypeptides to be used in a vaccine according to this invention would be genetically engineered MN proteins. Preferred recombinant MN protein are the GEX-3X-MN, MN 20-19, MN-Fc and MN-PA proteins.

20 Other exemplary vaccines include vaccinia-MN (live vaccinia virus with full-length MN cDNA), and baculovirus-MN (full length MN cDNA inserted into baculovirus vector, e.g. in suspension of infected insect cells). Different vaccines may be combined and vaccination periods can be prolonged.

25 A preferred exemplary use of such a vaccine of this invention would be its administration to patients whose MN-carrying primary cancer had been surgically removed. The vaccine may induce active immunity in the patients and prevent recidivism or metastasis.

30 It will further be appreciated that anti-idiotypic antibodies to antibodies to MN proteins/polypeptides are also useful as vaccines and can be similarly formulated.

An amino acid sequence corresponding to an epitope of an MN protein/polypeptide either in monomeric or multimeric
35 form may also be obtained by chemical synthetic means or by purification from biological sources including genetically modified microorganisms or their culture media. [See Lerner, "Synthetic Vaccines", Sci. Am. 248(2): 66-74 (1983).] The

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protein/polypeptide may be combined in an amino acid sequence with other proteins/polypeptides including fragments of other proteins, as for example, when synthesized as a fusion protein, or linked to other antigenic or non-antigenic polypeptides of synthetic or biological origin. In some instances, it may be desirable to fuse a MN protein or polypeptide to an immunogenic and/or antigenic protein or polypeptide, for example, to stimulate efficacy of a MN-based vaccine.

10 The term "corresponding to an epitope of an MN protein/polypeptide" will be understood to include the practical possibility that, in some instances, amino acid sequence variations of a naturally occurring protein or polypeptide may be antigenic and confer protective immunity
15 against neoplastic disease and/or anti-tumorigenic effects. Possible sequence variations include, without limitation, amino acid substitutions, extensions, deletions, truncations, interpolations and combinations thereof. Such variations fall within the contemplated scope of the invention provided the
20 protein or polypeptide containing them is immunogenic and antibodies elicited by such a polypeptide or protein cross-react with naturally occurring MN proteins and polypeptides to a sufficient extent to provide protective immunity and/or anti-tumorigenic activity when administered as a vaccine.

25 Such vaccine compositions will be combined with a physiologically acceptable medium, including immunologically acceptable diluents and carriers as well as commonly employed adjuvants such as Freund's Complete Adjuvant, saponin, alum, and the like. Administration would be in immunologically
30 effective amounts of the MN proteins or polypeptides, preferably in quantities providing unit doses of from 0.01 to 10.0 micrograms of immunologically active MN protein and/or polypeptide per kilogram of the recipient's body weight. Total protective doses may range from 0.1 to about 100
35 micrograms of antigen. Routes of administration, antigen dose, number and frequency of injections are all matters of optimization within the scope of the ordinary skill in the art.

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The following examples are for purposes of illustration only and not meant to limit the invention in any way.

Example 1

5 Immunohistochemical Staining of Tissue Specimens

To study and evaluate the tissue distribution range and expression of MN proteins, the monoclonal antibody M75 was used to stain immunohistochemically a variety of human tissue specimens. The primary antibody used in these
10 immunohistochemical staining experiments was the M75 monoclonal antibody. A biotinylated second antibody and streptavidin-peroxidase were used to detect the M75 reactivity in sections of formalin-fixed, paraffin-embedded tissue samples. A commercially available amplification kit,
15 specifically the DAKO LSAB™ kit [DAKO Corp., Carpinteria, CA (USA)] which provides matched, ready made blocking reagent, secondary antibody and streptavidin-horseradish peroxidase was used in these experiments.

M75 immunoreactivity was tested according to the
20 methods of this invention in multiple-tissue sections of breast, colon, cervical, lung and normal tissues. Such multiple-tissue sections were cut from paraffin blocks of tissues called "sausages" that were purchased from the City of Hope [Duarte, CA (USA)]. Combined in such a multiple-tissue
25 section were normal, benign and malignant specimens of a given tissue; for example, about a score of tissue samples of breast cancers from different patients, a similar number of benign breast tissue samples, and normal breast tissue samples would be combined in one such multiple-breast-tissue section. The
30 normal multiple-tissue sections contained only normal tissues from various organs, for example, liver, spleen, lung, kidney, adrenal gland, brain, prostate, pancreas, thyroid, ovary, and testis.

Also screened for MN gene expression were multiple
35 individual specimens from cervical cancers, bladder cancers, renal cell cancers, and head and neck cancers. Such specimens were obtained from U.C. Davis Medical Center in Sacramento, CA

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and from Dr. Shu Y. Liao [Department of Pathology; St. Joseph Hospital; Orange, CA (USA)].

Controls used in these experiments were the cell lines CGL3 (H/F-T hybrid cells) and CGL1 (H/F-N hybrid cells) which are known to stain respectively, positively and negatively with the M75 monoclonal antibody. The M75 monoclonal antibody was diluted to a 1:5000 dilution wherein the diluent was either PBS [0.05 M phosphate buffered saline (0.15 M NaCl), pH 7.2-7.4] or PBS containing 1% protease-free BSA as a protein stabilizer.

Immunohistochemical Staining Protocol

The immunohistochemical staining protocol was followed according to the manufacturer's instructions for the DAKO LSABTM kit. In brief, the sections were dewaxed, rehydrated and blocked to remove non-specific reactivity as well as endogenous peroxidase activity. Each section was then incubated with dilutions of the M75 monoclonal antibody. After the unbound M75 was removed by rinsing the section, the section was sequentially reacted with a biotinylated antimouse IgG antibody and streptavidin conjugated to horseradish peroxidase; a rinsing step was included between those two reactions and after the second reaction. Following the last rinse, the antibody-enzyme complexes were detected by reaction with an insoluble chromogen (diaminobenzidine) and hydrogen peroxide. A positive result was indicated by the formation of an insoluble reddish-brown precipitate at the site of the primary antibody reaction. The sections were then rinsed, counterstained with hematoxylin, dehydrated and cover slipped. Then the sections were examined using standard light microscopy.

Interpretation. A deposit of a reddish brown precipitate over the plasma membrane was taken as evidence that the M75 antibody had bound to a MN antigen in the tissue. The known positive control (CGL3) had to be stained to validate the assay. Section thickness was taken into consideration to compare staining intensities, as thicker

sections produce greater staining intensity independently of other assay parameters.

Results

Preliminary examination of cervical specimens showed
5 that 62 of 68 squamous cell carcinoma specimens (91.2%)
stained positively with M75. Additionally, 2 of 6
adenocarcinomas and 2 of 2 adenosquamous cancers of the cervix
also stained positively. In early studies, 55.6% (10 of 18)
of cervical dysplasias stained positively. A total of 9
10 specimens including both cervical dysplasias and tumors,
exhibited some MN expression in normal appearing areas of the
endocervical glandular epithelium, usually at the basal layer.
In some specimens, whereas morphologically normal-looking
areas showed expression of MN antigen, areas exhibiting
15 dysplasia and/or malignancy did not show MN expression.

M75 positive immunoreactivity was most often
localized to the plasma membrane of cells, with the most
apparent stain being present at the junctions between adjacent
cells. Cytoplasmic staining was also evident in some cells;
20 however, plasma membrane staining was most often used as the
main criterion of positivity.

M75 positive cells tended to be near areas showing
keratin differentiation in cervical specimens. In some
specimens, positive staining cells were located in the center
25 of nests of non-staining cells. Often, there was very little,
if any, obvious morphological difference between staining
cells and non-staining cells. In some specimens, the positive
staining cells were associated with adjacent areas of
necrosis.

30 In most of the squamous cell carcinomas of the
cervix, the M75 immunoreactivity was focal in distribution,
i.e., only certain areas of the specimen stained. Although
the distribution of positive reactivity within a given
specimen was rather sporadic, the intensity of the reactivity
35 was usually very strong. In most of the adenocarcinomas of
the cervix, the staining pattern was more homogeneous, with
the majority of the specimen staining positively.

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Among the normal tissue samples, intense, positive and specific M75 immunoreactivity was observed only in normal stomach tissues, with diminishing reactivity in the small intestine, appendix and colon. No other normal tissue stained extensively positively for M75. Occasionally, however, foci of intensely staining cells were observed in normal intestine samples (usually at the base of the crypts) or were sometimes seen in morphologically normal appearing areas of the epithelium of cervical specimens exhibiting dysplasia and/or malignancy. In such, normal appearing areas of cervical specimens, positive staining was seen in focal areas of the basal layer of the ectocervical epithelium or in the basal layer of endocervical glandular epithelium. In one normal specimen of human skin, cytoplasmic MN staining was observed in the basal layer. The basal layers of these epithelia are usually areas of proliferation, suggesting the MN expression may be involved in cellular growth. In a few cervical biopsied specimens, MN positivity was observed in the morphologically normal appearing stratified squamous epithelium, sometimes associated with cells undergoing koilocytic changes.

Some colon adenomas (4 of 11) and adenocarcinomas (9 of 15) were positively stained. One normal colon specimen was positive at the base of the crypts. Of 15 colon cancer specimens, 4 adenocarcinomas and 5 metastatic lesions were MN positive. Fewer malignant breast cancers (3 of 25) and ovarian cancer specimens (3 of 15) were positively stained. Of 4 head and neck cancers, 3 stained very intensely with M75.

Although normal stomach tissue was routinely positive, 4 adenocarcinomas of the stomach were MN negative. Of 3 bladder cancer specimens (1 adenocarcinoma, 1 non-papillary transitional cell carcinoma, and 1 squamous cell carcinoma), only the squamous cell carcinoma was MN positive. Approximately 40% (12 of 30) of lung cancer specimens were positive; 2 of 4 undifferentiated carcinomas; 3 of 8 adenocarcinomas; 2 of 8 oat cell carcinomas; and, 5 of 10 squamous cell carcinomas. One hundred percent (4 of 4) of the renal cell carcinomas were MN positive.

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In summary, MN antigen, as detected by M75 and immunohistochemistry in the experiments described above, was shown to be prevalent in tumor cells, most notably in tissues of cervical cancers. MN antigen was also found in some cells of normal tissues, and sometimes in morphologically normal appearing areas of specimens exhibiting dysplasia and/or malignancy. However, MN is not usually extensively expressed in most normal tissues, except for stomach tissues where it is extensively expressed and in the tissues of the lower gastrointestinal tract where it is less extensively expressed. MN expression is most often localized to the cellular plasma membrane of tumor cells and may play a role in intercellular communication or cell adhesion. Representative results of experiments performed as described above are tabulated in Table 2.

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TABLE 2
Immunoreactivity of M75 in Various Tissues

	<u>TISSUE</u>	<u>TYPE</u>	<u>POS/NEG</u> <u>(#pos/#tested)</u>
5	liver, spleen, lung, kidney, adrenal gland, brain, prostate, pancreas, thyroid, ovary, testis	normal	NEG (all)
10	skin	normal	POS (in basal layer) (1/1)
	stomach	normal	POS
	small intestine	normal	POS
	colon	normal	POS
	breast	normal	NEG (0/10)
15	cervix	normal	NEG (0/2)
	breast	benign	NEG (0/17)
	colon	benign	POS (4/11)
	cervix	benign	POS (10/18)
	breast	malignant	POS (3/25)
20	colon	malignant	POS (9/15)
	ovarian	malignant	POS (3/15)
	lung	malignant	POS (12/30)
	bladder	malignant	POS (1/3)
	head & neck	malignant	POS (3/4)
25	kidney	malignant	POS (4/4)
	stomach	malignant	NEG (0/4)
	cervix	malignant	POS (62/68)

The results recorded in this example indicate that the presence of MN proteins in a tissue sample from a patient may, in general, depending upon the tissue involved, be a marker signaling that a pre-neoplastic or neoplastic process is occurring. Thus, one may conclude from these results that diagnostic/prognostic methods that detect MN antigen may be particularly useful for screening patient samples for a number of cancers which can thereby be detected at a pre-neoplastic stage or at an early stage prior to obvious morphologic

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changes associated with dysplasia and/or malignancy being evident or being evident on a widespread basis.

Example 2

Vaccine -- Rat Model

5 As shown above in Example 7 of WO 93/18152
(International Publication Date: 16 September 1993), in some
rat tumors, for example, the XC tumor cell line (cells from a
rat rhabdomyosarcoma), a rat MN protein, related to human MN,
is expressed. Thus a model was afforded to study antitumor
10 immunity induced by experimental MN-based vaccines. The
following representative experiments were performed.

Nine- to eleven-day-old Wistar rats from several
families were randomized, injected intraperitoneally with 0.1
ml of either control rat sera (the C group) or with rat serum
15 against the MN fusion protein GEX-3X-MN (the IM group).
Simultaneously both groups were injected subcutaneously with
10⁶ XC tumor cells.

Four weeks later, the rats were sacrificed, and
their tumors weighed. The results are shown in Figure 2.
20 Each point on the graph represents a tumor from one rat. The
difference between the two groups -- C and IM -- was
significant by Mann-Whitney rank test ($U = 84$, $\alpha < 0.025$).
The results indicate that the IM group of baby rats developed
tumors about one-half the size of the controls, and 5 of the
25 18 passively immunized rats developed no tumor at all,
compared to 1 of 18 controls.

Example 3

Expression of Full-Length MN cDNA in NIH 3T3 Cells

The role of MN in the regulation of cell
30 proliferation was studied by expressing the full-length cDNA
in NIH 3T3 cells. That cell line was chosen since it had been
used successfully to demonstrate the phenotypic effect of a
number of proto-oncogenes [Weinberg, R.A., Cancer Res., 49:
3713 (1989); Hunter, T., Cell, 64: 249 (1991)]. Also, NIH
35 3T3 cells express no endogenous MN-related protein that is
detectable by Mab M75.

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The full length MN cDNA was obtained by ligation of the two cDNA clones using the unique BamHI site and subcloned from pBluescript into KpnI-SacI sites of the expression vector pSG5C. pSG5C was kindly provided by Dr. Richard Kettman
5 [Department of Molecular Biology, Faculty of Agricultural Sciences, B-5030 Gembloux, Belgium]. pSG5C was derived from pSG5 [Stratagene] by inserting a polylinker consisting of a sequence having several neighboring sites for the following restriction enzymes: EcoRI, XhoI, KpnI, BamHI, SacI, 3 times
10 TAG stop codon and BglII.

The recombinant pSG5C-MN plasmid was co-transfected in a 10:1 ratio (10 μ g : 1 μ g) with the pSV2neo plasmid [Southern and Berg, J. Mol. Appl. Genet., 1: 327 (1982)] which contains the neo gene as a selection marker. The co-
15 transfection was carried out by calcium phosphate precipitation method [Mammalian Transfection Kit; Stratagene] into NIH 3T3 cells plated a day before at a density of 1×10^5 per 60 mm dish. As a control, pSV2neo was co-transfected with empty pSG5C.

20 Transfected cells were cultured in DMEM medium supplemented with 10% FCS and 600 μ g ml⁻¹ of G418 [Gibco BRL] for 14 days. The G418-resistant cells were clonally selected, expanded and analysed for expression of the transfected cDNA by Western blotting using iodinated Mab M75.

25 For an estimation of cell proliferation, the clonal cell lines were plated in triplicates (2×10^4 cells/well) in 24-well plates and cultivated in DMEM with 10% FCS and 1% FCS, respectively. The medium was changed each day, and the cell number was counted using a hemacytometer.

30 To determine the DNA synthesis, the cells were plated in triplicate in 96-well plate at a density of 10^4 /well in DMEM with 10% FCS and allowed to attach overnight. Then the cells were labeled with ³H-thymidine for 24 hours, and the incorporated radioactivity was counted.

35 For the anchorage-independent growth assay, cells (2×10^4) were suspended in a 0.3% agar in DMEM containing 10% FCS and overlaid onto 0.5% agar medium in 60 mm dish. Colonies grown in soft agar were counted two weeks after plating.

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Several clonal cell lines constitutively expressing both 54 and 58 kd forms of MN protein in levels comparable to those found in LCMV-infected HeLa cells were obtained. Selected MN-positive clones and negative control cells (mock-
5 transfected with an empty pSG5C plasmid) were subjected to further analyses directed to the characterization of their phenotype and growth behavior.

The MN-expressing NIH 3T3 cells displayed spindle-shaped morphology, and increased refractility; they were less
10 adherent to the solid support and smaller in size. The control (mock transfected cells) had a flat morphology, similar to parental NIH 3T3 cells. In contrast to the control cells that were aligned and formed a monolayer with an ordered pattern, the cells expressing MN lost the capacity for growth
15 arrest and grew chaotically on top of one another. Correspondingly, the MN-expressing cells were able to reach significantly higher (more than 2x) saturation densities (Table 3) and were less dependent on growth factors than the control cells.

20 MN transfectants also showed faster doubling times (by 15%) and enhanced DNA synthesis (by 10%), as determined by the amount of [³H]-thymidine incorporated in comparison to control cells. Finally, NIH 3T3 cells expressing MN protein grew in soft agar. The diameter of colonies grown for 14 days
25 ranged from 0.1 to 0.5 mm; however, the cloning efficiency of MN transfectants was rather low (2.4%). Although that parameter of NIH 3T3 cells seems to be less affected by MN than by conventional oncogenes, all other data are consistent with the idea that MN plays a role in cell growth control.

30

Table 3

Growth Properties of NIH 3T3 Cells Expressing MN Protein

35

Transfected DNA	pSG5C/ pSV2neo	pSG5C-MN/ pSV2neo
Doubling time ^a (hours)	27.9 ± 0.5	24.1 ± 1.3

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Saturation density ^b (cells x 10 ⁴ /cm ²)	4.9 ± 0.2	11.4 ± 0.4
Cloning efficiency (%) ^c	< 0.01	2.4 ± 0.2

5

^aFor calculation of the doubling time, the proliferation rate of exponentially growing cells was used. ^bThe saturation cell density was derived from the cell number 4 days after reaching confluency. ^cColonies greater than 0.1 mm in diameter were
10 scored at day 14. Cloning efficiency was estimated as a percentage of colonies per number of cells plated, with correction for cell viability.

Example 4

Acceleration of G1 Transit and Decrease in Mitomycin C 15 Sensitivity Caused by MN Protein

For the experiments described in this example, the stable MN transfectants of NIH 3T3 cells generated as described in Example 3 were used. Four selected MN-positive clones and four control mock-transfected clones were either
20 used individually or in pools.

Flow cytometric analyses of asynchronous cell populations. Cells that had been grown in dense culture were plated at 1 x 10⁶ cells per 60 mm dish. Four days later, the cells were collected by trypsinization, washed, resuspended in
25 PBS, fixed by dropwise addition of 70% ethanol and stained by propidium iodine solution containing RNase. Analysis was performed by FACStar using DNA cell cycle analysis software [Becton Dickinson; Franklin Lakes, NJ (USA)].

Exponentially growing cells were plated at 5 x 10⁵
30 cells per 60 mm dish and analysed as above 2 days later. Forward light scatter was used for the analysis of relative cell sizes. The data were evaluated using Kolmogorov-Smirnov test [Young, J. Histochem. Cytochem., 25: 935 (1977)].

The flow cytometric analyses revealed that clonal
35 populations constitutively expressing MN protein showed a

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decreased percentage of cells in G1 phase and an increased percentage of cells in G2-M phases. Those differences were more striking in cell populations grown throughout three passages in high density cultures than in exponentially growing subconfluent cells. That observation supports the idea that MN protein has the capacity to perturb contact inhibition.

Also observed was a decrease in the size of MN expressing cells seen in both exponentially proliferating and high density cultures. It is possible that the MN-mediated acceleration of G1 transit is related to the above-noted shorter doubling time (by about 15%) of exponentially proliferating MN-expressing NIH 3T3 cells. Also, MN expressing cells displayed substantially higher saturation density and lower serum requirements than the control cells. Those facts suggest that MN-transfected cells had the capacity to continue to proliferate despite space limitations and diminished levels of serum growth factors, whereas the control cells were arrested in G1 phase.

Limiting conditions. The proliferation of MN-expressing and control cells was studied both in optimal and limiting conditions. Cells were plated at 2×10^4 per well of 24-well plate in DMEM with 10% FCS. The medium was changed at daily intervals until day 4 when confluency was reached, and the medium was no longer renewed. Viable cells were counted in a hemacytometer at appropriate times using trypan blue dye exclusion. The numbers of cells were plotted versus time wherein each plot point represents a mean value of triplicate determination.

The results showed that the proliferation of MN expressing and control cells was similar during the first phase when the medium was renewed daily, but that a big difference in the number of viable cells occurred after the medium was not renewed. More than half of the control cells were not able to withstand the unfavorable growth conditions. In contrast, the MN-expressing cells continued to proliferate even when exposed to increasing competition for nutrients and serum growth factors.

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Those results were supported also by flow cytometric analysis of serum starved cells grown for two days in medium containing 1% FCS. While 83% of control cells accumulated in G0-G1 phase (S = 5%, G2-M = 12%), expression of MN protein partially reversed the delay in G1 as indicated by cell cycle distribution of MN transfectants (G0-G1 = 65%, S = 10%, G2-M = 26%). The results of the above-described experiments suggest that MN protein might function to release the G1/S checkpoint and allow cells to proliferate under unfavorable conditions.

10 MMC. To test that assumption, unfavorable conditions were simulated by treating cells with the DNA damaging drug mitomycin C (MMC) and then following their proliferation and viability. The mechanism of action of MMC is thought to result from its intracellular activation and subsequent DNA alkylation and crosslinking [Yier and Szybalski, Science, 145: 55 (1964)]. Normally, cells respond to DNA damage by arrest of their cell cycle progression to repair defects and prevent acquisition of genomic instability. Large damage is accompanied by marked cytotoxicity. However, many studies [for example, Peters et al., Int. J. Cancer, 54: 450 (1993)] concern the emergence of drug resistant cells both in tumor cell populations and after the introduction of oncogenes into nontransformed cell lines.

25 The response of MN-transfected NIH 3T3 cells to increasing concentrations of MMC was determined by continuous [³H]-thymidine labeling. Cells were plated in 96-well microtiter plate concentration of 10⁴ per well and incubated overnight in DMEM with 10% FCS to attach. Then the growth medium was replaced with 100 µl of medium containing increasing concentrations of MMC from 1 µl/ml to 32 µg/ml. All the drug concentrations were tested in three replicate wells. After 5 hours of treatment, the MMC was removed, cells were washed with PBS and fresh growth medium without the drug was added. After overnight recovery, the fractions of cells that were actively participating in proliferation was determined by continuous 24-hr labeling with [³H]-thymidine. The incorporation by the treated cells was compared to that of the control, untreated cells, and the proliferating fractions

were considered as a percentage of the control's incorporation.

The viability of the treated cells was estimated three days later by a CellTiter 96 AQ Non-Radioactive Cell Proliferation Assay [Promega] which is based on the bio-reduction of methotrexate (MTX) into a water soluble formazan that absorbs light at 490 nm. The percentage of surviving cells was derived from the values of absorbance obtained after subtraction of background.

The control and MN-expressing NIH 3T3 cells showed remarkable differences in their responses to MMC. The sensitivity of the MN-transfected cells appeared considerably lower than the control's in both sections of the above-described experiments. The results suggested that the MN-transfected cells were able to override the negative growth signal mediated by MMC.

ATCC Deposits. The material listed below was deposited with the American Type Culture Collection (ATCC) at 12301 Parklawn Drive, Rockville, MD 20852 (USA). The deposits were made under the provisions of the Budapest Treaty on the International Recognition of Deposited Microorganisms for the Purposes of Patent Procedure and Regulations thereunder (Budapest Treaty). Maintenance of a viable culture is assured for thirty years from the date of deposit. The hybridomas and plasmids will be made available by the ATCC under the terms of the Budapest Treaty, and subject to an agreement between the Applicants and the ATCC which assures unrestricted availability of the deposited hybridomas and plasmids to the public upon the granting of patent from the instant application. Availability of the deposited strain is not to be construed as a license to practice the invention in contravention of the rights granted under the authority of any Government in accordance with its patent laws.

	<u>Hybridoma</u>	<u>Deposit Date</u>	<u>ATCC #</u>
35	VU-M75	September 17, 1992	HB 11128
	MN 12.2.2	June 9, 1994	HB 11647

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<u>Plasmid</u>	<u>Deposit Date</u>	<u>ATCC #</u>
A4a	June 6, 1995	97199
XE1	June 6, 1995	97200
XE3	June 6, 1995	97198

5 The description of the foregoing embodiments of the
invention have been presented for purposes of illustration and
description. They are not intended to be exhaustive or to
limit the invention to the precise form disclosed, and
obviously many modifications and variations are possible in
10 light of the above teachings. The embodiments were chosen and
described in order to explain the principles of the invention
and its practical application to enable thereby others skilled
in the art to utilize the invention in various embodiments and
with various modifications as are suited to the particular use
15 contemplated.

All references cited herein are hereby incorporated
by reference.

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CLAIMS

1. An isolated nucleic acid containing at least twenty-seven nucleotides wherein the nucleotide sequence for said nucleic acid is selected from the group consisting of:

- 5 (a) SEQ. ID. NOS.: 1, 5, and 27-49 and nucleotide sequences complementary to SEQ. ID. NOS.: 1, 5 and 27-49;
- (b) nucleotide sequences that hybridize under stringent hybridization conditions to one or more of the following nucleotide sequences: SEQ. ID. NOS.: 1, 5, and 27-
10 49 and the respective complements of SEQ. ID. NOS.: 1, 5 and 27-49; and
- (c) nucleotide sequences that differ from the nucleotide sequences of (a) and (b) in codon sequence due to the degeneracy of the genetic code.

- 15 2. An isolated nucleic acid according to Claim 1 wherein said nucleotide sequence is selected from the group consisting of:
- (a) SEQ. ID. NO.: 5 and its complement;
- (b) nucleotide sequences that hybridize under
20 stringent conditions to SEQ. ID. NO.: 5 or to its complement;
- (c) nucleotide sequences that differ from the nucleotide sequences of (a) or (b) in codon sequence due to the degeneracy of the genetic code.

3. An isolated nucleic acid containing at least
25 sixteen nucleotides wherein the nucleotide sequence therefor is selected from the group consisting of:

- (a) the MN nucleotide sequences contained in plasmids A4a, XE1 and XE3 which were deposited at the American Type Culture Collection (ATCC) in Rockville, Maryland in the
30 United State of America under the respective ATCC Nos. 97199, 97200, and 97198;
- (b) nucleotide sequences that hybridize under stringent conditions to the MN nucleotide sequences of (a);
and

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(c) nucleotide sequences that differ from the nucleotide sequences of (a) or (b) in codon sequence due to the degeneracy of the genetic code.

4. An isolated nucleic acid according to Claim 3
5 which functions as a polymerase chain reaction primer for MN nucleic acid sequences, and which is from 16 to about 50 nucleotides in length.

10 5. An isolated nucleic acid, containing at least fifty nucleotides, encoding an MN protein or polypeptide that is specifically bound either by monoclonal antibodies designated M75 secreted by the hybridoma VU-M75 deposited at the American Type Culture Collection (ATCC) in Rockville, Maryland in the United States of America under ATCC No. HB 11128, or by monoclonal antibodies designated MN12 secreted by
15 the hybridoma MN 12.2.2 deposited at the ATCC under ATCC No. 11647, or by both of said monoclonal antibodies.

20 6. An isolated nucleic acid which is operatively linked to an expression control sequence within a vector wherein said nucleic acid is selected from the group consisting of:

(a) SEQ. ID. NO.: 1 and its complement;

(b) nucleic acids that hybridize under stringent hybridization conditions to SEQ. ID. NO.: 1 or to its complement; and

25 (c) nucleic acids that differ from the nucleic acid sequences of (a) and (b) due to the degeneracy of the genetic code.

30 7. A unicellular host, which is either prokaryotic or eukaryotic, that is transformed or transfected with the isolated nucleic acid operatively linked to an expression control sequence in a vector according to Claim 6.

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8. A method of recombinantly producing an MN protein, MN fusion protein or MN polypeptide comprising the steps of:

- 5 (a) transforming a unicellular host with the isolated nucleic acid operatively linked to an expression control sequence in a vector according to Claim 6;
- (b) culturing said unicellular host so that said MN protein or polypeptide is expressed; and
- 10 (c) extracting and isolating said MN protein or polypeptide.

9. A recombinant nucleic acid encoding a fusion protein, that consists essentially of an MN protein or polypeptide and a non-MN protein or polypeptide, wherein the nucleotide sequence for the portion of the nucleic acid
15 encoding the MN protein or polypeptide is selected from the group consisting of:

- (a) SEQ. ID. NO.: 1;
 - (b) nucleotide sequences that hybridize under stringent conditions to SEQ. ID. NO.: 1 or to its complement;
20 and
 - (c) degenerate variants of SEQ. ID. NO.: 1 and of the nucleotide sequences of (b);
- wherein the nucleic acid encoding said MN protein or polypeptide contains at least twenty-nine nucleotides.

25 10. A method of detecting mutations in an isolated MN gene and/or fragment(s) thereof comprising the steps of:
amplifying one or more fragment(s) of said gene by the polymerase chain reaction (PCR); and
determining whether said one or more fragments
30 contain any mutations.

11. An MN protein, MN fusion protein or MN polypeptide, wherein said MN protein, MN polypeptide or the MN protein portion of said MN fusion protein is encoded by a nucleic acid of at least twenty-nine nucleotides which is
35 selected from the group consisting of:

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(a) SEQ. ID. NO.: 1;

(b) nucleotide sequences that hybridize under stringent hybridization conditions to SEQ. ID. NO.: 1 or to its complement; and

5 (c) nucleotide sequences that differ from SEQ. ID. NO.: 1 or from the sequences of (b) in codon sequence due to the degeneracy of the genetic code.

10 12. An MN protein, MN fusion protein, or MN polypeptide wherein said MN protein or polypeptide has, and wherein said MN fusion protein contains, an amino acid sequence selected from the group consisting of SEQ. ID. NOS.: 2, 6, 10-16, and 50-53.

15 13. A vaccine comprising an immunogenic amount of one or more MN proteins, MN fusion proteins, and/or MN polypeptides according to Claims 11 or 12 dispersed in a physiologically acceptable, nontoxic vehicle, which amount is effective to immunize a vertebrate against a neoplastic disease associated with expression of MN antigen.

20 14. An antibody which specifically binds to an MN protein, an MN fusion protein and/or an MN polypeptide according to Claims 11 or 12.

25 15. A monoclonal antibody according to Claim 14 which is designated MN12 and is secreted by the hybridoma MN 12.2.2 which was deposited at the American Type Culture Collection (ATCC) in Rockville, Maryland in the United States of America under ATCC No. HB 11647.

30 16. An antibody according to Claim 14 which specifically binds to an MN antigen epitope selected from the group of epitopes represented by the following amino acid sequences: SEQ. ID. NOS. 10-16.

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17. An antibody according to Claim 14 which is linked to an imaging agent, to a chemotherapeutic agent or to a toxic agent.

18. A method of imaging pre-neoplastic or
5 neoplastic disease in a patient comprising:

(a) injecting said patient with antibody linked to an imaging agent according to Claim 17; and

(b) detecting the binding of said antibody.

19. A hybridoma designated MN 12.2.2 which produces
10 the monoclonal antibody MN12, and which was deposited at the American Type Culture Collection (ATCC) in Rockville Maryland in the United States of America under ATCC Accession No. HB 11647.

20. A method of delivering a chemotherapeutic agent
15 or toxic agent to a cancer cell which comprises contacting said cell with an antibody linked to a chemotherapeutic agent or to a toxic agent according to Claim 17.

21. A method of treating neoplastic disease in a patient comprising administering to said patient a
20 therapeutically effective amount of a composition comprising antibodies which specifically bind to an MN protein, an MN fusion protein and/or an MN polypeptide according to Claims 11 or 12.

22. A method of detecting and/or quantitating in a
25 vertebrate sample MN antigen comprising the steps of:

(a) contacting said sample with one or more antibodies according to Claims 14, 15 or 16; and

(b) detecting and/or quantitating binding of said antibody in said sample.

23. A method according to Claim 22 wherein said
30 vertebrate sample is a human tissue specimen, such as, a cell

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smear, a histological section from a biopsied tissue or organ, or an imprint preparation.

24. A method according to Claim 23 wherein said tissue specimen is ovarian, endometrial, or cervical.

5 25. A method of detecting and/or quantitating MN-specific antibodies in a vertebrate sample comprising the steps of:

 (a) contacting and incubating the vertebrate sample with an MN protein, an MN fusion protein and/or an MN
10 polypeptide according to Claims 11 or 12; and

 (b) detecting and/or quantitating binding of said MN protein, MN fusion protein and/or MN polypeptide to antibody in said sample.

15 26. A method of treating neoplastic disease and/or pre-neoplastic disease comprising inhibiting the expression of MN genes by administering one or more antisense nucleic acid sequences that hybridize under stringent conditions to mRNA transcribed from MN genes.

20 27. Vectors containing an MN nucleic acid sequence wherein said MN nucleic acid sequence is selected from the group consisting of:

 (a) SEQ. ID. NO.: 5 and its complement;

 (b) nucleic acids that hybridize to SEQ. ID. NO.: 5 or to its complement; and

25 (c) nucleic acids that differ from the nucleic acids of (a) or (b) due to the degeneracy of the genetic code;

 wherein said nucleic acid is at least twenty-nine nucleotides in length.

30 28. Vectors containing an MN nucleic acid sequence according to Claim 27 selected from the group consisting of the plasmids A4a, XE1 and XE3 which are deposited at the American Type Culture Collection (ATCC) in Rockville, Maryland

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in the United States of America under the respective ATCC
accession numbers 97199, 97200 and 97198.

1 / 14

1	1	ACA	GTC	AGC	CGC	ATG	GCT	CCC	CTG	TGC	CCC	AGC	CCC	TGG	CTC	CCT	CTG	12	48
13	49	L	I	P	A	P	A	P	G	L	T	V	Q	L	L	L	S	28	96
29	97	L	L	L	L	M	P	V	H	P	Q	R	L	P	R	M	Q	44	144
45	145	E	D	S	P	L	G	G	G	S	S	G	E	D	D	P	L	60	192
61	193	G	E	E	D	L	P	S	E	E	D	S	P	R	E	E	D	76	240
77	241	P	P	G	E	E	D	L	P	G	E	E	D	L	P	G	E	92	288
93	289	E	D	L	P	E	V	K	P	K	S	E	E	E	G	S	L	108	336
109	337	K	L	E	D	L	P	T	V	E	A	P	G	D	P	Q	E	124	384
125	385	P	Q	N	A	H	R	D	K	E	G	D	D	D	Q	S	H	140	432
141	433	W	R	Y	G	D	P	P	P	W	P	R	V	S	P	A	C	156	480
157	481	A	G	R	F	Q	S	P	V	D	I	R	P	Q	L	A	A	172	528

FIG.-1A

SUBSTITUTE SHEET (RULE 26)

FIG. 1B

SUBSTITUTE SHEET (RULE 26)

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349   V   M   L   S   A   K   Q   L   H   T   L   S   D   T   L   W   364
1057 GTG ATG CTG AGT GCT AAG CAG CTC CAC ACC CTC TCT GAC ACC CTG TGG 1104

365   G   P   G   D   S   R   L   Q   L   N   F   R   A   T   Q   P   380
1105 GGA CCT GGT GAC TCT CGG CTA CAG CTG AAC TTC CGA GCG ACG CAG CCT 1152

381   L   N   G   R   V   I   E   A   S   F   P   A   G   V   D   S   396
1153 TTG AAT GGG CGA GTG ATT GAG GCC TCC TTC CCT GCT GGA GTG GAC AGC 1200

397   S   P   R   A   A   E   P   V   Q   L   N   S   C   L   A   A   412
1201 AGT CCT CGG GCT GCT GAG CCA GTC CAG CTG AAT TCC TGC CTG GCT GCT 1248

413   G   D   I   L   A   L   V   F   G   L   L   F   A   V   T   S   428
1249 GGT GAC ATC CTA GCC CTG GTT TTT GGC CTC CTT TTT GCT GTC ACC AGC 1296

429   V   A   F   L   V   Q   M   R   R   Q   H   R   R   G   T   K   444
1297 GTC GCG TTC CTT GTG CAG ATG AGA AGG CAG CAC AGA AGG GGA ACC AAA 1344

445   G   G   V   S   Y   R   P   A   E   V   A   E   T   G   A   *   460
1345 GGG GGT GTG AGC TAC CGC CCA GCA GAG GTA GCC GAG ACT GGA GCC TAG 1392

1393 AGG CTG GAT CTT GGA GAA TGT GAG AAG CCA GCC AGA GGC ATC TGA GGG 1440

1441 GGA GCC GGT AAC TGT CCT GTC CTC ATT ATG CCA CTT CCT TTT AAC 1488

1489 TGC CAA GAA ATT TTT TAA AAT AAA TAT TTA TAA T 1522

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FIG._1C

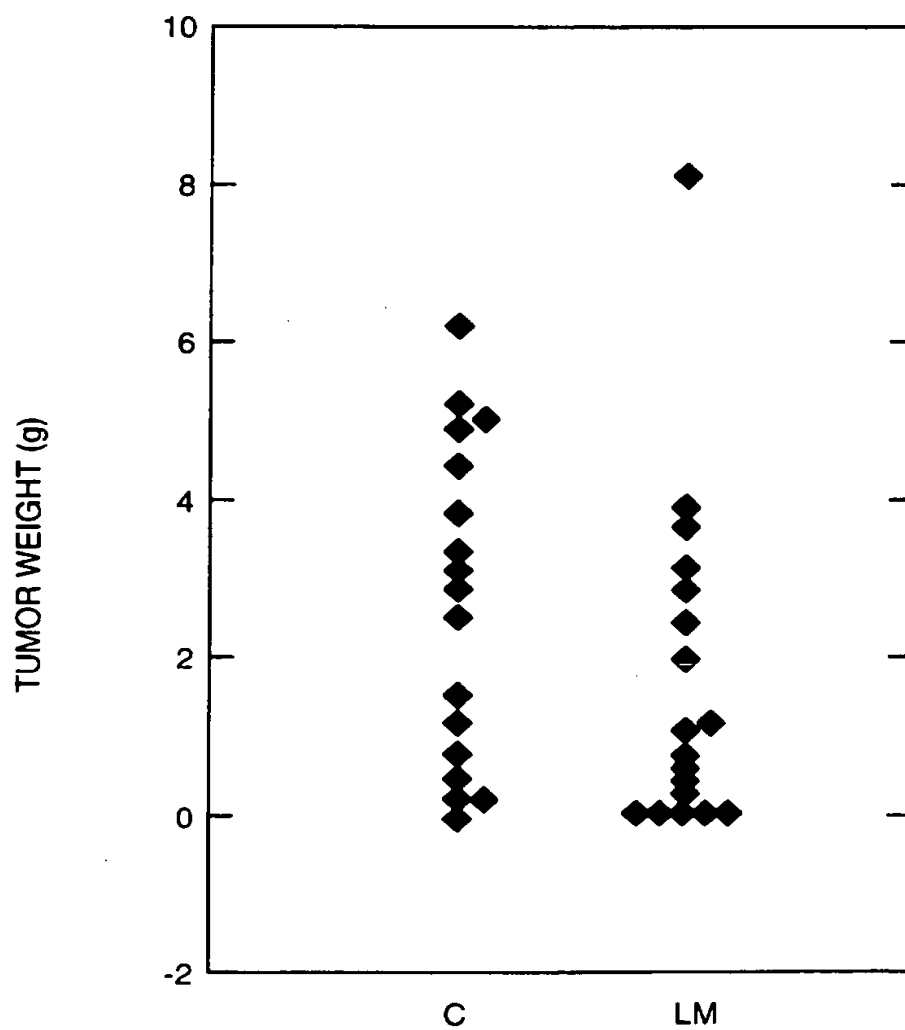
FIG._1A

FIG._1B

FIG._1C

FIG._1

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**FIG. 2****SUBSTITUTE SHEET (RULE 26)**

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FIG.-3A

1 ggatcctgtt gactcgtgac cttaccccca accctgtgct ctctgaacaa tgagctgtgt
61 ccactcaggg ttaaatggat taaggcggt gcaagatgtg ctttgttaaa cagatgcttg
121 aaggcagcat gctcgttaag agtcatcacc aatccctaata ctcaagtaaa caggacaca
181 aacactcgg aaggcgcag ggtcctctgc ctaggaaaac cagagacctt tgttcaactg
241 ttatctgac ctccctcca ctattgtcca tgacctgccc aaatccccct ctgtgagaaa
301 caccacaagaa ttatcaataa aaaaataaat ttaaaaaaaa aatacaaaa aaaaaaaa
361 aaaaaaaa gacttacgaa tagttattga taaatgaata gctattggta aagccaagta
421 aatgatcata ttcaaaaacca gacggccatc atcacagctc aagctacctt gatttgatct
481 ctttatcatt gtcattcttt gatttacctt gattagtcac catcctcaaa attctcccc
541 aagttctaata tacgttccaa acatttaggg gttacatgaa gcttgaacct actaccttct
601 ttgcttttga gccatgagtt gtaggaatga tgagtttaca ccttacctgc tgggatttaa
661 tttaaaacttt acctctaagt cagttgggta gcctttggct tatttttgta gctaatttg
721 tagttaatgg atgcaactgt aatcttgcta tgatagtttt cctccacct ttgccactag
781 gggtaggtag gtactcagtt ttccagtaatt gcttacctaa gacctaaag cctatttctc
841 ttgtactggc ctttatctgt aatatgggca tatttaatac aatataattt ttggagtttt
901 ttgtttgtt tggtgacctc tgggacctc gcaagctcca cctcccgagt tcacggcatt
961 ggagtagcag tgggacctc gtagctggga ctacaggcg cggccacct gccgggctaa
1021 ttccctgcctc agcctcccga gtagctggga caccgtgtta gccagaatgg tctcgatctc
1081 ttttttgtat ttttggtaga gacggggttt ccaaagttct gggattacag gtgtgagcca
1141 ctgacttcgt gatccaccg cctcgccctc aagtaaaaat agtcttcta agctggtaac
1201 ccgcacctgg ccaatttttt gagtctttta aagtaaaaat atgtcttcta agctggtaac
1261 tatggtacat ttccttttat taatgtgggtg ctgacgggtca tataggttct tttgagtttg
1321 gcatgcataat gctacttttt gcagtccttt cattacattt ttctctcttc atttgaagag
1381 catgttataat ctttttagctt cacttggctt aaaagggttct ctcatagcc taacacagt
1441 tcattgttgg taccacttgg atcataagt ttcagggtgaa tctgacacta agaaactccc ctacctgagg
1501 cttgttttga agaggatga ttcagggtgaa ttcagggtgaa ttcagggtgaa ttcagggtgaa
1561 tctgagattc ctctgacatt gctgtatata ggtttttctt ttgacagcct gtgactgagg
1621 actatttttc ttaagcaaga tatgtctaaag ttttgtgagc ctttttccag agagaggtct
1681 catatctgca tcaagtgaaga acatataatg tctgcatgtt tccatatctc agaatgttt
1741 gcttgtgttt tatgttttta tatagacagg gaaacttgtt cctcagtgac ccaaaaagagg
1801 tgggaattgt tattggatat catcatggc ccacgctttc tgaccttggg aacaattaa
1861 ggttcataat ctcaattctg tcagaattgg tacaagaaat agctgctatg tttcttgaca
1921 ttccacttgg taggaaataa gaatgtgaaa ctcttcagtt ggtgtgtgtc cct?gtttt

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FIG.-3B

1981 ttgcaatttc cttcttactg tgttaaaaa aagtatgata ttgctctgag aggtgaggca
2041 ttcttaataca tgatctttaa agatcaataa tataatcctt tcaaggatta tgtctttatt
2101 ataataaaga taatttgtct ttaacagaat caataataa atccctaaa ggattatatac
2161 tttgctgggc gcagtggctc acacctgtaa tcccagcaat ttgggtggcc aaggtggaag
2221 gatcaaatat gatcaaatat gcctacttct atattatctt ctaaacgaga attcatctct cttccctcaa
2281 tatgatgata ttgacagggg ttgcccctac tcactagatt gtgagctcct gctcagggca
2341 ggtagcggtt tttgtttttg tttttgtttt ttttttttga gacaggggtct tgtctgttca
2401 ccaggccag agtgcaatgg tacagtctca gctcactgca gcctcaaccg cctcgggtca
2461 aaccatcatc ccatttcagc ctccctgagta gctgggacta caggcacatg ccattacacc
2521 tggctaattt ttttgtattt ctagttaga caggggtttg ccatgttgcc cgggctggtc
2581 tcgaactcct ggactcaagc aatccaccca cctcagcctc ccaaatgag ggaccgtgtc
2641 ttatttcatt ccattgtcct gaatgcaata gtaaatagca ttccagggag caagaactag attaacaag
2701 aatatttgtt gtggtaaaaa gtttggagaa aaaaataata gtttaatttg gctagagtat gagggagagt
2761 gtggtaaaaa agtaggagac aagatggaaa ggtctcttgg gcaagggttt gaaggaagt ggaagtcaga
2821 agtacacaat gtgcatactg ttgaaaaata aatataggtt aaacctatca gagccccctt gacacataca
2941 gagtaattgt ttgaaaaata aagtttgtct cccacatacc cattacttaa ctacacctcg
3001 ctgtcttttc atccaagctc gcagcctgcc ctacctctt acctgcttcc tgggtgagtc aggatgtat
3061 ggctccccca acatgagctg ctttccctct cagccagagg acatgggggg cccagctcc cctgccttc
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3181 ccttctgtg cctggagctg ccaggagag cctgcatagt gccaggtgtt gccttgggtt ccaagctagt
3241 ctgggtggg gataaccttc tgccctgtgca cagacctgccc cctcactcca ccccatcct
3301 ccattggccc agctttggta tgggggagag ggcagcctgt gactcagcct gctccccctc cccccccc
3361 agctttggta tgggggagag ggcagcctgt gactcagcct gactcagcct gactcagcct cccccccc
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3481 cagctctcgt ttccaatgca cgtacagccc gtaacacccg tgtgctggga cccccacag
3541 TCAGCCGCAT GGCTCCCCCTG TGCCCCAGCC CCTGGCTCCC TCTGTTGATC CCGGCCCTG
3601 CTCCAGGCCT CACTGTGCAA CTGCTGCTGT CACTGCTGCT TCTGCTGCT GTCCATCCCC
3661 AGAGGTTGCC CCGGATGCCAG GAGGATTCCC CCTTGGGAGG AGGCTCTTCT GGGGAAGATG
3721 ACCCACTGGG CGAGGAGGAT CTGCCAGTG AGAGGATTC ACCCAGAGAG GAGGATCCAC
3781 CCGGAGAGGA GGATCTACCT GGAGAGGAGG ATCTACCTGG AGAGGAGGAT CTACCTGAAG
3841 TTAAGCCTAA ATCAGAAGAA GAGGGCTCCC TGAAGTTAGA GGATCTACCT ACTGTTGAGG
3901 CTCCTGGAGA TCCTCAAGAA CCCCAGAATA ATGCCACAG GGACAAAGAA Ggtaagtggc

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FIG._3C

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4081 tcccatacca atatcccat cccactctc ggaggtagaa agggacagat gtggagagaa
4141 aataaaaaagg gtgcaaaagg agagaggtga gctggatgag atgggagaga agggggaggc
4201 tggagaagag aaaggatga gaactgcaga tgagagaaaa aatgtgcaga cagaggaaaa
4261 aatataggtgg agaaggagag tcagagagtt tgaggggaaag agaaaaaggaa agcttgggag
4321 gtgaagtggg taccagagac aagcaagaag agctggtaga agtcattctca tcttaggcta
4381 caatgaggaa ttgagacctt ggaagaaggg acacagcagg tagagaaacg tggcttcttg
4441 actcccaagc caggaatttg gggaaaaggg ttggagacca tacaaggcag agggatgagt
4501 ggggagaaga aagaaggagg aaaggaaga tgggtactc actcatttg gactcaggac
4561 tgaagtgcc actcactttt ttttttttt tttttgagac aaactttcac tttgttgcc
4621 caggctggag tgcaatggcg cgatctcggc tcactgcaac ctccacctcc cgggttcaag
4681 tgattctcct gcctcagcct ctagccaagt agctgcgatt acaggcatgc gccaccacgc
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4861 cgtgagccac agcgctggc ctgaagcagc cactcactt tacagaccct aagacaatga
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4981 tctcctgtgc ttgacactg gcccgcttaa ggcatttgtt accgtaatg ctctgtaaag
5041 gcatctgcgt ttgtgacatc gttttggtcg ccaggaaggg atbggggctc taagcttgag
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5221 cgtccctgaa cactggtccc gggcgtecca cccgcgccc accgtcccac cccctcacct
5281 tttctaccg ggttccctaa gttcctgacc taggcgtcag acttctcac tatactctcc
5341 caccacagc GACCGCCCTT GGCCCGGGT GTCCCCAGCC TGCGCGGGCC GCTTCCAGTC
5401 CCCGGTGGAT ATCCGCCCCC AGCTCGCCGC CTCTGCCCCG GCCCTGCGCC CCTTGAACCT
5461 CCTGGGCTTC CAGCTCCCCG CGTCCCCAGA ACTGCGCCTG CGCAACAATG GCCACAGTGg
5521 tgagggggtc tccccgcga gacttgggga tggggcgggg cgcagggaaag ggaaccgtcg
5581 cgcagtgcct gcccggggt tgggctggcc ctaccggcg gggccggctc acttgcctct
5641 ccctacgcag TGCAACTGAC CCTGCCCTCTT GGGCTAGAGA TGGCTCTGGG TCCCCGGCGG
5701 GAGTACCGG CTCTGCAGCT GCATCTGCAC TGGGGGGCTG CAGGTCGTCC GGGCTCGGAG
5761 CACACTGTGG AAGGCCACCG TTTCCCTGCC GAGgtgagcg cggactggcc gagaaggggc
5821 aaaggagcgg ggcggacggg ggcagagac gtggccctct cctaccctcg tgccttttc
5881 agATCCACGT GGTTCACCTC AGCACCGCCT TTGCCAGAGT TGACGAGGCC TTGGGGCGCC

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5941 CGGAGGCCCT GCGCGTGTG GCCCCTTTC TGGAGgtacc agatcctgga caccacctac
6001 tccccgctt cccatcccat gctcctccc gactctatcg tggagccaga gaccacctcc
6061 cagcaagctc actcaggccc ctggctgaca aactcattca cgcactgttt gttcatttaa
6121 caccactgt gaaccaggca ccagcccca ccaaggattc tgaagctgta ggtccttgcc
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6301 aggtgttcat tgcagaggaa acagaatgtg caaagactca gaatatggcc tatttaggga
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6481 ccaggctgga gtgcagtgt gtgatcttgg gtcactgcaa cttccgcctc ccgggttcaa
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6721 tgagccaccg tgcacagcca cactcactga tctttaaag ccagccacac agcacaaagt
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6841 cttaacatta ggttcataag caaataaaga aaaaagaata ataaataaaa gaagtggcat
6901 gtcaggacct cactgaaaa cccaaacaca gaatcatgaa ggtgaatgca gaggtgacac
6961 caacacaaag gtgtatatat ggtttcctgt ggggagtagt tacggaggca gcagtgaagt
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7141 cagcaagagt acatagagt tgaataaata cataggattt taagaggag acactgtctc
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7261 agcatctca gagctgagga atgggagagg actatgggaa ccccttcat gttccggcct
7321 tcagccatgg ccctggatac atgcactcat ctgtcttaca atgtcattcc ccagGAGGG
7381 CCCGGAAGAA AACAGTGCC TATGAGCAGTT GCTGTCTCGC TTGGAAGAAA TCGCTGAGGA
7441 AGgtcagttt gttggtctgg ccactaatct ctgtggccta gttcataaag aatcacctt
7501 tggagcttca ggtctgaggc tccctccag tgcaggaggg attgaagcat
7561 gagccagcgc tcatcttgat aataaccatg aagctgacag acacagttac ccgcaaacgg
7621 ctgcctacag attgaaaacc aagcaaaaac cgccgggcac ggtggctcac gcctgtaac
7681 ccagcacttt gggaggccaa ggcagggtgga tcacgaggtc aagagatcaa gaccatcctg
7741 gccaacatgg tgaaacccca tcttactaa aaatacgaaa aaatagccag gcgtggtggc
7801 ggggtgcctgt aatcccagct actcgggagg ctgaggcagg agaattggcat gaaccggga
7861 ggcagaagtt gcagtgagcc gagatcgtgc cactgcactc cagcctgggc aacagagcga

FIG. 3D

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FIG. 3E

7921 gactcttgct tcaaaaaaa gataaaaaa gaaaccaa gaaaccaa aatgagacaa
7981 aaaaaaag accaaaaaat ggtgtttgga aatgtcaag gtcaagctctg gagagctaaa
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8461 atggtacaca gagttaagag tgtagactca gacgggtctt cttctttcct tctcttcct
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8581 caggcctctt ccagttgctc caagccctg tactttttt ttgagtaacg tcttatggga
8641 agggcctgca cttagtgaag aagtgtctc aagtgtgagt taccttggt tctgggaggt
8701 gaaactgtat ccctataacc tgaagcttta aggggtgca atgtagatga gaccccaaca
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8881 CAGGGTGTCA TCTGGACTGT GTTTAACCCAG ACAGTGATGC TGAGTGCTAA GCAGgtggc
8941 ctggggtgtg tgtggacaca gtgggtgctg gggaaagag atgtaagatg agatgagaaa
9001 caggagaaga aagaaataca ggctgggctc tgtggttac gcctataatc ccaccagtt
9061 gggaggctga ggtggagaa tgggttgagc ccaggagttc aagacaaggc ggggcaacat
9121 agtgtgacct catctctacc aaaaaacc caacaaacc aaaaatagcc gggcatggtg
9181 gtatcgggcc tagtcccagc tactcaagga ggctgaggtg ggaagatcgc ttgattccag
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9301 atttatttat aaagaaatc aagaggctg atggggaata caggagctgg aggtggagc
9361 cctgaggtgc tggttgtgag ctggcctggg acccttgttt cctgtcatgc catgaaccca
9421 ccacactgt ccactgacct ccctagCTCC ACACCCTCTC TGACACCCCTG TGGGACCTG
9481 GTGACTCTCG GCTACAGCTG AACTTCCGAG CGACGCAGCC TTTGAATGGG CGAGTGATTG
9541 AGGCTCTCTT CCTGCTGGA GTGGACAGCA GTCTCGGGC TGCTGAGCCA Ggtacagctt
9601 tgtctggttt cccccagcc agtagtccct tatcctccca tgtgtgtgct agtgtctgtc
9661 attggtggtc acagccgccc tctcacatct cttttttctc tccagTCCAG CTGAATTCTT
9721 GCCTGGCTGC TGGtgagttt gccctctctc ttggtcctga tggcaggaga ctcctcagca
9781 ccattcagcc ccagggtgc tcaggaccgc ctctgctccc tctcctttc tgcagaacag
9841 acccaacc caatattaga gaggcagatc atggtgggga tcccccat gtccccagag

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9901 gctaattgat tagaatgaag cttgagaaat ctcccagcat ccctctcgca aaagaatccc
9961 cccccctttt tttaaagata gggctctcact ctgtttgccc caggctgggg tgttgtggca
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10501 atgcaaatga gctgctcctg ggccagtttt ctgattagcc ttctctgttg tgtacacaca
10561 GAAGGGGAAC CAAAGGGGT GTGAGCTACC GCCCAGCAGA GGTAGCCGAG ACTGGAGCCT
10621 AGAGGCTGGA TCTTGGAGAA TGTGAGAAGC CAGCCAGAGG CATCTGAGGG GGAGCCGGTA
10681 ACTGTCTGT CTTGCTCATT ATGCCACTTC CTTTAACTG CCAAGAAATT TTTTAAATA
10741 AATATTATA ATaaaatatg tgtagtcac ctttgttccc caaatcagaa ggaggatat
10801 gaatttccta ttactgttat tagcaccaat ttagtggtaa tgcatttatt ctattacagt
10861 tcggcctcct tccacacatc actccaatgt gttgctcc

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FIG._3F**FIG._3A****FIG._3B****FIG._3C****FIG._3D****FIG._3E****FIG._3F****FIG._3**



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-506 CTTGCTTTTC ATTCAAGCTC AAGTTTGTTCT CCCACATACC CATTACTTAA CTCACCCCTCG
-446 GGCTCCCCTA GCAGCCTGCC CTACCTCTTT ACCTGCTTCC TGGTGGAGTC AGGGATGTAT
AP2
-386 ACATGAGCTG CTTTCCCTCT CAGCCAGAGG ACATGGGGG CCCCAGCTCC CCTGCCCTTTC
-326 CCCTTCTGTG CCTGGAGCTG GGAAGCAGGC CAGGGTTAGC TGAGGCTGGC TGGCAAGCAG
-266 CTGGGTGGTG CCAGGGAGAG CCTGCATAGT GCCAGGTGGT GCCTTGGGTT CCAAGCTAGT
p53
-206 CCATGGCCCC GATAACCTTC TGCCTGTGCA CACACCTGCC CCTCACTCCA CCCCCATCCT
Inr
-146 AGCTTTGGTA TGGGGGAGAG GGCACAGGGC CAGACAAACC TGTGAGACTT TGGCTCCATC
Inr
-86 TCTGCAAAG GCGGCTCTGT GAGTCAGCCT GCTCCCCCTCC AGGCTTGCTC CTCCCCCACC
AP1 p53 AP2
-26 CAGCTCTCGT TTCCAATGCA CGTACAGCCC GTACACACCG TGTGCTGGGA CACCCACAG

FIG._6

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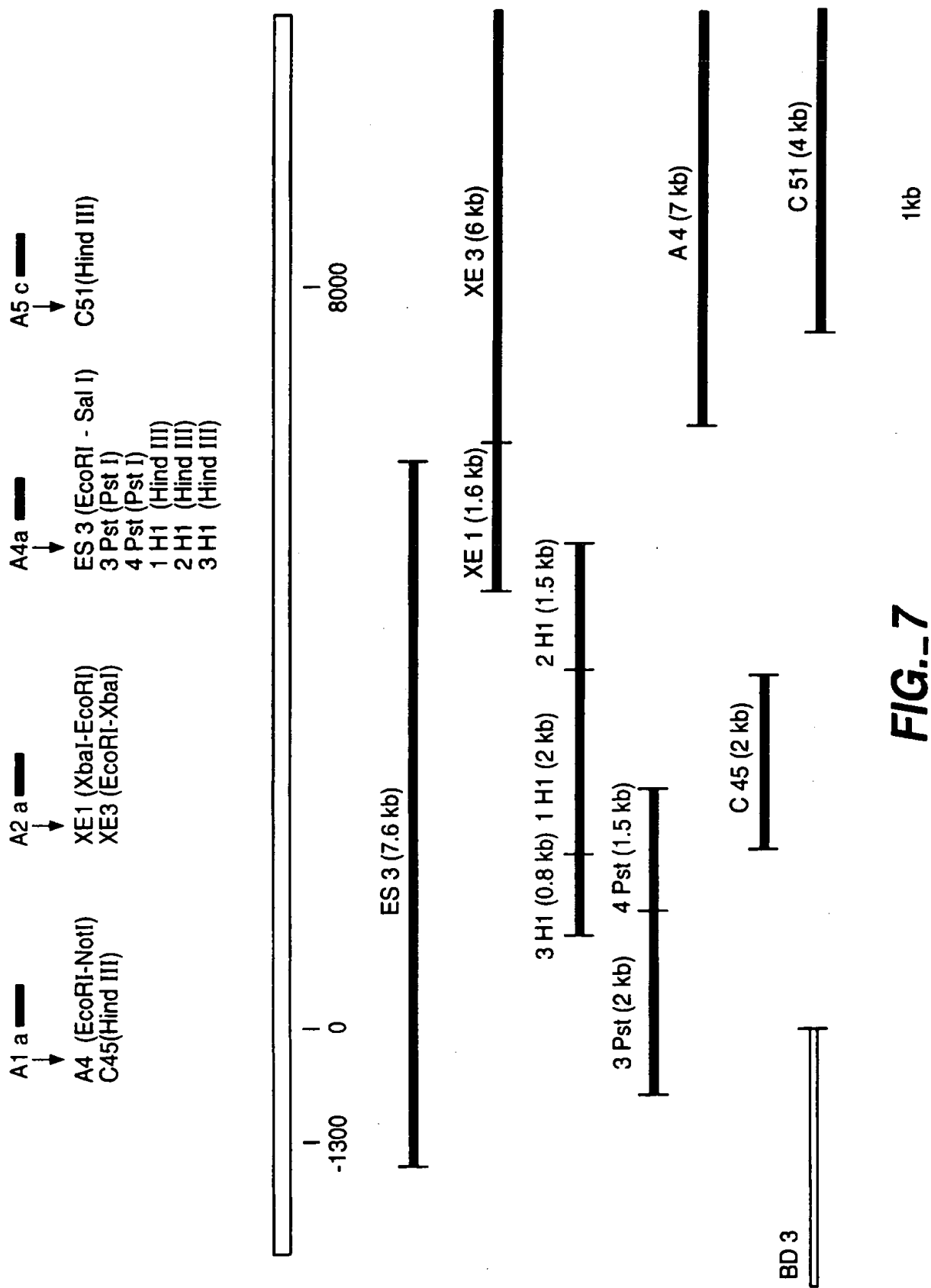
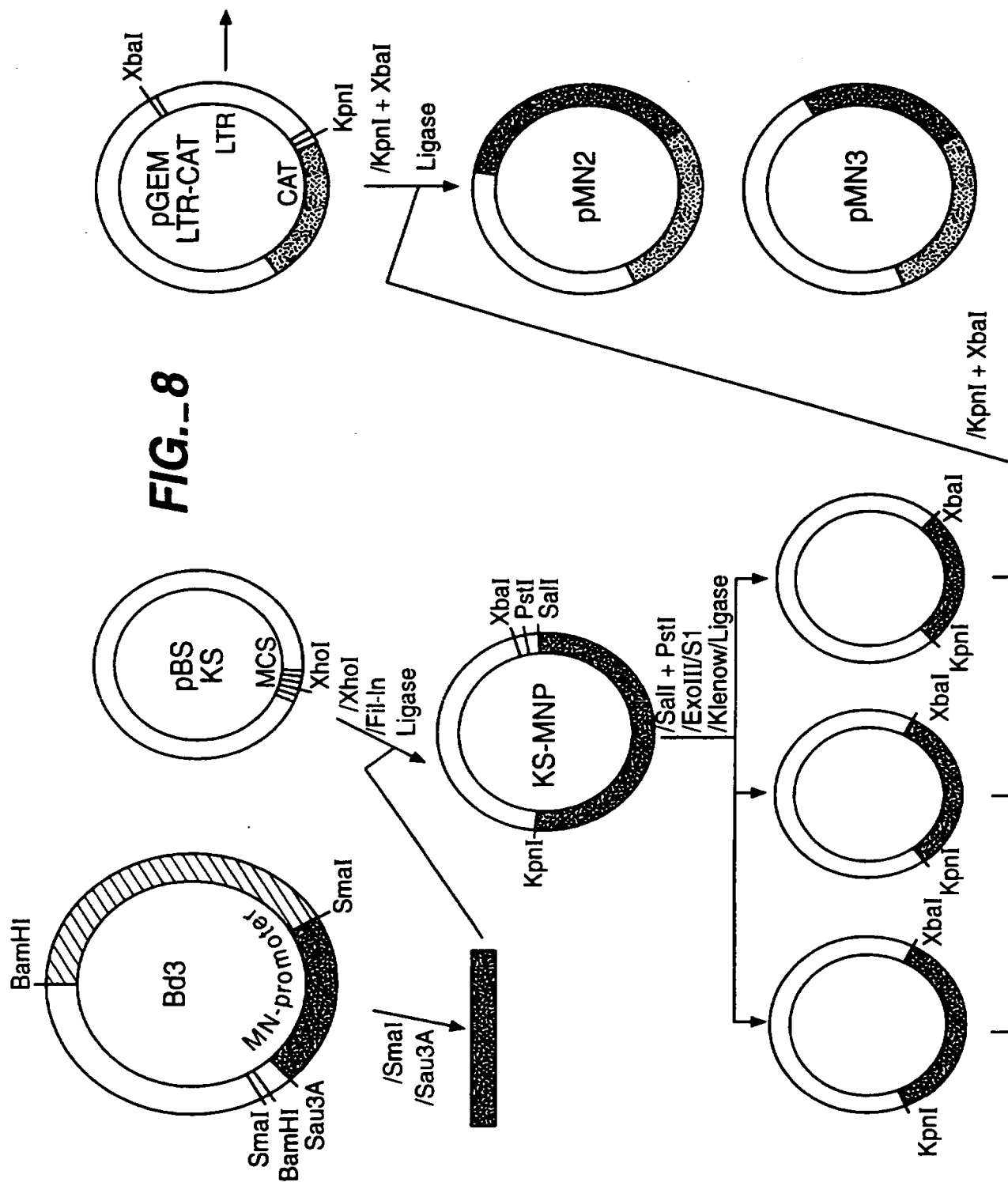


FIG. 7

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INTERNATIONAL SEARCH REPORT

International Application No.

PCT/US 95/07628

A. CLASSIFICATION OF SUBJECT MATTER

IPC 6 C12N15/12 C12N1/21 C12N15/62 C12Q1/68 A61K39/00
 C07K14/82 C12N5/16 C12N15/87 C07K16/30 G01N33/50
 A61K48/00

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 C12N A61K C07K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO,A,93 18152 (CIBA CORNING DIAGNOSTICS CORP. AND INSTITUTE OF VIROLOGY) 16 September 1993 cited in the application see the whole document ---	1-14, 16-18, 21-27
X	PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF USA, vol. 85, December 1988 WASHINGTON US, pages 8998-9002, M.A.FROHMAN ET AL. 'Rapid production of full-length cDNAs from rare transcripts: Amplification using a single gene-specific oligonucleotide primer' see page 8998 --- -/--	10,28



Further documents are listed in the continuation of box C.



Patent family members are listed in annex.

* Special categories of cited documents:

- *A* document defining the general state of the art which is not considered to be of particular relevance
- *E* earlier document but published on or after the international filing date
- *L* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- *O* document referring to an oral disclosure, use, exhibition or other means
- *P* document published prior to the international filing date but later than the priority date claimed

T later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

X document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

Y document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

& document member of the same patent family

Date of the actual completion of the international search

4 January 1996

Date of mailing of the international search report

16.01.96

Name and mailing address of the ISA

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 Fax (+31-70) 340-3016

Authorized officer

Cupido, M

INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 95/07628

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	VIROLOGY, v 1. 187, no. 2, April 1992 ORLANDO US, pages 620-626, S.PASTOREKOVA ET AL. 'A novel quasi-viral agent, MaTu, is a two-component system' see the whole document ---	1-28
P,X	ONCOGENE, vol. 9, no. 10, October 1994 pages 2877-2888, J.PASTOREK ET AL. 'Cloning and characterization of MN, a human tumor-associated protein with a domain homologous to carbonic anhydrase and a putative helix-loop-helix DNA binding segment' see the whole document -----	1-14, 16-18, 21-27

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US97/07628

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This international search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☒ Claims Nos.: 18,20,21,26
because they relate to subject matter not required to be searched by this Authority, namely:
Remark: Although claims 18,20,21 and 26 are directed to methods of treatment of, and diagnostic methods practised on the human body the search has been carried out and based on the alleged effects of the composition.
2. ☐ Claims Nos.:
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
- ☐ No protest accompanied the payment of additional search fees.

Information on patent family members

PCT/US 95/07628

Form PCT/ISA/210 (patent family annex) (July 1992)